

REVIEW

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# Effects of abiotic stress on plants: a systems biology perspective

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## Abstract

The natural environment for plants is composed of a complex set of abiotic stresses and biotic stresses. Plant responses to these stresses are equally complex. Systems biology approaches facilitate a multi-targeted approach by allowing one to identify regulatory hubs in complex networks. Systems biology takes the molecular parts (transcripts, proteins and metabolites) of an organism and attempts to fit them into functional networks or models designed to describe and predict the dynamic activities of that organism in different environments. In this review, research progress in plant responses to abiotic stresses is summarized from the physiological level to the molecular level. New insights obtained from the integration of omics datasets are highlighted. Gaps in our knowledge are identified, providing additional focus areas for crop improvement research in the future.

## Reviews

Recent advances in biotechnology have dramatically changed our capabilities for gene discovery and functional genomics. For the first time, we can now obtain a holistic “snapshot” of a cell with transcript, protein and metabolite profiling. Such a “systems biology” approach allows for a deeper understanding of physiologically complex processes and cellular function [1]. New models can be formed from the plethora of data collected and lead to new hypotheses generated from those models.

Understanding the function of genes is a major challenge of the post-genomic era. While many of the functions of individual parts are unknown, their function can sometimes be inferred through association with other known parts, providing a better understanding of the biological system as a whole. High throughput omics technologies are facilitating the identification of new genes and gene function. In addition, network reconstructions at the genome-scale are key to quantifying and characterizing the genotype to phenotype relationships [2].

In this review, we summarize recent progress on systematic analyses of plant responses to abiotic stress to include transcriptomics, metabolomics, proteomics, and

other integrated approaches. Due to space limitations, we try to emphasize important perspectives, especially from what systems biology and omics approaches have provided in recent research on environmental stresses.

## Plant responses to the environment are complex

Plants are complex organisms. It is difficult to find an estimate of the total number of cells in a plant. Estimates of the number of cells in the adaxial epidermal layer and palisade mesophyll of a simple *Arabidopsis* leaf are approximately 27,000 and 57,000 cells, respectively [3]. Another estimate of the adaxial side of the epidermal layer of the 7<sup>th</sup> leaf of *Arabidopsis* was close to 100,000 cells [4] per cm<sup>2</sup> of leaf area. An *Arabidopsis* plant can grow as large as 14 g fresh weight with a leaf area of 258 cm<sup>2</sup> (11 g fresh weight) [5]. Thus, we estimate that a single *Arabidopsis* plant could have approximately 100 million cells (range of 30 to 150 million cells assuming 2.4 to 11 million cells per g fresh weight). A one million Kg redwood tree could possibly have 70 trillion cells assuming a cell size 100 times larger than an *Arabidopsis* cell. Combine that with developmental changes, cell differentiation and interactions with the environment and it is easy to see that there are an infinite number of permutations to this complexity.

There is additional complexity within the cell with multiple organelles, interactions between nuclear, plastidial and mitochondrial genomes, and between cellular territories that behave like symplastically isolated

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domains that are able to exchange transcription factors controlling gene expression and developmental stages across the plasmodesmata. A typical plant cell has more than 30,000 genes and an unknown number of proteins, which can have more than 200 known post-translational modifications (PTMs). The molecular responses of cells (and plants) to their environment are extremely complex.

### Environmental limits to crop production

In 1982, Boyer indicated that environmental factors may limit crop production by as much as 70% [6]. A 2007 FAO report stated that only 3.5% of the global land area is not affected by some environmental constraint (see Table three point seven in <http://www.fao.org/docrep/010/a1075e/a1075e00.htm>). While it is difficult to get accurate estimates of the effects of abiotic stress on crop production (see different estimates in Table 1), it is evident that abiotic stress continues to have a significant impact on plants based upon the percentage of land area affected and the number of scientific publications directed at various abiotic stresses (Table 1). If anything

the environmental impacts are even more significant today; yields of the “big 5” food crops are expected to decline in many areas in the future due to the continued reduction of arable land, reduction of water resources and increased global warming trends and climate change [7].

This growing concern is reflected in the increasing number of publications focused on abiotic stresses. For example, since the pivotal review of systems biology by Kitano in 2002 [1], the number of papers published on abiotic stress in plants using a systems biology approach has increased exponentially (Figure 1).

### Multiple factors limit plant growth

Fundamentally, plants require energy (light), water, carbon and mineral nutrients for growth. Abiotic stress is defined as environmental conditions that reduce growth and yield below optimum levels. Plant responses to abiotic stresses are dynamic and complex [8,9]; they are both elastic (reversible) and plastic (irreversible).

The plant responses to stress are dependent on the tissue or organ affected by the stress. For example,

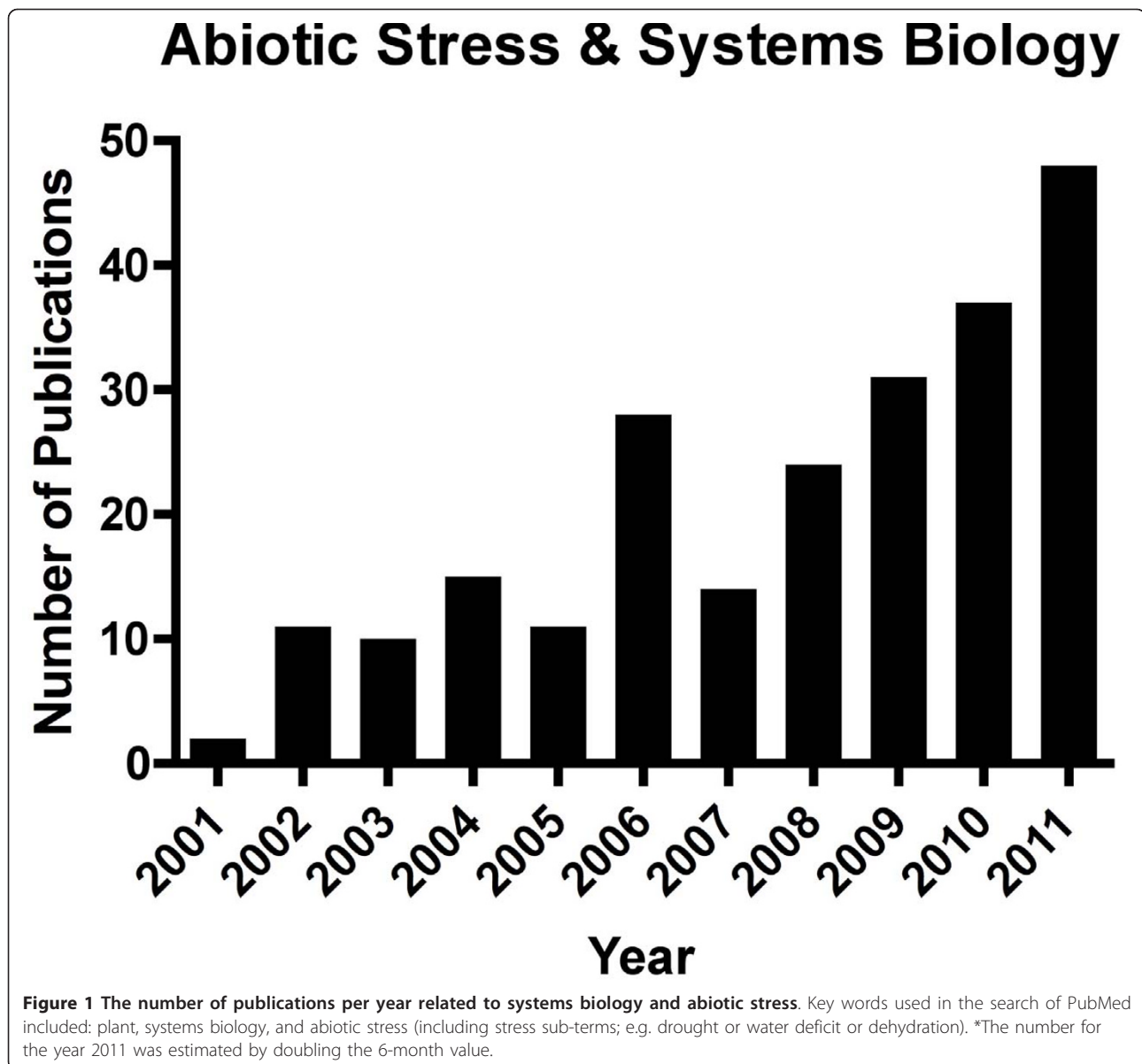
**Table 1 Estimates of the impacts of abiotic stresses on crop production and published research**

