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# Exogenous application of NaBiF<sub>4</sub> nanoparticle affects wheat root development

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## Abstract

**Background:** Nanoparticle causes soil pollution, which affected plant development and then resulted in biomass decreased, especially in crops. However, little is known how sodium nanoparticles affect wheat root development at plant physiological level.

**Results:** We used NaBiF<sub>4</sub> (size of 50–100 nm) to analyze the effect in wheat development at plant physiological level. Under exogenous application of 50 μM NaBiF<sub>4</sub> for treatment, wheat root elongation was inhibited, but fresh weight and dry weight were increased. We also found that NaBiF<sub>4</sub> induced that the plant had lower content of sodium than negative control. Used no-sodium nanoparticle of BiF<sub>3</sub> for another negative control, it was also supported that NaBiF<sub>4</sub> entered into cell to replace of sodium and exported sodium out of plant. These results implied NaBiF<sub>4</sub> might induce sodium export to maintain the balance between sodium and potassium elements. Additionally, metabolism analysis demonstrated that SOD activity was increased, but CAT and POD activity reduced under exogenous treatment of NaBiF<sub>4</sub> nanoparticles.

**Conclusions:** Sodium nanoparticles (NaBiF<sub>4</sub>) inhibited plant development by nanoparticle accumulation and sodium homeostasis broken, and then involved reactive oxygen species (ROS) signaling system response. These results provided more sights of sodium nanoparticle effect in plant development.

**Keywords:** Wheat, Root, Development, Nanoparticle, NaBiF<sub>4</sub>, Sodium, Homeostasis

## Background

In the past several decades, the world's population has been increased year by year. And cereal production similarly increased from 1.2 billion tons in 1969 to 2.8 billion tons in 2014 (FAOSTAT 1. data). Environmental factors play an essential role in crop plants development, such as temperature, light, drought, soil quality, nutrition, nanoparticles and so on. Environmental pollution, especially in soil, caused the crops production reduced due

to affect root activity and impeded substance transport activity.

Many nanoparticles contribute to their promising suitability for solar cells, drug delivery, temperature sensors, indoor illumination, and field emission displays. Once nanoparticle is taken in through root pathway, it resulted in beneficial or opposite effect in plant development. Until now, several nanoparticles have been reported on the interactions with the plants, including carbonaceous nanomaterials (fullerenes and nanotubes), metal oxides, zero-valent metals, nanopolymers, QDs and other NPs (Ni(OH)<sub>2</sub> and NaYF<sub>4</sub>) [1–8]. Actually, nanoparticles are different with nutrition, which are

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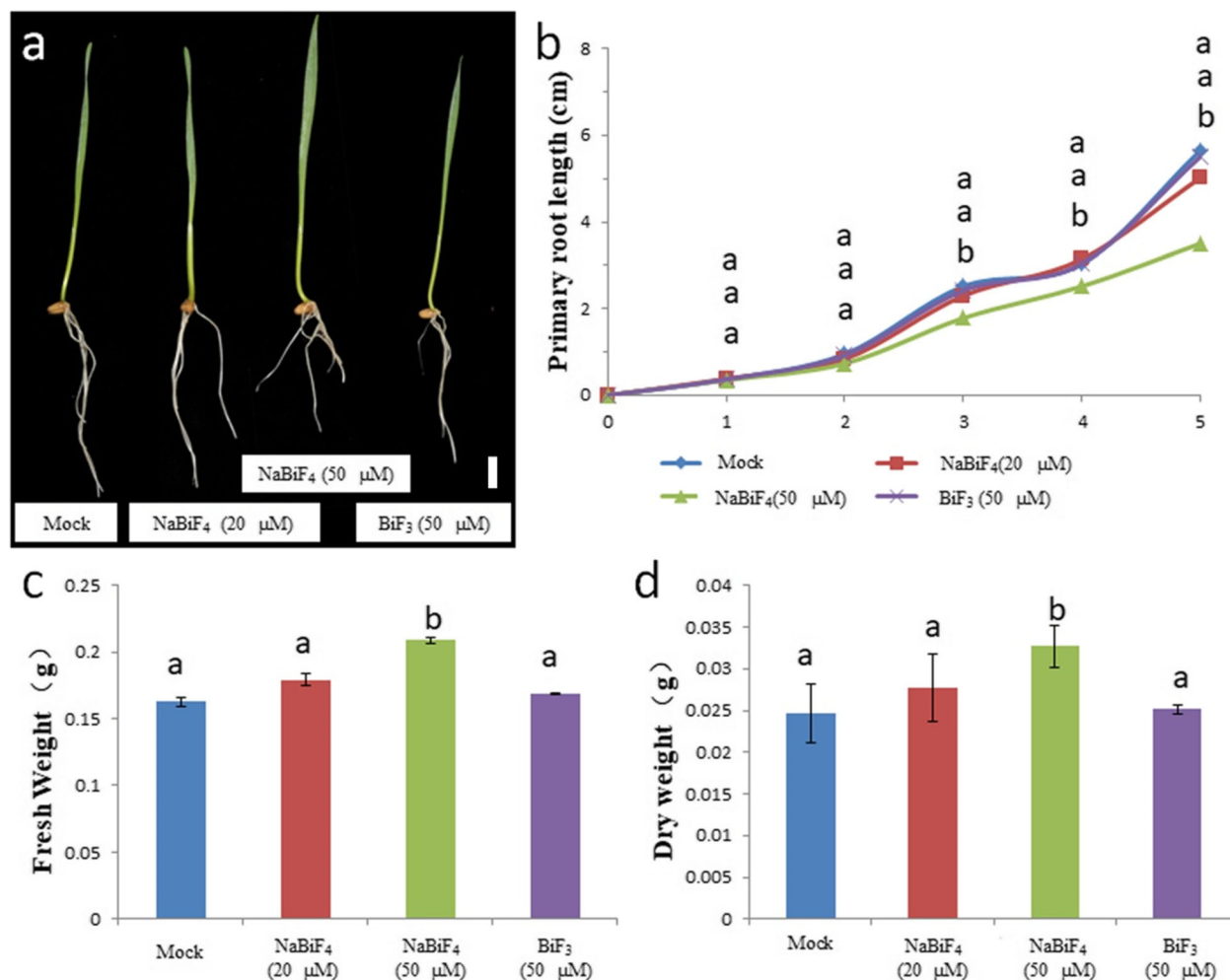
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assimilated by root as anion or cation type. Base on the nanoparticle physical characteristic of composition, size, concentration and coating of nanoparticle, it plays different roles. To some degree, high concentrations or low concentrations of nanoparticles have opposite functions in plant development, as inhibited or promoted plants, respectively. Nevertheless, magnetic  $\text{Fe}_3\text{O}_4$  even at the concentration of 2 mM does not cause serious injury in pumpkin (*Cucurbita maxima*) [9]. These positive effects of nanomaterials on plants were mainly reported for Au or Ag nanoparticles, Cu nanoparticles, Al related nanoparticles,  $\text{TiO}_2$  nanoparticles,  $\text{CeO}_2$  nanoparticles,  $\text{SiO}_2$  nanoparticles and carbonnanotubes [10–15].