Stress Type	% of global land area affected*	% of global rural land area affected**	Number of Publications***
<b>Abiotic Stress</b>		96.5	<b>35,363</b>
<b>Water</b>			<b>4819</b>
Deficit or Drought	64	16	4137
Flooding or Anoxia	13	10	682
<b>Temperature</b>			<b>9715</b>
Cold	57	26	3798
Chilling			187
Freezing			350
High or heat			5380
<b>Light</b>			<b>7659</b>
Low			3081
High			4578
<b>Chemical/Soil</b>		50	<b>12391</b>
Salt or salinity	6	6	3498
Mineral deficiency or low fertility	9	39	222
Mineral toxicity			437
Acid soil	15		3646
Air pollutants			
Ozone			1369
Sulfur dioxide			378
NO <sub>x</sub> oxide			2001
Elevated CO <sub>2</sub>			840
Miscellaneous (e.g. wind, mechanical, etc.)			<b>779</b>

\*based on FAO World Soil Resources Report 2000 <ftp://ftp.fao.org/agl/agll/docs/wsr.pdf>.

\*\* based on Tables three point six and three point seven of 2007 FAO Report <http://www.fao.org/docrep/010/a1075e/a1075e00.htm>

\*\*\* data based on simple searches in PubMed between 2001 and July 7, 2011.



transcriptional responses to stress are tissue or cell specific in roots and are quite different depending on the stress involved [10]. In addition, the level and duration of stress (acute vs chronic) can have a significant effect on the complexity of the response [11,12].

Water deficit inhibits plant growth by reducing water uptake into the expanding cells, and alters enzymatically the rheological properties of the cell wall; for example, by the activity of ROS (reactive oxygen species) on cell wall enzymes [8]. In addition, water deficit alters the cell wall nonenzymatically; for example, by the interaction of pectate and calcium [13]. Furthermore, water conductance to the expanding cells is affected by aquaporin activity and xylem embolism [14-17]. The initial growth

inhibition by water deficit occurs prior to any inhibition of photosynthesis or respiration [18,19].

The growth limitation is in part due to the fundamental nature of newly divided cells encasing the xylem in the growing zone [20,21]. These cells act as a resistance to water flow to the expanding cells in the epidermis making it necessary for the plant to develop a larger water potential gradient. Growth is limited by the plant's ability to osmotically adjust or conduct water. The epidermal cells can increase the water potential gradient by osmotic adjustment, which may be largely supplied by solutes from the phloem. Such solutes are supplied by photosynthesis that is also supplying energy for growth and other metabolic functions in the plant. With long-

term stress, photosynthesis declines due to stomatal limitations for CO<sub>2</sub> uptake and increased photoinhibition from difficulties in dissipating excess light energy [12].

One of the earliest metabolic responses to abiotic stresses and the inhibition of growth is the inhibition of protein synthesis [22-25] and an increase in protein folding and processing [26]. Energy metabolism is affected as the stress becomes more severe (e.g. sugars, lipids and photosynthesis) [12,27,28]. Thus, there are gradual and complex changes in metabolism in response to stress.

### Central regulators limit key plant processes

The plant molecular responses to abiotic stresses involve interactions and crosstalk with many molecular pathways [29]. Systems biology and omics approaches have been used to elucidate some of the key regulatory pathways in plant responses to abiotic stress.

One of the earliest signals in many abiotic stresses involve ROS and reactive nitrogen species (RNS), which modify enzyme activity and gene regulation [30-32]. ROS signaling in response to abiotic stresses and its interactions with hormones has been thoroughly reviewed [32]. ROS and RNS form a coordinated network that regulates many plant responses to the environment; there are a large number of studies on the oxidative effects of ROS on plant responses to abiotic stress, but only a few studies documenting the nitrosative effects of RNS [30].

Hormones are also important regulators of plant responses to abiotic stress (Figure 2). The two most important are abscisic acid (ABA) and ethylene [33]. ABA is a central regulator of many plant responses to environmental stresses, particularly osmotic stresses [9,34-36]. Its signaling can be very fast without involving transcriptional activity; a good example is the control of stomatal aperture by ABA through the biochemical regulation of ion and water transport processes [35]. There are slower responses to ABA involving transcriptional responses that regulate growth, germination and protective mechanisms.

Recently, the essential components of ABA signaling have been identified, and their mode of action was clarified [37]. The current model of ABA signaling includes three core components, receptors (PYR/PYL/RCAR), protein phosphatases (PP2C) and protein kinases (SnRK2/OST1) [38,39]. The PYR/PYL/RCAR proteins were identified as soluble ABA receptors by two independent groups [38,39]. The 2C-type protein phosphatases (PP2C) including ABI1 and ABI2, were first identified from the ABA-insensitive Arabidopsis mutants *abi1-1* and *abi2-1*, and they act as global negative regulators of ABA signaling [40]. SNF1-related protein kinase 2 (SnRK2) is a family of protein kinases isolated

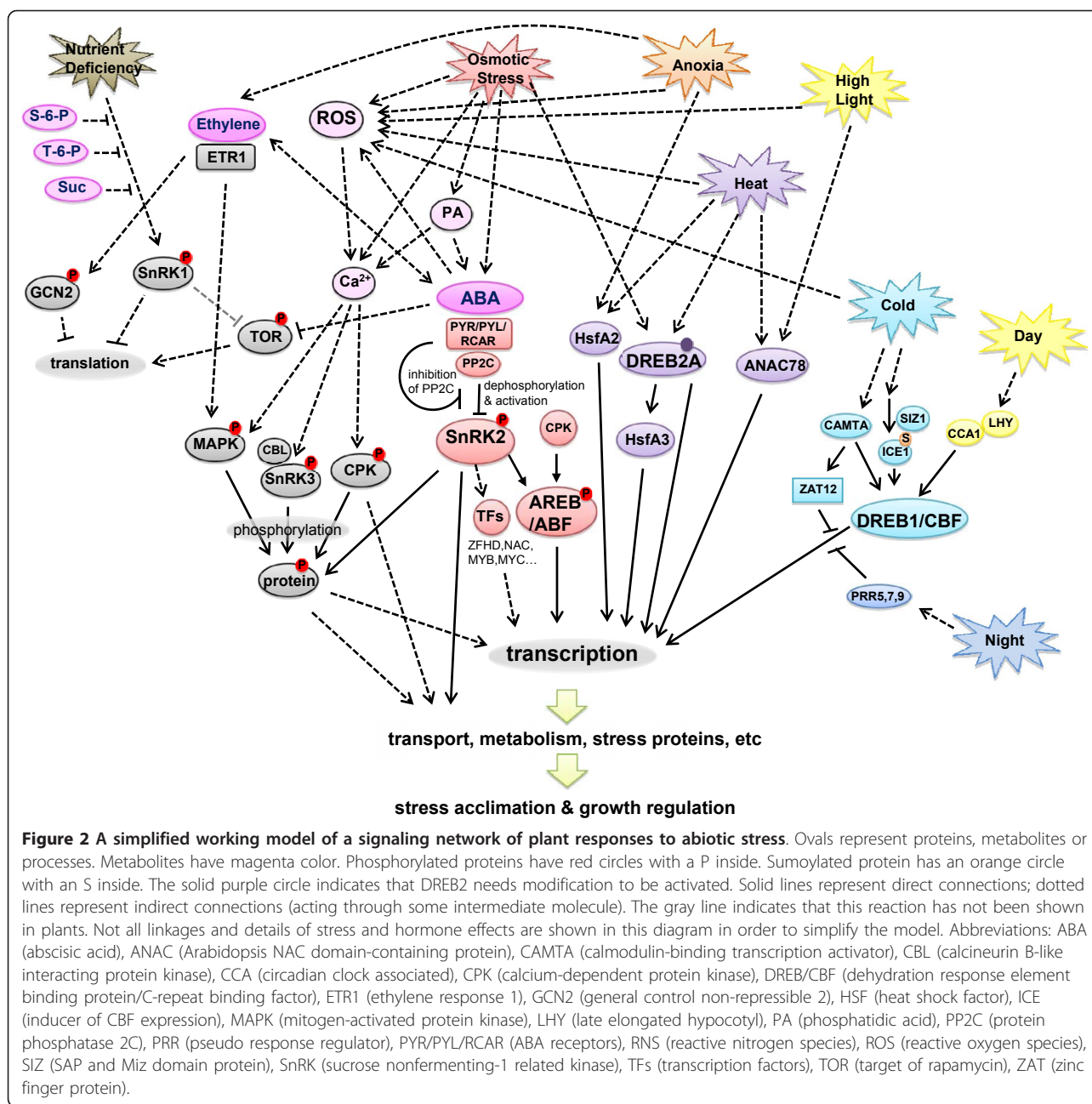
as ABA-activated protein kinases [41,42]. In Arabidopsis, three members of this family, SRK2D/SnRK2.2, SRK2E/OST1/SnRK2.6, and SRK2I/SnRK2.3, regulate ABA signaling positively and globally, as shown in the triple knockout mutant *srk2d srk2e srk2i (srk2dei)/snrk2.2 snrk2.3 snrk2.6*, which lacks ABA responses [43]. The PYR/PYL/RCAR - PP2C - SnRK2 complex plays a key role in ABA perception and signaling.

Studies of the transcriptional regulation of dehydration and salinity stresses have revealed both ABA-dependent and ABA-independent pathways [44]. Cellular dehydration under water limited conditions induces an increase in endogenous ABA levels that trigger downstream target genes encoding signaling factors, transcription factors, metabolic enzymes, and others [44]. In the vegetative stage, expression of ABA-responsive genes is mainly regulated by bZIP transcription factors (TFs) known as AREB/ABFs, which act in an ABA-responsive-element (ABRE) dependent manner [45-47]. Activation of ABA signaling cascades result in enhanced plant tolerance to dehydration stress. In contrast, a dehydration-responsive cis-acting element, DRE/CRT sequence and its DNA binding ERF/AP2-type TFs, DREB1/CBF and DREB2A, are related to the ABA-independent dehydration and temperature responsive pathways [44]. DREB1/CBFs function in cold-responsive gene expression [48,49], whereas DREB2s are involved in dehydration-responsive and heat-responsive gene expression [50].

Ethylene is also involved in many stress responses [51-53], including drought, ozone, flooding (hypoxia and anoxia), heat, chilling, wounding and UV-B light [31,33,53]. Ethylene signaling is well defined [51,52], and will not be discussed in detail here. There are known interactions between ethylene and ABA during drought [31], fruit ripening [54,55], and bud dormancy [56]. All of these interactions make the plant response to stress very complex [12,31,52].

In yeast, the well-documented central regulators of protein synthesis and energy are SnRK1 (Snf1/AMPK), TOR1 and GCN2 [57-60]. These proteins are largely controlled by the phosphorylation of enzymes; all three are protein kinases acting as key hubs in the coordination of metabolism during stressful conditions [61]. In plants, TOR activity is inhibited by osmotic stress and ABA [62] and GCN2 activity is stimulated by UV-light, amino acid starvation, ethylene, and cold stress [63]. SnRK1 responds to energy depletion, such as low light, nutrient deprivation or hypoxic conditions [64,65], and interacts with both glucose and ABA signaling pathways [66]. One of the results of this coordinated response is the inhibition of protein synthesis.

Many abiotic stresses directly or indirectly affect the synthesis, concentration, metabolism, transport and storage of sugars. Soluble sugars act as potential



signals interacting with light, nitrogen and abiotic stress [67-69] to regulate plant growth and development; at least 10% of *Arabidopsis* genes are sugar-responsive [68]. Mutant analysis has revealed that sugar signaling interacts with ethylene [70], ABA [71,72], cytokinins [73], and light [74,75]. In grapevine, sugar and ABA signaling pathways interact to control sugar transport. An ASR (ABA, stress-, and ripening-induced) protein isolated from grape berries is upregulated synergistically by ABA and sugars, and upregulates the expression of a hexose transporter [76]. VVSK1, a GSK3 type protein kinase, is also induced by

sugars and ABA, and upregulates the expression of several hexose transporters [77].

Stresses such as sugar starvation and lack of light stimulate SnRK1 activity ([64]. Suc-P synthase (SPS), 3-hydroxy-3-methylglutaryl-CoA reductase, nitrate reductase, and trehalose-6-P synthase are negatively regulated by SnRK1 phosphorylation [78], indicating that SnRK1 modulates metabolism by phosphorylating key metabolic enzymes. Post-translational redox modulation of ADPG-pyrophosphorylase, a key control of starch synthesis, by SnRK1 provides an interesting example of interactions between phosphorylation, redox control and sugar



metabolism [79]. In *Arabidopsis*, SnRK1 kinase activity is itself increased by GRIK1 and GRIK2, which phosphorylate a threonine residue of the SnRK1 catalytic subunit [78]. SnRK2 interacts with ABA for the control of stomatal aperture and participates in the regulation of plant primary metabolism. Constitutive expression of SnRK2.6 drastically boosts sucrose and total soluble sugar levels in leaves, presumably by controlling SPS expression [80].

### Systems biology approach to abiotic stress

In the post-genomic era, comprehensive analyses using three systematic approaches or omics have increased our understanding of the complex molecular regulatory networks associated with stress adaptation and tolerance. The first one is 'transcriptomics' for the analysis of coding and noncoding RNAs, and their expression profiles. The second one is 'metabolomics' that is a powerful tool to analyze a large number of metabolites. The third one is 'proteomics' in which protein and protein modification profiles offer an unprecedented understanding of regulatory networks. Protein complexes involved in signaling have been analyzed by a proteomics approach [81,82]. Integration of the different omics analyses facilitates abiotic stress signaling studies allowing for more robust identifications of molecular targets for future biotechnological applications in crops and trees.

### Co-expression analyses identify regulatory hubs

An important application of transcriptomics data is co-expression analysis of target genes using on-line analytical tools, such as ATTED-II (reviewed by [83]). This approach is very promising for understanding gene-gene correlations and finding master genes in target conditions.

In a series of pioneering papers, Hirai et al. [84,85] identified MYB transcription factors regulating glucosinolate biosynthesis in *Arabidopsis* in response to S and N deficiency using an integrated transcriptomics and metabolomics approach. Genes and metabolites in glucosinolate metabolism were found to be coordinately regulated [84]. Co-expression analysis was used to identify two MYB transcription factors that positively regulate glucosinolate metabolism [85]. Then a knock out mutant and ectopic expression of one of the transcription factors was used to validate its positive role in glucosinolate metabolism. Previously unidentified genes were assigned to this biosynthetic pathway and a regulatory network model was constructed [85].

Mao et al. [86] performed a gene co-expression network analysis of 1094 microarrays of *Arabidopsis* using a non-targeted approach. They identified 382 modules in this network. The top three modules with the most

nodes were: photosynthesis, response to oxidative stress and protein synthesis. Many of the modules also involved responses to environmental stresses. They constructed a cold-induced gene network from a subset of microarrays. The response to auxin stimulus was the most over-represented of the 18 significant modules.

Carrera et al. [87] used the InferGene application to construct a regulatory model of the *Arabidopsis* genome. They used datasets from 1,486 microarray experiments. Ten genes were predicted to be the most central regulatory hubs influencing the largest number of genes. Included in this set were transcription factor genes involved in auxin (KAN3), gibberellin (MYB29), abscisic acid (MYB121), ethylene (ERF1), and stress responses (ANAC036). They computed the top 12 gene subnetworks; four of these were related to biotic and abiotic stresses. Eighty-five percent of the predicted interactions of the 25% most connected transcription factors were validated in AtRegNet, the *Arabidopsis thaliana* Regulator Network <http://arabidopsis.med.ohio-state.edu/more-Network.html>.

Lorenz et al [88] investigated the drought response of loblolly pine roots and identified a number of hubs in the transcriptional network. Highly ranked hubs included thioredoxin, an inositol transporter, cardiolipin synthase/phosphatidyl transferase, 9-cis-epoxycarotenoid dioxygenase, zeatin O-glucosyltransferase and a SnRK2 kinase. These genes are involved in phospholipid metabolism, ABA biosynthesis and signaling, and cytokinin metabolism; they appear to be important in stress mediation.