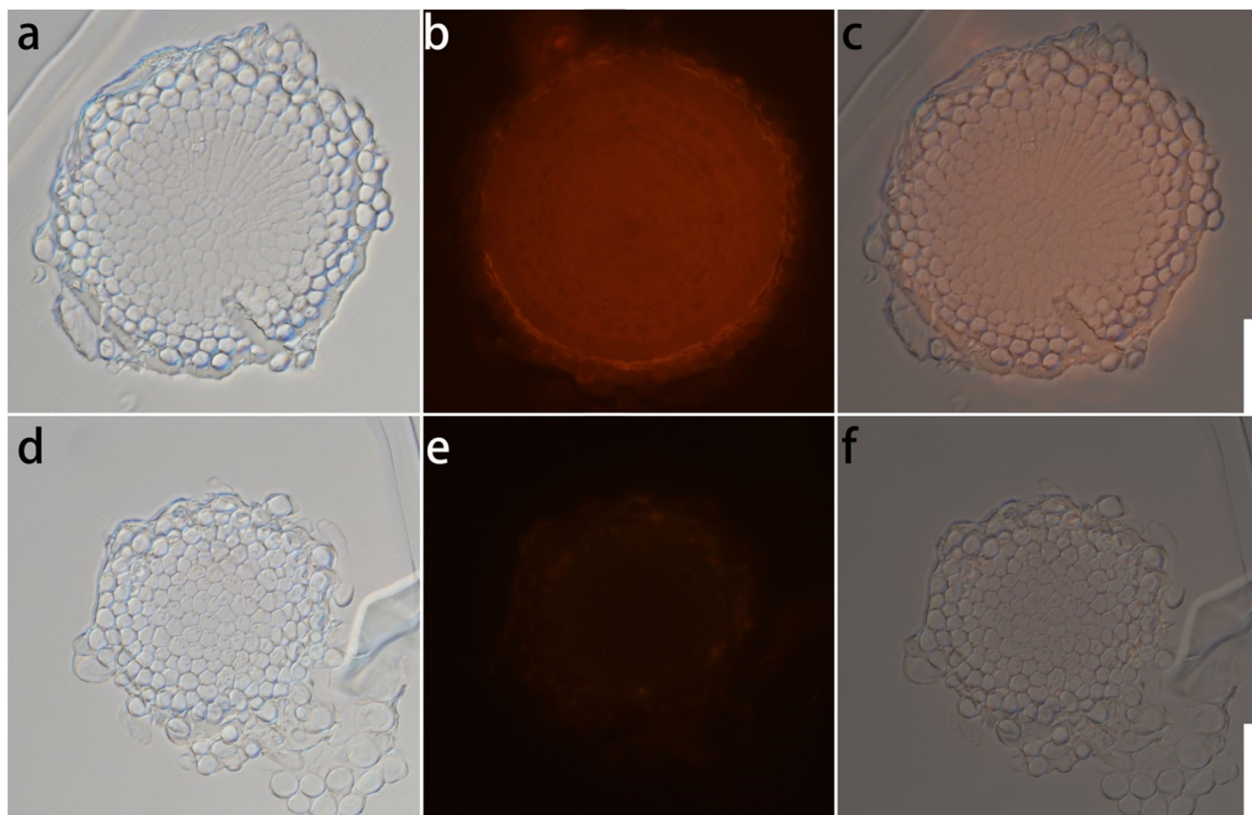
Always, most of high concentrations of the nanoparticles caused phytotoxicity by toxic ions, cell or tissue damage, production of excess ROS, catalytic reactions [16–20]. To detect nanoparticles in the plant tissues,

there are several different detection mechanisms of nanoparticles, such as fluorescence signaling, QDs, in situ analysis, nanoparticles color and so on [21]. Until now, little is known how nanoparticle affects crop plant development at metabolism level.