Weston et al [89] used weighted co-expression analysis to define six modules for *Arabidopsis* responses to abiotic stress. Two hubs in the common response module were an ankyrin-repeat protein and genes involved in Ca signaling. They created a compendium of genomic signatures and linked them to their co-expression analysis. Using the same approach, they extended their analyses to the responses of three different plant species to heat and light [90]. Species-specific responses were found involving heat tolerance, heat-shock proteins, ROS, oligosaccharide metabolism and photosynthesis.

### Time-series analyses reveal multiple phases in stress responses

Time-series analyses allow one to distinguish between primary and secondary responses to stress. In a comprehensive time-series transcriptomics analysis of 7 abiotic stresses on different *Arabidopsis* organs [28], a core set of genes (50% were transcription factors) of non-specific responses for all stresses were elucidated. Included in this set were the AZF2, ZAT10 and ZAT12 transcription factors. This initial response is thought to be involved in the readjustment of energy homeostasis in

response to the stress. With time (after 1 h) more stress-specific profiles developed.

Sun et al [91] applied a complexity metric to a set of time series data of Arabidopsis with 9 different abiotic stresses. They found that genes with a higher complexity metric had longer 5' intergenic regions and a greater density of cis-regulatory motifs than the genes with a low complexity metric. Many of the cis-regulatory motifs identified were associated with previously characterized stress responses.

Vanderauwera et al. [92] investigated the effects of hydrogen peroxide ( $H_2O_2$ ) signaling during high light stress using microarray analyses. They found that  $H_2O_2$  was not only heavily involved in signaling in high light stress, but also salinity, water deficit, heat and cold stress.  $H_2O_2$  was a key regulator of small and 70 kD heat shock proteins and many genes of the anthocyanin metabolic pathway. Anthocyanins appear to play an important role as antioxidants in plants. A specific UDP-glycosyltransferase (UGT74E2) was highly regulated by  $H_2O_2$ . In a subsequent study [93], UGT74E2 responded quickly to  $H_2O_2$  and glycosylated indole-3-butyric acid (IBA) modifying auxin homeostasis, plant morphology and improving stress tolerance to salinity and water deficit. Furthermore, auxin was found to interact with ABA, increasing the ABA sensitivity of the plant. Silencing a poly(ADP-ribose) polymerase improved high light stress tolerance in Arabidopsis [94,95]. Part of the improved abiotic stress tolerance was ascribed to improved energy-use efficiency and reduced oxidative stress [94,95].

Kusano et al. [96] conducted a time-series experiment on the effects of UV-B light on Arabidopsis using both metabolomics and transcriptomics analyses. They found that plants responded in two phases with an upregulation of primary metabolites in the first phase and the induction of protective secondary metabolites, especially phenolics, in the second phase. The induction of phenolics corresponded to transcripts involved in the phenylpropanoid pathway, but the transcripts for primary metabolism were less consistent indicating that this pathway may be regulated by other mechanisms (e.g. kinases).

The transcriptomic response to drought can vary with the time of day [97]. These responses seem to interact with hormonal and other stress pathways that naturally vary during the course of the day. A smaller set of core genes were identified that responded at all times of the day. This set was compared to two previous studies and was whittled down to just 19 genes, including a NF-YB transcription factor, several PP2Cs, a CIPK7, and a sulfate transporter.

Drought stress studies and microarray analyses of three different genotypes of poplar clones grown in two

different locations revealed epigenetic regulation to the environment [98]. The tree clones that had a longer history in the environment showed greater changes in DNA methylation, thereby influencing their response to drought.

Shoot tip growth of grapevines was found to be much more sensitive to osmotic stress than gene expression in a time-series experiment of the effects of gradual osmotic stress on grapevine [27]. Proteomics data indicated that changes in protein expression preceded and were not well correlated with gene expression (G.R. Cramer, unpublished results). The integration of transcriptomics data and metabolomics data indicated distinct differences of the responses of salinity and an isosmotic water deficit [27]. Drought-stressed plants induced greater responses in processes needed for osmotic adjustment and protection against ROS and photoinhibition. Salinity induced greater responses in processes involved in energy metabolism, ion transport, protein synthesis and protein fate. A comparison to similar short-term stresses [11] indicated that a gradual, chronic stress response was more complex than an acute stress response.

The effect of water-deficit on Cabernet Sauvignon berries (a red wine grape) in the field was studied using transcriptomics, proteomics and metabolomics [99-102]. Integrated analyses confirmed that the phenylpropanoid pathway (including anthocyanin and stilbene biosynthesis) was upregulated by water deficit in a tissue-specific manner in the skins of the berries. Other metabolic pathways in the berries were affected by water deficit including ABA, amino acid, carotenoid, lipid, sugar and acid metabolism. Most of these changes were associated with improved quality characteristics of the fruit.

Likewise, Zamboni et al. [103] investigated berry development and withering in grapevine at the transcriptomics, proteomics and metabolomics levels. A multistep hypothesis-free approach from four developmental stages and three withering intervals, with integration achieved using a hierarchical clustering strategy (multivariate O2PLS technique), identified stage-specific functional networks of linked transcripts, proteins and metabolites, providing important insights into the key molecular processes that determine wine quality. A hypothesis-driven approach identified transcript, protein and metabolite variables involved in the molecular events underpinning withering, which predominantly reflected a general stress response. Berry ripening and withering are characterized by the accumulation of secondary metabolites such as acylated anthocyanins, but withering also involves the activation of osmotic and oxidative stress response genes and the production of stilbenes and taxifolin.

Usadel et al. [104] investigated the effects of cold temperatures over time using transcriptomics, metabolomics

and enzyme activities. They found some enzyme activities and metabolites changed rapidly, whereas others changed more slowly. The early changes (6 h) in enzyme activities were poorly correlated with transcript abundance, but after 78 h these correlations were greatly improved. Much of the long-term changes in metabolism could be ascribed to the CBF regulon.

Caldana et al. [105] conducted a complex time-series experiment (22 time points) with differing temperatures and light intensities using both metabolomics and transcriptomics analyses. This high-resolution time series experiment revealed that metabolic activities respond more quickly than transcriptional activities, indicating a disconnect between metabolism and transcription in the early phases of stress response and indicating that enzymatic activities may play a significant role. There were common metabolic responses to the changing environment within 1 h of the change including a decrease in energy metabolism and translation and an increase in the transcription of genes involved in signaling cascades. At later time points, condition-dependent metabolism was revealed. For example, protein degradation and energy metabolism derived from amino acids occurred in warm temperatures and darkness. Amino acid catabolism appears to fuel the TCA cycle in the absence of photosynthesis.

Yun et al. [106] characterized the response of rice to a mild chilling stress (10°C). They found that transcriptional regulation consisted of three dynamic and complex phases over 96 h. The early transcriptional phase appeared to be triggered by oxidative signals (H<sub>2</sub>O<sub>2</sub>) and lead to the subsequent induction of cellular defense and rescue mechanisms. Combining temporal co-expression data from microarrays with promoter motif enrichment analyses and oxidative responses, transcriptional regulatory network models for the different response phases were constructed. A bZIP-TGA transcription factor module (as1/ocs/TGA), one of seven transcription factor modules, was the most connected regulatory module in phase one. Each of the transcription factor modules consisted of clusters of transcription factors exhibiting combinatorial control of the chilling regulon. The speed of the response of this network was associated with chilling tolerance. Chilling-resistant genotypes had a much more rapid and pronounced response of this transcriptional regulatory network than chilling-sensitive genotypes. In addition, the transcription factors identified in this study were located within known growth and stress QTLs in the rice genome.

### **Integration of omics analysis identifies molecular networks functioning in abiotic stress responses**

Integrated omics analyses have markedly increased our understanding of plant responses to various stresses.

These analyses are important for comprehensive analyses of abiotic stress responses, especially the final steps of stress signal transduction pathways.