Wheat (*Triticum aestivum* L.) is one of the most important crop plants in the world, which supports the 1/3 of the food for human. Previously, it was reported that  $\text{TiO}_2$  nanoparticles with diameters ranging from 14 nm to 655 nm, were accumulated in wheat root. And  $\text{TiO}_2$  nanoparticles did not affect wheat seed germination, biomass and transpiration [22]. As the nanoparticles enter into plant cell, there are several different pathways for transport, such as: vascular system, membrane system, plasmodesmata system and so on. Base on the size of pathway in vascular, membrane, plasmodesmata or other system, we found nanoparticle size from 50 to 100 nm



**Fig. 1** Effect of  $\text{NaBiF}_4$  nanoparticle in wheat development at seedling stage. **a** Images of wheat plants grown in MSO medium with various concentrations of  $\text{NaBiF}_4$  nanoparticles at 10 DAG. Bar = 1 cm. **b** Primary root length as a function of DAG. **c** Fresh weight treated with different  $\text{NaBiF}_4$  nanoparticles at 10 DAG. **d** Dry weight treated with different  $\text{NaBiF}_4$  nanoparticles at 10 DAG. Error bars represent standard error for at least 5 samples. Values in the same column with different letters are significantly different ( $P < 0.05$ )



**Fig. 2** Cross section images of the wheat root tip treated with 50  $\mu\text{M}$   $\text{NaBiF}_4\text{:Eu}^{3+}$  nanoparticles at 4 DAG: **a** Bright channel, **b** RFP channel and **c** merged channel. Cross section images of the wheat root tip cultivated in MS0 medium without nanoparticles at 4 DAG: **d** Bright channel, **e** RFP channel and **f** merged channel. Bar = 50  $\mu\text{m}$

only depended on membrane. Previously, we used  $\text{NaBiF}_4$  and  $\text{BiF}_3$  for analysis the roles in rice root development [23, 24]. We found that  $\text{NaBiF}_4$  inhibited rice root elongation, but promoted more crown root formation. We analyzed several ROS signaling genes, which displayed transcript level of *OsOVPI*, *OsNIP2:1*, and *OsMT2* was reduced, but expression of *OsMT2b* increased [24]. Exogenous application of nanoparticle of  $\text{BiF}_3$  for treatment, which did not reduce rice root elongation, but not mediate *OsOVPI*, *OsNIP2:1*, *OsMT2*, and *OsMT2b* transcript level changed [23]. Because the composition of these two nanoparticles, only one element (sodium) shows difference, which might interrupt the native balance system, for example, homeostasis of sodium-potassium balance.

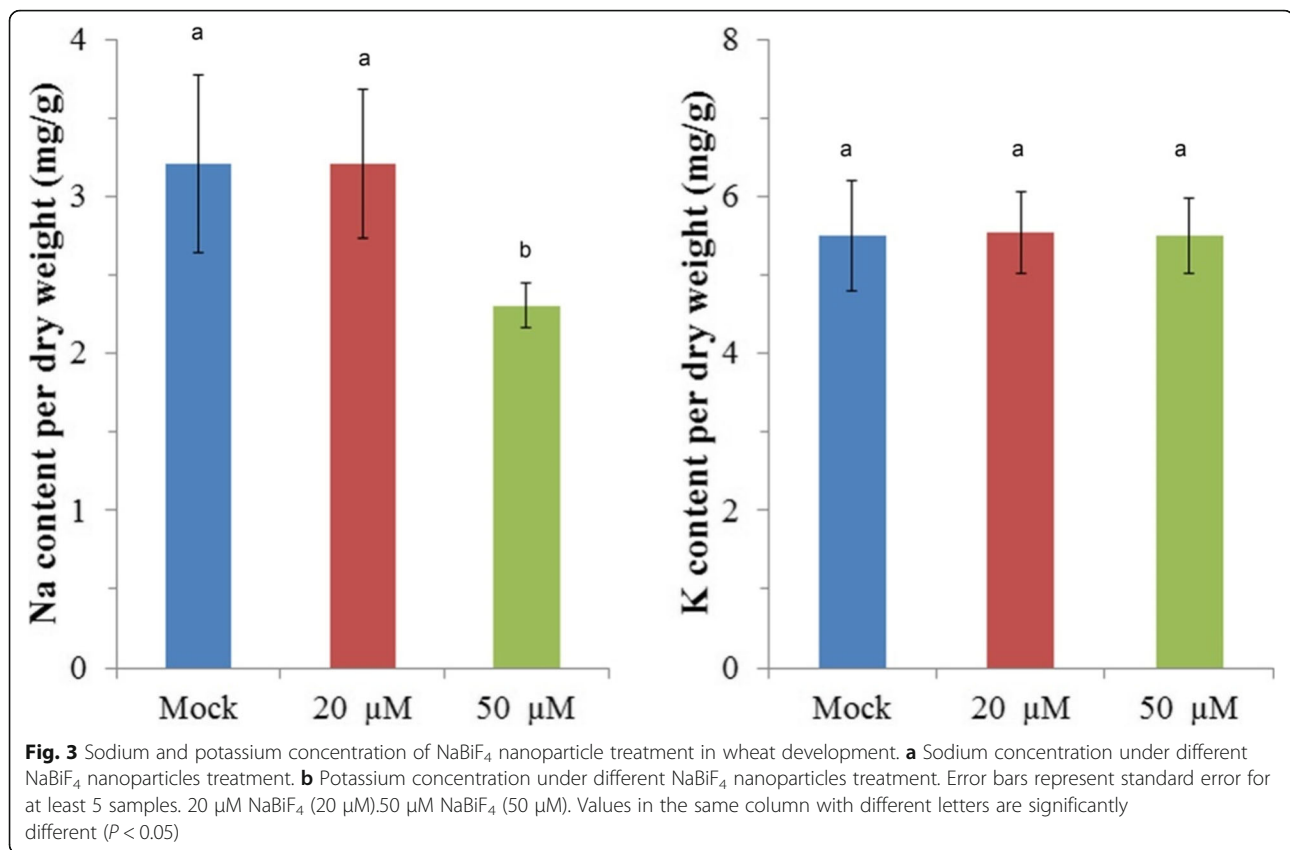
Plants generally maintain a stable  $\text{K}^+/\text{Na}^+$  ratio and a negative electrical membrane potential difference across the plasma membrane under a normal physiological state.  $\text{Na}^+$  enters into the roots through different channels and transporters [25]. However, if the balance was broken, plant may start ROS response reactions. In this study, we found that wheat root was much more sensitive to  $\text{NaBiF}_4$  nanoparticles than  $\text{BiF}_3$  nanoparticles in

root development, which caused the balance of sodium potassium pump affected.