Integrated analyses of the transcriptome and the metabolome successfully demonstrate connections between genes and metabolites, elucidating a wide range of signal output from ABA under dehydration [107] and the DREB1/CBF transcription factors in response to low temperature [108,109]. Metabolite profiling reveals that ABA accumulates during dehydration, regulating the accumulation of various amino acids and sugars such as glucose and fructose. In particular, the dehydration-inducible accumulation of BCAAs (branch-chain amino acids), saccharopine, proline, and agmatine are correlated with the dehydration-inducible expression of their key biosynthetic genes (*BCAT2*, *LKR/SDH*, *P5CS1*, and *ADC2*, respectively), which are regulated by endogenous ABA [107]. In addition, metabolome analysis of transgenic *Arabidopsis* overexpressing DREB1A/CBF3 reveals that there is a striking similarity between the low-temperature regulated metabolome (monosaccharides, disaccharides, oligosaccharides and sugar alcohols) and that regulated by the DREB1A/CBF3 transcription factor [108,109]. In particular, the low-temperature-inducible accumulation of galactinol and raffinose is correlated with the expression of the *Gols3* gene, which is a direct target of DREB1A/CBF3 [108,109]. Maruyama et al. [109] also analyzed DREB2A overexpression, which did not increase the level of any low-temperature regulated metabolites in transgenic plants. Overexpression of DREB2A-CA in transgenic plants increased their tolerance to dehydration stress, but only slightly increased their tolerance to freezing stress [50]. These results indicate that the increased tolerance to freezing stress in transgenic plants overexpressing DREB1A may depend on the accumulation of low-temperature regulated metabolites, especially sucrose, raffinose, galactinol, and *myo*-inositol. Similarly, transcriptomics and metabolomics analyses of PSEUDO RESPONSE REGULATOR (PRR) arrhythmic triple mutant revealed that the DREB1A/CBF gene and raffinose amounts appear to be regulated by the circadian clock, varying between day and night as if in anticipation of the colder night temperatures [110].

Comparing metabolomics between dehydration, salinity, light, heat or low temperature stress have identified metabolites that are generally important in abiotic stress responses or are specific to each stress [27,95,105,111,112]. In a metabolite profiling study of *Arabidopsis* responses to combined dehydration and heat stresses [95], heat stress reduced the toxicity of proline, indicating that during the more severe combined stress treatment, sucrose replaces proline in plants as the major osmoprotectant. Comparative metabolite



analysis between *Arabidopsis* responding to heat shock and cold shock revealed that the majority of metabolites in response to heat shock overlapped with those produced in response to cold shock [109,113]. These results indicate that a metabolic network of compatible solutes includes proline, monosaccharides (glucose and fructose), galactinol, and raffinose, which have an important role in tolerance to temperature stress. Wienkoop et al. [112] identified a RNA-binding protein (ATGRP7) that increased in response to low temperature stress and decreased in response to high temperature stress. Its abundance was significantly correlated with glutamine and proline concentrations. While raffinose and galactinol concentrations were significant markers for temperature responses, their response was independent of the responses of ATGRP7, proline and glutamine.

Transcriptomics, metabolomics and enzyme activities were integrated in a comprehensive study of K deficiency [114]. Carbon and nitrogen metabolism were significantly affected by K deficiency. This integrated approach pinpointed that pyruvate kinase activity (not transcription) was inhibited directly by K deficiency and was primarily responsible for the metabolic disorders observed.

### **Systematic application of omics technologies has contributed to the development of stress-tolerant crops in the field**

Many genes affect stress tolerance, but few of the identified genes have proven useful in the field. Due to the complexity of stress interactions and stress responses, relevant phenotyping needs to be performed (including field experiments) in abiotic stress studies if we are to make significant progress [113]. The following studies are discussed to highlight good examples of systems biology and omics approaches that have been used to identify key genes regulating stress tolerance and then followed with validation of those responses and phenotypes in multiple experiments including field conditions.

A SNAC1 gene was identified from microarray experiments of stress treatments on rice [115]. SNAC1 is a NAC transcription factor that induces the expression of a number of stress-tolerance genes and improves the drought and salt tolerance of rice in the field. The transgenic plants exhibited increased sensitivity to ABA and reduced water loss. In another drought stress study, a LEA (late embryogenesis abundant) gene was identified from microarray experiments of rice and was transformed and tested in the field under drought conditions through the T3 generation [116]. Spikelet fertility appears to be the main factor contributing to improved yields under drought conditions.

An exhaustive screen of greater than 1500 transcription factors in *Arabidopsis* identified approximately 40

transcription factors that when overexpressed, improved stress tolerance [117]. One of these transcription factors NF-YB1 was further characterized and shown to display significant drought tolerance in *Arabidopsis*. Microarray data of this overexpressing line showed few differences in gene expression and the genes identified were not known previously to be involved in drought tolerance. This functional genomics approach provided a new strategy for improving drought tolerance in plants. A homolog of NF-YB1 was cloned in maize (ZmNF-YB2), overexpressed and tested for drought tolerance in the greenhouse and field plots. The transgenic maize lines were more drought tolerant having increased chlorophyll content, photosynthesis, stomatal conductance and grain yields. One line consistently had more than 50% yield improvement in drought conditions over two different years.

Oh et al. [118] used microarrays to identify 42 AP2 transcription factors whose expressions were affected by stress. Two of these transcription factors, AP37 and AP59 were functionally characterized. The two transcription factors are closely related but have distinct differences in affecting rice phenotype. AP37 responded to drought, salinity, cold and ABA; over-expression improved stress tolerance to all three environmental conditions. AP59 responded to drought and salinity, but not cold or ABA, and improved stress tolerance to drought and salinity only. Both overexpressing lines showed improved photosynthetic efficiency under stress conditions. Overexpression of the transcription factors induced common and distinct sets of genes. T5 homozygous overexpressing lines of AP59, but not AP37, had yield penalties under normal paddy conditions in the field, whereas AP37 overexpressing lines, but not AP59, had enhanced yields under drought conditions in the field. The reduced yields of the overexpressing lines of AP59 were attributed to effects on spikelet development. This study emphasizes the point that it is important to characterize gene effects on yield under field conditions.

### **Mapping stress responses has provided new insights and identified gaps in our knowledge of abiotic stress responses**

From a meta-analysis of drought-stress related papers from the last 15 years, a complex model for plant responses to drought stress was produced [12]. This model details the interactions of sugars, ROS/RNS, hormones (ABA, ethylene, auxins, cytokinins, salicylic acid, gibberellin and brassinosteroids) and nitrogen metabolism. It highlights the highly complex nature of stress responses.

From this review, we have constructed a simplified working model summarizing some of the known plant signaling responses to abiotic stress (Figure 2). Much of

the signaling involves phosphorylation cascades that react quickly in the plant cell, emphasizing the need for proteomics data as well as transcriptomics data in future models. The PYR/PYL/RCAR-PP2C-SnRK2 pathway illustrates that protein phosphorylation and dephosphorylation are the most important factors in ABA signaling. Similar phosphorylation and dephosphorylation processes are involved in ethylene and other abiotic stress signaling pathways (Figure 2). Not all connections could be drawn in this two-dimensional figure without obscuring many other connections. For example, the interactions of ROS with abiotic stresses and hormones [32] are too complex to display here. In addition, the actual signaling response will be dependent upon the signaling pathway present in that organ, tissue or cell at the time of the response. One needs to use more sophisticated bioinformatics programs like Cytoscape [119] and its plug-ins to visualize the interactions comprehensively in two dimensional or three-dimensional space [120] or with time series views [121], which would allow these data to be viewed in four dimensions.

Although there are still some technological issues that must be solved to produce a complete picture of protein phosphorylation, several technologies have been developed for the large-scale analysis of phosphoproteins, known as 'phosphoproteomics' [122]. Mass spectrometry analyses have identified thousands of phosphoproteins in *Arabidopsis*, rice, and *Medicago truncatula* [123-125]. In addition, two studies have reported ABA-responsive changes in the phosphoproteome [126,127]. Phosphoproteomics analyses of mutants for abiotic stress signaling (e.g. PP2C or SnRK) will identify the relevant network of protein phosphorylation events in abiotic stress signaling.

Transcriptome analysis technologies have advanced to the point where high-throughput DNA sequencers and high-density microarrays such as tiling arrays are readily available. These technologies provide new opportunities to analyze noncoding RNAs and can clarify aspects of epigenetic regulation of gene expression [128,129]. Similar approaches [130,131] have revealed the global transcriptomes of plants exposed to abiotic stresses such as dehydration, cold, heat, high-salinity, osmotic stress, and ABA. These analyses indicate that these stresses increase or decrease transcript abundance from not only previously identified stress-responsive genes, but also from thousands of unannotated non-protein-coding regions. Matsui et al. [130] estimated that approximately 80% of previously unannotated upregulated transcripts arise from antisense strands of sense transcripts. There was a significant linear correlation between the expression ratios (stress-treated/untreated) of the sense transcripts and the ratios of the antisense transcripts. Interestingly, the data suggested that such stress-responsive antisense

transcripts are derived from antisense strands of the stress-responsive genes, *RD29A* and *CYP707A1*. Clearly, transcriptional regulation is far more complicated than we previously imagined. Whether or not such antisense transcripts have biological functions is an important issue that remains to be resolved.

Much more research is required in order to fully map plant responses to abiotic stress. The nature of the pathway responses will vary and is highly dependent on the species, organ, tissue, cell type, developmental stage of the plant, the stress or stresses affecting the plant, the level and duration of the stress. Despite the vast amount of research collected on abiotic stress in the last decade, there are still significant gaps in our knowledge. We still do not understand completely how plants perceive stress. We don't know all of the receptors and their sites of action (organs, tissues and cellular components). While we know a lot about downstream signaling (i.e. transcriptional pathways), we know very little about the primary signaling (i.e. proteomics). Most of the literature on abiotic stress responses in plants is based upon transcriptomics data rather than proteomics data. This is not surprising as transcriptomics technology is more advanced, easier to perform and less expensive. However, transcriptomics analyses are insufficient as there is an overall poor correlation of transcriptomics profiles with proteomics profiles [101,132,133] or enzyme activities [104,114]. There are only a few studies describing phosphorylation cascades and other post-translation modification activities in response to stress [134]. Recent efforts to map the hormone [126,127] and light-regulated [135] phosphoproteomes are good first steps. Finally, we need better tools to facilitate systems biology analyses especially in the area of bioinformatics. Transcriptomics data can be collected in a matter of days or weeks, but the data analyses often take more than a year.

## Conclusions

We have made great progress in understanding the responses of plants to abiotic stress. There are inherent physical, morphological and molecular limitations to the plant's ability to respond to stress. Systems biology approaches have given us a more holistic view of the molecular responses. Transcriptomics studies are well advanced, but proteomics analyses are lagging behind, especially the study of post-translational modifications. Plant responses to abiotic stress are dynamic and complex. The integration of multiple omics studies has revealed new areas of interactions and regulation. Time series experiments have revealed the kinetics of stress responses, identifying multiple response phases involving core sets of genes and condition-dependent changes. One consistent trend in response to abiotic stress is the

early down regulation of energy metabolism and protein synthesis. This may indicate a conservation of energy by the plant and may reflect a shift from plant growth to protective mechanisms. In many examples presented in this review, ABA signaling mediates the plant responses to abiotic stress. Co-expression analyses are useful in that they have revealed key regulatory hubs that can be manipulated to produce different phenotypes. To get a comprehensive understanding of plant responses to abiotic stress, more extensive mapping of these responses at the organ, tissue and cellular level are needed. Such network analyses need to be extended to the proteomics and enzyme activities levels. Models need to be constructed and linked to phenotypic traits. The linkage of key regulatory hubs to phenotypic traits will allow for more rapid progress in the genetic manipulation and production of crop plants. Current progress is exemplified by the identification and validation of several key genes that improved stress tolerance of crops in the field. It is expected that progress in the plant sciences and systems biology will continue to accelerate in the near future.

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#### Authors' contributions

GRC contributed to all aspects of this manuscript. KU and KS contributed to the metabolomics and ABA signaling sections and Figure 2. SD contributed to the introduction, ABA and sugar signaling sections. MP contributed to the time-series analyses section. All authors read and approved the final version of this manuscript.

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#### References

1. Kitano H: Systems biology: a brief overview. *Science* 2002, **295**(5560):1662-1664.
2. Feist AM, Palsson BO: The growing scope of applications of genome-scale metabolic reconstructions using *Escherichia coli*. *Nat Biotechnol* 2008, **26**(6):659-667.
3. Bemis SM, Torii KU: Autonomy of cell proliferation and developmental programs during *Arabidopsis* aboveground organ morphogenesis. *Dev Biol* 2007, **304**(1):367-381.
4. Riefler M, Novak O, Strnad M, Schmulling T: *Arabidopsis* cytokinin receptor mutants reveal functions in shoot growth, leaf senescence, seed size, germination, root development, and cytokinin metabolism. *Plant Cell* 2006, **18**(1):40-54.
5. Gibeault DM, Hulett J, Cramer GR, Seemann JR: Maximal biomass of *Arabidopsis thaliana* using a simple, low-maintenance hydroponic method and favorable environmental conditions. *Plant Physiol* 1997, **115**:317-319.
6. Boyer JS: Plant productivity and environment. *Science* 1982, **218**(4571):443-448.
7. Lobell DB, Schlenker W, Costa-Roberts J: Climate trends and global crop production since 1980. *Science* 2011, **333**(6042):616-620.
8. Skirycz A, Inze D: More from less: plant growth under limited water. *Curr Opin Biotechnol* 2010, **21**(2):197-203.
9. Cramer GR: Abiotic stress & plant responses from the whole vine to the genes. *Aust J Grape Wine Res* 2010, **16**:86-93.
10. Dinneny JR, Long TA, Wang JY, Jung JW, Mace D, Pointer S, Barron C, Brady SM, Schiefelbein J, Benfey PN: Cell identity mediates the response of *Arabidopsis* roots to abiotic stress. *Science* 2008, **320**(5878):942-945.
11. Tattersall EA, Grimplet J, Deluc L, Wheatley MD, Vincent D, Osborne C, Ergul A, Lomen E, Blank RR, Schlauch KA, Cushman JC, Cramer GR: Transcript abundance profiles reveal larger and more complex responses of grapevine to chilling compared to osmotic and salinity stress. *Funct Integr Genomics* 2007, **7**(4):317-333.
12. Pinheiro C, Chaves MM: Photosynthesis and drought: can we make metabolic connections from available data? *J Exp Bot* 2011, **62**(3):869-882.
13. Boyer JS: Evans Review: Cell wall biosynthesis and the molecular mechanism of plant enlargement. *Funct Plant Biol* 2009, **36**(5):383-394.
14. Parent B, Hachez C, Redondo E, Simonneau T, Chaumont F, Tardieu F: Drought and abscisic acid effects on aquaporin content translate into changes in hydraulic conductivity and leaf growth rate: a trans-scale approach. *Plant Physiol* 2009, **149**(4):2000-2012.
15. Boursiac Y, Boudet J, Postaire O, Luu DT, Tournaire-Roux C, Maurel C: Stimulus-induced downregulation of root water transport involves reactive oxygen species-activated cell signalling and plasma membrane intrinsic protein internalization. *Plant J* 2008, **56**(2):207-218.
16. Vandeleur RK, Mayo G, Shelden MC, Gilliam M, Kaiser BN, Tyerman SD: The role of PIP aquaporins in water transport through roots: diurnal and drought stress responses reveal different strategies between isohydric and anisohydric cultivars of grapevine. *Plant Physiol* 2008.
17. Nardini A, Lo GMA, Salleo S: Refilling embolized xylem conduits: is it a matter of phloem unloading? *Plant Sci* 2011, **180**(4):604-611.
18. Hummel I, Pantin F, Sulpice R, Piques M, Rolland G, Dauzat M, Christophe A, Pervent M, Bouteille M, Stitt M, Gibon Y, Muller B: *Arabidopsis* plants acclimate to water deficit at low cost through changes of carbon usage: an integrated perspective using growth, metabolite, enzyme, and gene expression analysis. *Plant Physiol* 2010, **154**(1):357-372.
19. Cramer GR, Alberico GJ, Schmidt C: Leaf expansion limits dry matter accumulation of salt-stressed maize. *Aust J Plant Physiol* 1994, **21**:663-674.
20. Nonami H, Wu YJ, Boyer JS: Decreased growth-induced water potential: primary cause of growth inhibition at low water potentials. *Plant Physiol* 1997, **114**(2):501-509.
21. Tang AC, Boyer JS: Growth-induced water potentials and the growth of maize leaves. *J Exp Bot* 2002, **53**(368):489-503.
22. Good AG, Zaplachinski ST: The effects of drought stress on free amino acid accumulation and protein synthesis in brassica napus. *Physiol Plant* 1994, **90**(1):9-14.
23. Vincent D, Ergul A, Bohlman MC, Tattersall EA, Tillett RL, Wheatley MD, Woolsey R, Quilici DR, Joets J, Schlauch K, Schooley DA, Cushman JC, Cramer GR: Proteomic analysis reveals differences between *Vitis vinifera* L. cv. Chardonnay and cv. Cabernet Sauvignon and their responses to water deficit and salinity. *J Exp Bot* 2007, **58**(7):1873-1892.
24. Ben-Zioni A, Itai C, Vaadia Y: Water and salt stresses, kinetin and protein synthesis in tobacco leaves. *Plant Physiol* 1967, **42**:361-365.
25. Dhindsa RS, Cleland RE: Water stress and protein synthesis: I. Differential inhibition of protein synthesis. *Plant Physiol* 1975, **55**(4):778-781.
26. Liu JX, Howell SH: Endoplasmic reticulum protein quality control and its relationship to environmental stress responses in plants. *Plant Cell* 2010, **22**(9):2930-2942.
27. Cramer GR, Ergul A, Grimplet J, Tillett RL, Tattersall EA, Bohlman MC, Vincent D, Sonderegger J, Evans J, Osborne C, Quilici D, Schlauch KA, Schooley DA, Cushman JC: Water and salinity stress in grapevines: early and late changes in transcript and metabolite profiles. *Funct Integr Genomics* 2007, **7**(2):111-134.
28. Kilian J, Whitehead D, Horak J, Wanke D, Weinel S, Batistic O, D'Angelo C, Bornberg-Bauer E, Kudla J, Harter K: The AtGenExpress global stress expression data set: protocols, evaluation and model data analysis of UV-B light, drought and cold stress responses. *Plant J* 2007, **50**(2):347-363.
29. Takahashi S, Seki M, Ishida J, Satou M, Sakurai T, Narusaka M, Kamiya A, Nakajima M, Enju A, Akiyama K, Yamaguchi-Shinozaki K, Shinozaki K: Monitoring the expression profiles of genes induced by hyperosmotic, high salinity, and oxidative stress and abscisic acid treatment in