## Results

### Effect of nanoparticles on the wheat root development

To analyze the effect of synthesized nanoparticles in wheat root development, wild type (WT) (*Triticum aestivum* L cultivar Yangmai 13) were grown in MS medium without sucrose (MS0), but with multiple concentrations of  $\text{NaBiF}_4$  nanoparticles. The images of the cultivated wheat were shown at 10 days after germination (DAG) in Fig. 1a. As demonstrated, the development of wheat root was significantly reduced by the 50  $\mu\text{M}$  concentration of nanoparticles. Clearly, compared with that of the wheat grown on MS0 medium without nanoparticles as a negative control (Mock), the elongation speed of primary roots was much slower for the seedlings treated with 50  $\mu\text{M}$  concentration of  $\text{NaBiF}_4$  nanoparticles (WT-HT) (Fig. 1b). And the length of WT-HT root reduced about 57.14%. Nevertheless, when the concentration of nanoparticles was declined to as low as 20  $\mu\text{M}$  (WT-LT), the length of the primary roots was not significantly changed compared with the Mock (Fig. 1a-b). When the



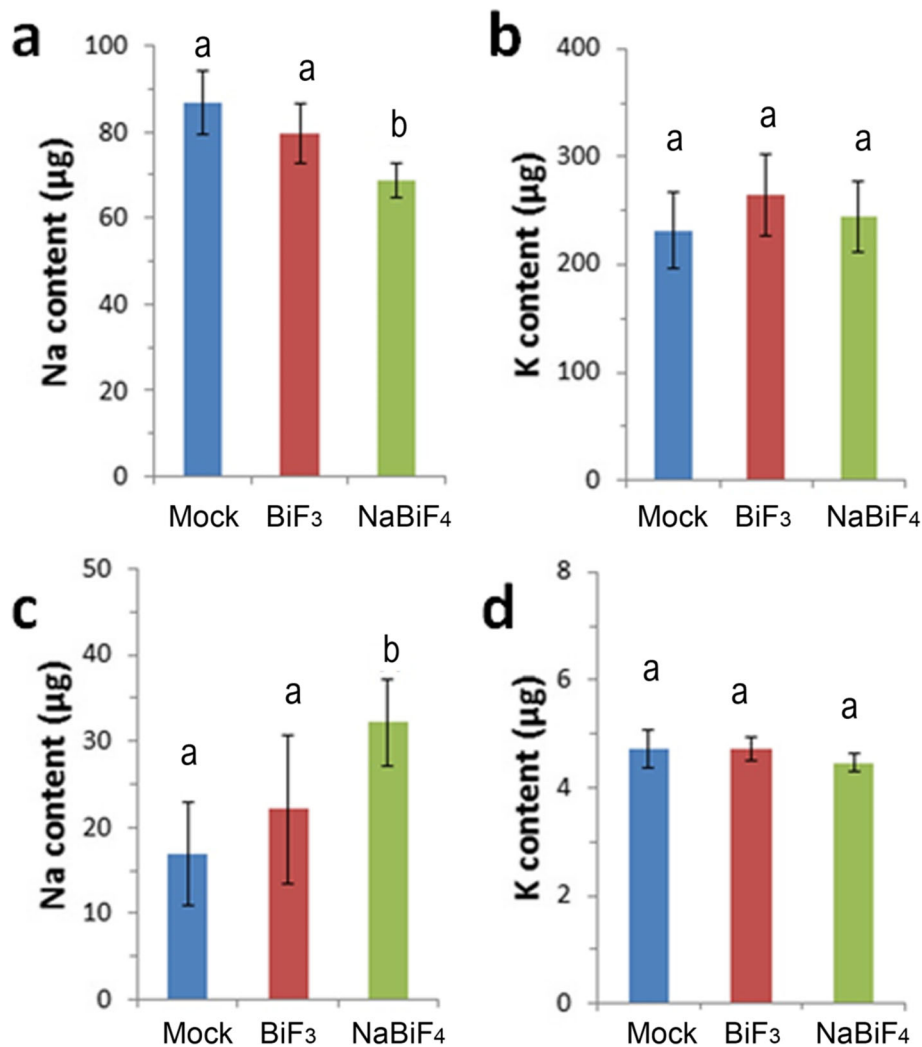
seedlings plants were treated with high concentration of NaBiF<sub>4</sub> nanoparticles, the fresh weight and dry weight were measured. Interestingly, although the primary root elongation was inhibited, the fresh weight and dry weight were increased up to 131.25 and 130%, respectively (Fig. 1c-d). Here, we also used BiF<sub>3</sub> nanoparticles as another controls, these data indicated that 50  $\mu$ M NaBiF<sub>4</sub> nanoparticles induced wheat biomass accumulation.

#### Nanoparticles caused sodium export from wheat seedling plant

Previously, we reported that NaBiF<sub>4</sub> nanoparticles caused rice root elongation inhibited due to phytotoxicity [24]. Eu acts as one type of earth element, which was visualized as red emission in the RFP channel. And, the NaBiF<sub>4</sub>:Eu<sup>3+</sup> nanoparticles not only emitted dazzling visible red emission under the NUV excitation but also exhibited similar characteristic as the NaBiF<sub>4</sub> nanoparticles in rice [24]. To get deep insight into the location of the nanoparticles, the cross section of root tip further confirmed that the NaBiF<sub>4</sub>:Eu<sup>3+</sup> nanoparticles were distributed in the cells (Fig. 2a-c). Similarly, the negative results did not have any obvious signals in the wheat root grown in the MS0 medium (Fig. 2d-f). These results demonstrated that nanoparticles were accumulated in root tip. These results were similar with in rice, as the previous reported (Du et al., 2018).