- Arabidopsis cell culture using a full-length cDNA microarray. *Plant Mol Biol* 2004, **56**(1):29-55.**
30. Molassiotis A, Fotopoulos V: **Oxidative and nitrosative signaling in plants: two branches in the same tree?** *Plant Signal Behav* 2011, **6**(2):210-214.
  31. Wilkinson S, Davies WJ: **Drought, ozone, ABA and ethylene: new insights from cell to plant to community.** *Plant Cell Environ* 2009.
  32. Mittler R, Vanderauwera S, Suzuki N, Miller G, Tognetti VB, Vandepoel K, Gollery M, Shulaev V, Van BF: **ROS signaling: the new wave?** *Trends Plant Sci* 2011, **16**(6):300-309.
  33. Goda H, Sasaki E, Akiyama K, Maruyama-Nakashita A, Nakabayashi K, Li W, Ogawa M, Yamauchi Y, Preston J, Aoki K, Kiba T, Takatsuto S, Fujioaka S, Asami T, Nakano T, Kato H, Mizuno T, Sakakibara H, Yamaguchi S, Nambara E, Kamiya Y, Takahashi H, Hirai MY, Sakurai T, Shinozaki K, Saito K, Yoshida S, Shimada Y: **The AtGenExpress hormone and chemical treatment data set: experimental design, data evaluation, model data analysis and data access.** *Plant J* 2008, **55**(3):526-542.
  34. Hubbard KE, Nishimura N, Hitomi K, Getzoff ED, Schroeder JI: **Early abscisic acid signal transduction mechanisms: newly discovered components and newly emerging questions.** *Genes Dev* 2010, **24**(16):1695-1708.
  35. Kim TH, Bohmer M, Hu H, Nishimura N, Schroeder JI: **Guard cell signal transduction network: advances in understanding abscisic acid, CO<sub>2</sub>, and Ca<sup>2+</sup> signaling.** *Annu Rev Plant Biol* 2010, **61**:561-591.
  36. Chinnusamy V, Gong Z, Zhu JK: **Abscisic acid-mediated epigenetic processes in plant development and stress responses.** *J Integr Plant Biol* 2008, **50**(10):1187-1195.
  37. Umezawa T: **Systems biology approaches to abscisic acid signaling.** *J Plant Res* 2011, **124**(4):539-548.
  38. Ma Y, Szostkiewicz I, Korte A, Moes D, Yang Y, Christmann A, Grill E: **Regulators of PP2C phosphatase activity function as abscisic acid sensors.** *Science* 2009, **324**(5930):1064-1068.
  39. Park SY, Jung P, Nishimura N, Jensen DR, Fujii H, Zhao Y, Lumba S, Santiago J, Rodrigues A, Chow TF, Alfred SE, Bonetta D, Finkelstein R, Provart NJ, Desveaux D, Rodriguez PL, McCourt P, Zhu JK, Schroeder JI, Volkman BF, Cutler SR: **Abscisic acid inhibits type 2C protein phosphatases via the PYR/PYL family of START proteins.** *Science* 2009, **324**(5930):1068-1071.
  40. Leung J, Giraudat J: **Abscisic acid signal transduction.** *Annu Rev Plant Physiol Plant Mol Biol* 1998, **49**:199-222.
  41. Yoshida R, Hobo T, Ichimura K, Mizoguchi T, Takahashi F, Aronso J, Ecker JR, Shinozaki K: **ABA-activated SnRK2 protein kinase is required for dehydration stress signaling in Arabidopsis.** *Plant Cell Physiol* 2002, **43**(12):1473-1483.
  42. Mustilli AC, Merlot S, Vavasour A, Fenzi F, Giraudat J: **Arabidopsis OST1 protein kinase mediates the regulation of stomatal aperture by abscisic acid and acts upstream of reactive oxygen species production.** *Plant Cell* 2002, **14**(12):3089-3099.
  43. Umezawa T, Nakashima K, Miyakawa T, Kuromori T, Tanokura M, Shinozaki K, Yamaguchi-Shinozaki K: **Molecular basis of the core regulatory network in aba responses: sensing, signaling and transport.** *Plant Cell Physiol* 2010, **51**(11):1821-1839.
  44. Yamaguchi-Shinozaki K, Shinozaki K: **Transcriptional regulatory networks in cellular responses and tolerance to dehydration and cold stresses.** *Annu Rev Plant Biol* 2006, **57**:781-803.
  45. Fujita Y, Fujita M, Satoh R, Maruyama K, Parvez MM, Seki M, Hiratsu K, Ohme-Takagi M, Shinozaki K, Yamaguchi-Shinozaki K: **AREB1 is a transcription activator of novel ABRE-dependent ABA signaling that enhances drought stress tolerance in Arabidopsis.** *Plant Cell* 2005, **17**(12):3470-3488.
  46. Kang JY, Choi HI, Im MY, Kim SY: **Arabidopsis basic leucine zipper proteins that mediate stress-responsive abscisic acid signaling.** *Plant Cell* 2002, **14**(2):343-357.
  47. Yoshida T, Fujita Y, Sayama H, Maruyama K, Mizoi J, Shinozaki K, Yamaguchi-Shinozaki K: **AREB1, AREB2, and ABF3 are master transcription factors that cooperatively regulate ABRE-dependent ABA signaling involved in drought stress tolerance and require ABA for full activation.** *Plant J* 2010, **61**(4):672-685.
  48. Fowler S, Thomashow MF: **Arabidopsis transcriptome profiling indicates that multiple regulatory pathways are activated during cold acclimation in addition to the CBF cold response pathway.** *Plant Cell* 2002, **14**(8):1675-1690.
  49. Maruyama K, Sakuma Y, Kasuga M, Ito Y, Seki M, Goda H, Shimada Y, Yoshida S, Shinozaki K, Yamaguchi-Shinozaki K: **Identification of cold-inducible downstream genes of the Arabidopsis DREB1A/CBF3 transcriptional factor using two microarray systems.** *Plant J* 2004, **38**(6):982-993.
  50. Sakuma Y, Maruyama K, Osakabe Y, Qin F, Seki M, Shinozaki K, Yamaguchi-Shinozaki K: **Functional analysis of an Arabidopsis transcription factor, DREB2A, involved in drought-responsive gene expression.** *Plant Cell* 2006, **18**(5):1292-1309.
  51. Stepanova AN, Alonso JM: **Ethylene signaling and response: where different regulatory modules meet.** *Curr Opin Plant Biol* 2009, **12**(5):548-555.
  52. Yoo SD, Cho Y, Sheen J: **Emerging connections in the ethylene signaling network.** *Trends Plant Sci* 2009, **14**(5):270-279.
  53. Morgan PW, Drew MC: **Ethylene and plant responses to stress.** *Physiol Plant* 1997, **100**(3):620-630.
  54. Zhang M, Leng P, Zhang G, Li X: **Cloning and functional analysis of 9-cis-epoxycarotenoid dioxygenase (NCED) genes encoding a key enzyme during abscisic acid biosynthesis from peach and grape fruits.** *J Plant Physiol* 2009, **166**(12):1241-1252.
  55. Sun L, Zhang M, Ren J, Qi J, Zhang G, Leng P: **Reciprocity between abscisic acid and ethylene at the onset of berry ripening and after harvest.** *BMC Plant Biol* 2010, **10**:257.
  56. Ophir R, Pang X, Halaly T, Venkateswari J, Lavee S, Galbraith D, Or E: **Gene-expression profiling of grape bud response to two alternative dormancy-release stimuli expose possible links between impaired mitochondrial activity, hypoxia, ethylene-ABA interplay and cell enlargement.** *Plant Mol Biol* 2009, **71**(4-5):403-423.
  57. Petranovic D, Tyo K, Vemuri GN, Nielsen J: **Prospects of yeast systems biology for human health: integrating lipid, protein and energy metabolism.** *FEMS Yeast Res* 2010, **10**(8):1046-1059.
  58. Zaborske JM, Wu X, Wek RC, Pan T: **Selective control of amino acid metabolism by the GCN2 eIF2 kinase pathway in Saccharomyces cerevisiae.** *BMC Biochem* 2010, **11**:29.
  59. Staschke KA, Dey S, Zaborske JM, Palam LR, McClintock JN, Pan T, Edenberg HJ, Wek RC: **Integration of general amino acid control and target of rapamycin (TOR) regulatory pathways in nitrogen assimilation in yeast.** *J Biol Chem* 2010, **285**(22):16893-16911.
  60. Smeekens S, Ma J, Hanson J, Rolland F: **Sugar signals and molecular networks controlling plant growth.** *Curr Opin Plant Biol* 2010, **13**(3):274-279.
  61. Hey SJ, Byrne E, Halford NG: **The interface between metabolic and stress signalling.** *Ann Bot* 2010, **105**(2):197-203.
  62. Deprost D, Yao L, Sormani R, Moreau M, Letterreux G, Nicolai M, Bedu M, Robaglia C, Meyer C: **The Arabidopsis TOR kinase links plant growth, yield, stress resistance and mRNA translation.** *EMBO Rep* 2007, **8**(9):864-870.
  63. Lageix S, Lanet E, Pouch-Pelissier MN, Espagnol MC, Robaglia C, Deragon JM, Pelissier T: **Arabidopsis eIF2alpha kinase GCN2 is essential for growth in stress conditions and is activated by wounding.** *BMC Plant Biol* 2008, **8**:134.
  64. Baena-Gonzalez E, Rolland F, Thevelein JM, Sheen J: **A central integrator of transcription networks in plant stress and energy signalling.** *Nature* 2007, **448**(7156):938-942.
  65. Baena-Gonzalez E, Sheen J: **Convergent energy and stress signaling.** *Trends Plant Sci* 2008, **13**(9):474-482.
  66. Jossier M, Bouly JP, Meimoun P, Arjmand A, Lessard P, Hawley S, Grahame HD, Thomas M: **SnRK1 (SNF1-related kinase 1) has a central role in sugar and ABA signalling in Arabidopsis thaliana.** *Plant J* 2009, **59**(2):316-328.
  67. Obertello M, Krouk G, Katari MS, Runko SJ, Coruzzi GM: **Modeling the global effect of the basic-leucine zipper transcription factor 1 (bZIP1) on nitrogen and light regulation in Arabidopsis.** *BMC Systems Biol* 2010, **4**:111.
  68. Kang SG, Price J, Lin PC, Hong JC, Jang JC: **The arabidopsis bZIP1 transcription factor is involved in sugar signaling, protein networking, and DNA binding.** *Mol Plant* 2010, **3**(2):361-373.
  69. Osuna D, Usadel B, Morcuende R, Gibon Y, Blasing OE, Hohne M, Gunter M, Kamlage B, Trethewey R, Scheible WR, Stitt M: **Temporal responses of transcripts, enzyme activities and metabolites after adding sucrose to carbon-deprived Arabidopsis seedlings.** *Plant J* 2007, **49**(3):463-491.