Multiple factors affect ROS signaling response by phytotoxicity, such as sodium stress, nutrition transport disrupt, and so on. To further understand the mechanism by nanoparticles treatment, we measured sodium concentration. We found wheat seedling by 50  $\mu$ M NaBiF<sub>4</sub> nanoparticles treatment had lower level of sodium (71.874%) than Mock, but 20  $\mu$ M NaBiF<sub>4</sub> nanoparticles treatment was not significant changed (Fig. 3a). Here, we used potassium content for negative control, which demonstrated that there were no obvious changed (Fig. 3b) in these three groups. It implied that NaBiF<sub>4</sub> nanoparticles entered into cell resulted in less sodium in cell. Meanwhile, NaBiF<sub>4</sub> nanoparticles induced sodium export from cell.

To further confirm this hypothesis, we used 50  $\mu$ M NaBiF<sub>4</sub> nanoparticles treatment for the similar experiments. And the solution used water to instead of MS0 medium in case sodium contamination from MS0 medium. With the treatment of NaBiF<sub>4</sub> nanoparticles, sodium concentration was decreased about 81.39% compare with negative control. Also, we measured the sodium content in left solutions, sodium under NaBiF<sub>4</sub> nanoparticles treatment had more than WT-CK (137.5%). And we also measured potassium concentration that there was no affected in Fig. 4b-d. This stated clearly that NaBiF<sub>4</sub> nanoparticle caused extra sodium export out of plant into solution.



**Fig. 4** Sodium and Potassium concentration of NaBiF<sub>4</sub> and BiF<sub>3</sub> nanoparticle treatment in wheat development. **a** Sodium concentration under different NaBiF<sub>4</sub> nanoparticles treatment in wheat. **b** Potassium concentration under different NaBiF<sub>4</sub> nanoparticles treatment wheat. **c** Sodium concentration under different NaBiF<sub>4</sub> nanoparticles treatment export from cell into water. **d** Potassium concentration under different NaBiF<sub>4</sub> nanoparticles treatment export from cell into water. Error bars represent standard error for at least 5 samples. Values in the same column with different letters are significantly different ( $P < 0.05$ )

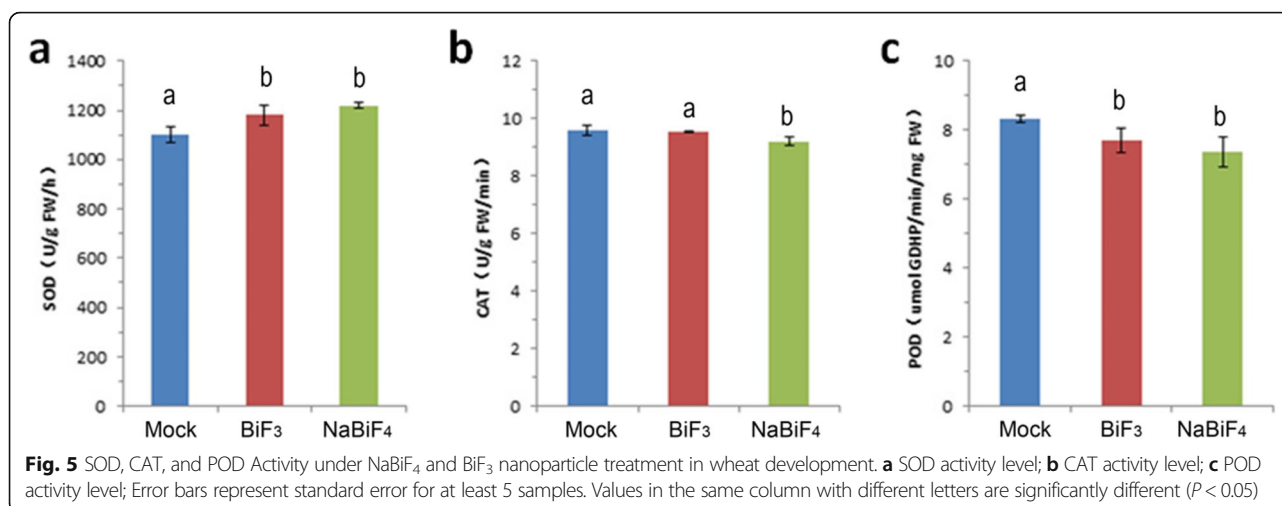
Additional, this phenotype might due to Bismuth (Bi) or Fluorine (F). we chose another nanoparticle BiF<sub>3</sub> for synchronization. Exogenous application of 50 µM nanoparticle BiF<sub>3</sub>, which does not have sodium, did not inhibit root elongation in rice (Du et al., 2018a), as well as in wheat (Fig. 1). Also, with 50 µM BiF<sub>3</sub> nanoparticles for treatment, sodium and potassium concentrations in plant were not affected in plant and export solutions (Fig. 4a-b). It further demonstrated that NaBiF<sub>4</sub> displaced the sodium in cell to maintain the balance of sodium and potassium.

#### ROS metabolism due to nanoparticles

As deduced above, less sodium and much NaBiF<sub>4</sub> nanoparticles entered into plant cells, which might affect cell

metabolism reaction (phytotoxicity). This reaction includes two parts: affect sodium content, and xenobiotic substance, which induced by the nanoparticles might be the main factor to affect the development of the wheat roots. To response the phytotoxicity, several ROS system metabolism could be response to the wheat root, such as the superoxide dismutase (SOD), catalase (CAT), and peroxidase (POD) [11]. To better comprehend the nanoparticles induced phytotoxicity in the wheat root, the activity level of SOD, CAT, POD to several phytotoxicity related metabolism were analyzed in Fig. 5a-c. Compared with Mock, the activity of the SOD was much higher in these wheat roots treated with the NaBiF<sub>4</sub> nanoparticles (50 µM), as well as treated with the BiF<sub>3</sub> nanoparticles (50 µM). Noted that, with the treatment of





the resultant nanoparticles, the activity level of the CAT and POD were reduced (Fig. 5b-c). Since the nanoparticles treated to the seedlings exhibited higher activity of SOD, and then lower activity of CAT and POD involved, it were expected to response to ROS system.

## Discussion

As the industry development, soil contaminated day by day due to heavy metal, salinization, nanoparticles accumulation. Previously, we used multiple concentrations of NaBiF<sub>4</sub> and BiF<sub>3</sub> for exogenous application to another crop plant (rice) for treatment. These results demonstrated that high content (100  $\mu$ M) of NaBiF<sub>4</sub> caused toxicity by the root length reduced and more crown root number. For the particles location, it is accumulated at division and elongation zone. Further phytotoxicity related genes, transcript level of *OsOVPI*, *OsNIP2;1*, and *OsMT2* was reduced and *OsMT2b* increased [24]. Similar content of BiF<sub>3</sub> exogenous treatment with NaBiF<sub>4</sub> to rice did not show any obvious phenotype, although BiF<sub>3</sub> also located at root tip, as NaBiF<sub>4</sub> [23]. It implied that NaBiF<sub>4</sub> and BiF<sub>3</sub> have significant and different roles in plant development. In this study, we reported that same unsoluble nanoparticles, NaBiF<sub>4</sub> and BiF<sub>3</sub>, which affected wheat development similar with in rice. Exogenous application of 50  $\mu$ M NaBiF<sub>4</sub> caused root length decreased, but BiF<sub>3</sub> not. Interestingly, higher activity of SOD and lower of POD by the treatments of NaBiF<sub>4</sub> and BiF<sub>3</sub> nanoparticles, reduced CAT activity by NaBiF<sub>4</sub>, which demonstrated that both NaBiF<sub>4</sub> and BiF<sub>3</sub> affected ROS response reaction by tissue or cell abnormal in wheat root. Previously, Wang reported that nanoparticles caused phytotoxicity might due to (i) the dissolution and release of toxic ions; (ii) size- or shape-dependent mechanical damage and clogging; (iii) the production of excess ROS; (iv) binding interactions caused surface reconstruction of biological molecular structures; (v) oxidation of biomolecules through catalytic

reactions [21]. Compare with BiF<sub>3</sub> nanoparticles, NaBiF<sub>4</sub> has one more element of sodium. We found less sodium concentration in plant than control, as well as used BiF<sub>3</sub> treatment for negative control. Meanwhile, the reduced the sodium exported from the tissue into the solutions. It means that NaBiF<sub>4</sub> play as sodium might cause sodium and potassium balance, BiF<sub>3</sub> acts as one type of the exogenous substance, which might due to tissue damage and pathway clogging [21]. These results, above, indicated that NaBiF<sub>4</sub> nanoparticles resulted in wheat root toxicity both in NaBiF<sub>4</sub> accumulation in root and sodium export out of plant, as depicted as Fig. 6a. And BiF<sub>3</sub> nanoparticles can also induce ROS signaling response only in BiF<sub>3</sub> accumulation in root (Fig. 6b).

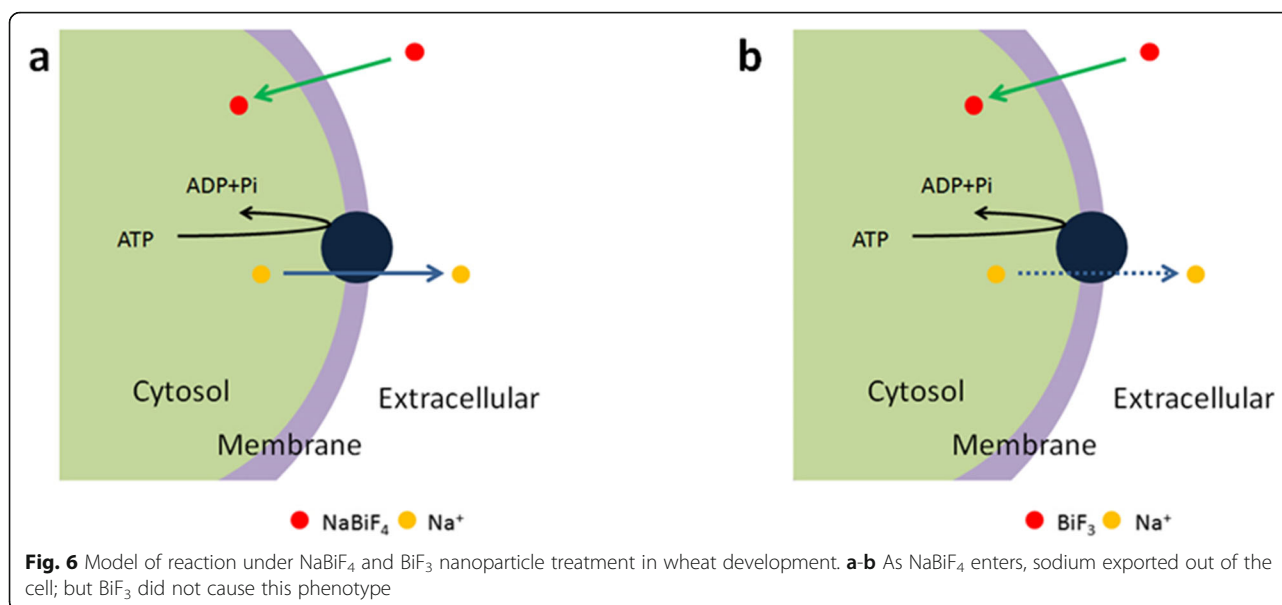
## Conclusion

Previously, we found that NaBiF<sub>4</sub> accumulated at rice root elongation zone, and then induced ROS system signaling response by several genes transcript level affected, such as, *OsOVPI*, *OsNIP2;1*, *OsMT2*, and *OsMT2b*. Here, we used another crop plant, wheat, to further analyze these phytotoxicity reactions from plant physiological level. As the root assimilated NaBiF<sub>4</sub> nanoparticle into cell, stable sodium from nanoparticle caused sodium export from root cell and then move into growth solution. Due to nanoparticle accumulation and less floating sodium level for plant physiological reaction, ROS related metabolism reactions were induced, which generated higher activity of SOD, and then lower activity of CAT, and POD. In the future, we will further analyze how nanoparticles move into cell.

## Methods

### Plant materials

The wheat cultivars selected in this study was Wheat (*Triticum aestivum* L. 'Ningmai13'), which were provided by the Lixiahe Agricultural Research Institute.



### Synthesis of NaBiF<sub>4</sub> and BiF<sub>3</sub> nanoparticles

High-purity powders of NaNO<sub>3</sub>, Bi(NO<sub>3</sub>)<sub>3</sub>·5H<sub>2</sub>O, and NH<sub>4</sub>F acted as the raw materials to prepare the nanoparticles [23]. To prepare the NaBiF<sub>4</sub> nanoparticles, two solutions were prepared. BiF<sub>3</sub>, NaBiF<sub>4</sub>, BiF<sub>3</sub>:Eu<sup>3+</sup> and NaBiF<sub>4</sub>:Eu<sup>3+</sup> were synthesized previously reported [23, 24].

### Determination of K<sup>+</sup> and Na<sup>+</sup> concentrations

The K<sup>+</sup> and Na<sup>+</sup> concentrations were measured as described previously [26–28].

### SOD, CAT, POD assay

The activities of SOD, CAT, and POD activity of wheat root was measured as described previously [29, 30]. 4 day after germination, the seedling wheat plants were moved to 50 μM NaBiF<sub>4</sub> and BiF<sub>3</sub> nanoparticles water solution for 3 days. About 100 mg of mixed material were harvested and ground in liquid nitrogen to a fine powder and then homogenized in 5 ml 10 mM PBS (pH 7.0) containing 1% PVP (w/v), 1 mM PMSF, 0.1% Triton-X100 (w/v) and 0.1 mM EDTA. The extraction was performed at 4 °C. After centrifugation at 12,000 g for 20 min, the supernatant solution was used as the preparation for individual enzyme activity. Then SOD and CAT activity were measured by spectrophotometer at 560 nm and 240 nm, respectively. The adrenochrome formation in the next 3 min was recorded at 470 nm in a UV–V is spectrophotometer.

### Statistical analysis

The experimental data was performed using t-test at a probability significance level of  $P < 0.05$  in SPSS.

### Abbreviations

ROS: Reactive oxygen species; SOD: Superoxide dismutase; CAT: Catalase; POD: Peroxidase; OsMT1: Metallothionein 1; OsMT2: Metallothionein 2; OsOVP1: Vacuolar H<sup>+</sup>-translocating inorganic pyrophosphatase; OsNIP2: 1: nodulin 26-like intrinsic protein; OsMT2b: Metallothionein2b

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### Authors' contributions

FX and YFW designed research; YFW, WMH P, QQJ performed the experiments; YFW wrote manuscript; FX, YFW, GC, QQJ, WMH P, ZDD, YRX corrected manuscript. All authors have read and approved the manuscript.

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### Availability of data and materials

All data generated or analyzed during this study are included in this manuscript.

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Competing interests

The authors declare that they have no competing interests.

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## References

- Wang B, Wang H, Chang J, Tso H, Chou Y. More spherical large fullerenes and multi-layer fullerene cages. *J Mol Struct Theochem*. 2001;540:171–6.
- Mauter MS, Elimelech M. environmental applications of carbon-based nanomaterials. *Environ Sci Technol*. 2008;42:5843–59.
- Wang P, Menzies NW, Lombi E, BA MK, Ohannessen B, Glover CJ, Kappen P, Kopittke PM. Fate of ZnO nanoparticles in soils and cowpea (*VignaUnguiculata*). *Environ Sci Technol*. 2013;47:13822–30.
- Zhai G, Walters KS, Peate DW, Alvarez PJ, Schnoor JL. Transport of gold nanoparticles through plasmodesmata and precipitation of gold ions in woody poplar. *Environ Sci Technol Lett*. 2014;1:146–51.
- Phenrat T, Cihan A, Kim H, Mital M, Illangasekare T, Lowry GV. Transport and deposition of polymer-modified FeO nanoparticles in 2-D heterogeneous porous media: effects of particle concentration, FeO content, and coatings. *Environ Sci Technol*. 2010;44:9086–93.
- Koo Y, Wang J, Zhang Q, Zhu H, Chehab EW, Colvin VL, Alvarez PJ, Braam J. fluorescence reports intact quantum dot uptake into roots and translocation to leaves of arabidopsis thaliana and subsequent ingestion by insect herbivores. *Environ Sci Technol*. 2014;49:626–32.
- Parsons JG, Lopez ML, Gonzalez CM, Peralta Videia JR, Gardea T, orresdey JL. Toxicity and biotransformation of uncoated and coated nickel hydroxide nanoparticles on mesquite plants. *Environ Toxicol Chem*. 2010;29:1146–54.
- Yin W, Zhou L, Ma Y, Tian G, Zhao J, Yan L, Zheng X, Zhang P, Yu J, Gu Z. Phytotoxicity, translocation, and biotransformation of NaYF<sub>4</sub> upconversion nanoparticles in a soybean plant. *Small*. 2015;11:4774–84.
- Zhu H, Han J, Xiao JQ, Jin Y. Uptake, translocation, and accumulation of manufactured iron oxide nanoparticles by pumpkin plants. *J Environ Monit*. 2008;10:713–7.
- Akiyama T, Nakada M, Terasaki N, Yamada S. Photocurrent enhancement in a porphyrin-gold nanoparticle nanostructure assisted by localized plasmon excitation. *Chem Commun*. 2006;4:395–7.
- Shah V, Belozero I. influence of metal nanoparticles on the soil microbial community and germination of lettuce seeds. *Water Air Soil Pollut*. 2009; 197:143–8.
- Su M, Liu H, Liu C, Qu C, Zheng L, Hong F. Promotion of nano-anatase TiO<sub>2</sub> on the spectral responses and photochemical activities of D1/D2/Cyt b559 complex of spinach. *Spectrochim Spectrochim Acta A*. 2009;72:1112–6.
- Wang X, Jia Y. Study on adsorption and remediation of heavy metals by poplar and larch in contaminated soil. *Environ Sci Pollut Res*. 2010;17:1331–8.
- Morales MI, Rico CM, Hernandez-Viezcas JA, Nunez JE, Barrios AC, Tafuya A, Flores-Marges JP, Peralta-Videa JR, Gardea-Torresdey JL. toxicity assessment of cerium oxide nanoparticles in cilantro (*Coriandrum sativum* L.) plants grown in organic soil. *J Agric Food Chem*. 2010;61:6224–30.
- Lin BS, Diao SH, Li CH, Fang LJ, Qiao SC, Yu M. Effect of TMS (Nanostructured Silicon Dioxide) on growth of changbai larch seedlings. *J For Res*. 2004;15:138–40.
- Asli S, Neumann PM. Colloidal suspensions of clay or titanium dioxide nanoparticles can inhibit leaf growth and transpiration via physical effects on root water transport. *Plant Cell Environ*. 2009;32:577–84.
- Nel AE, Mädler L, Velegol D, Xia T, Hoek EMV, Somasundaran P, Klaessig F, Castranova V, Thompson M. Understanding biophysicochemical interactions at the nano-bio interface. *Nat Mater*. 2009;8:543–57.
- Shen C, Zhang Q, Li J, Bi F, Yao N. Induction of programmed cell death in Arabidopsis and rice by single-wall carbon nanotubes. *Am J Bot*. 2010;97: 1602–9.
- Atha DH, Wang H, Petersen EJ, Cleveland D, Holbrook RD, Jaruga P, Dizdargolu M, Xing B, Nelson BC. Copper oxide nanoparticle mediated DNA damage in terrestrial plant models. *Environ Sci Technol*. 2012;46:1819–27.
- Zhao L, Peng B, Hernandez-Viezcas JA, Rico C, Sun Y, Peralta-Videa JR, Tang X, Niu G, Jin L, Varela-Ramirez A, Zhang J, Gardea-Torresdey JL. Stress response and tolerance of *zea mays* to CeO<sub>2</sub> nanoparticles: cross talk among H<sub>2</sub>O<sub>2</sub>, heat shock protein, and lipid peroxidation. *ACS Nano*. 2012;6: 9615–22.
- Wang P, Lombi E, Zhao F, Kopittke PM. Nanotechnology: A New Opportunity in Plant Sciences. *Trends Plant Sci*. 2016;21:699–712.
- Larue C, Castillo-Michel H, Sobanska S, Trcera N, Sorieul S, Cécillon L, Ouerdane L, Legros S, Sarret G. Fate of pristine TiO<sub>2</sub> nanoparticles and aged paint-containing TiO<sub>2</sub> nanoparticles in lettuce crop after foliar exposure. *J Hazard Mater*. 2014;273:17–26.
- Du P, Wu Y, Yu JS. Synthesis and luminescent properties of Eu<sup>3+</sup>-activated BiF<sub>3</sub> nanoparticles for optical thermometry and fluorescent imaging in rice root. *RSC Advances*. 2018a;8(12):6419–24.
- Du P, Wu Y, Yu JS. Real-time detection of the nanoparticle induced phytotoxicity in rice root tip through the visible red emissions of Eu<sup>3+</sup> ions. *Photochem Photobiol Sci*. 2018b;17:499.
- Keisham M, Mukherjee S, Bhatla SC. Mechanisms of Sodium Transport in Plants-Progresses and Challenges. *Int J Mol Sci*. 2018;19(3).
- Deng P, Jiang D, Dong Y, Shi X, Jing Wand Zhang W. Physiological characterisation and fine mapping of a salt-tolerant mutant in rice (*Oryza sativa*). *Funct Plant Biol*. 2015a;42:1026–35.
- Deng P, Shi X, Zhou J, Wang F, Dong Y, Jing W, Zhang W. Identification and fine mapping of a mutation conferring salt-sensitivity in rice (*Oryza sativa* L.). *Crop Sci*. 2015b;55:219–28.
- Zhou J, Wang F, Deng P, Jing W, Zhang W. Characterization and mapping of a salt-sensitive mutant in rice (*Oryza sativa* L.). *J Integr Plant Biol*. 2013;55: 504–13.
- Hasanuzzaman M, Nahar K, Alam MM, Fujita M. Modulation of antioxidant machinery and the methylglyoxal detoxification system in selenium-supplemented Brassica napus seedlings confers tolerance to high temperature. *Biol Trace Elem Res*. 2014;161:297–307.
- Zhao Q, Zhou L, Liu J, Cao Z, Du X, Huang F, Pan G, Cheng F. Involvement of CAT in the detoxification of HT-induced ROS burst in rice anther and its relation to pollen fertility. *Plant Cell Rep*. 2018;37(5):741–57.

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