70. Zhou L, Jang JC, Jones TL, Sheen J: **Glucose and ethylene signal transduction crosstalk revealed by an Arabidopsis glucose-insensitive mutant.** *Proc Natl Acad Sci USA* 1998, **95**(17):10294-10299.
71. Arenas-Huetero F, Arroyo A, Zhou L, Sheen J, Leon P: **Analysis of Arabidopsis glucose insensitive mutants, gin5 and gin6, reveals a central role of the plant hormone ABA in the regulation of plant vegetative development by sugar.** *Genes Dev* 2000, **14**(16):2085-2096.
72. Finkelstein RR, Gibson SL: **ABA and sugar interactions regulating development: cross-talk or voices in a crowd?** *Curr Opin Plant Biol* 2002, **5**(1):26-32.
73. Franco-Zorrilla JM, Martin AC, Leyva A, Paz-Ares J: **Interaction between phosphate-starvation, sugar, and cytokinin signaling in Arabidopsis and the roles of cytokinin receptors CRE1/AHK4 and AHK3.** *Plant Physiol* 2005, **138**(2):847-857.
74. Mita S, Suzuki-Fujii K, Nakamura K: **Sugar-inducible expression of a gene for beta-amylase in Arabidopsis thaliana.** *Plant Physiol* 1995, **107**(3):895-904.
75. Thum KE, Shasha DE, Lejay LV, Coruzzi GM: **Light- and carbon-signaling pathways. Modeling circuits of interactions.** *Plant Physiol* 2003, **132**(2):440-452.
76. Cakir B, Agasse A, Gaillard C, Saumonneau A, Delrot S, Atanassova R: **A grape ASR protein involved in sugar and abscisic acid signaling.** *Plant Cell* 2003, **15**(9):2165-2180.
77. Lecourieux F, Lecourieux D, Vignault C, Delrot S: **A sugar inducible protein kinase, VvSK1, regulates hexose transport and sugar accumulation in grapevine cells.** *Plant Physiol* 2010, **52**:1096-1106.
78. Shen W, Reyes MI, Hanley-Bowdoin L: **Arabidopsis protein kinases GRIK1 and GRIK2 specifically activate SnRK1 by phosphorylating its activation loop.** *Plant Physiol* 2009, **150**(2):996-1005.
79. Tiessen A, Prescha K, Branscheid A, Palacios N, McKibbin R, Halford NG, Geigenberger P: **Evidence that SNF1-related kinase and hexokinase are involved in separate sugar-signalling pathways modulating post-translational redox activation of ADP-glucose pyrophosphorylase in potato tubers.** *Plant J* 2003, **35**(4):490-500.
80. Zheng Z, Xu X, Crosley RA, Greenwalt SA, Sun Y, Blakeslee B, Wang L, Ni W, Sopko MS, Yao C, Yau K, Burton S, Zhuang M, McCaskill DG, Gachotte D, Thompson M, Greene TW: **The protein kinase SnRK2.6 mediates the regulation of sucrose metabolism and plant growth in Arabidopsis.** *Plant Physiol* 2010, **153**(1):99-113.
81. Lenz T, Fischer JJ, Dregler M: **Probing small molecule-protein interactions: A new perspective for functional proteomics.** *J Proteomics* 2011.
82. Kaufmann K, Smaczniak C, de VS, Angenent GC, Karlava R: **Proteomics insights into plant signaling and development.** *Proteomics* 2011, **11**(4):744-755.
83. Usadel B, Obayashi T, Mutwil M, Giorgi FM, Bassel GW, Tanimoto M, Chow A, Steinhäuser D, Persson S, Provart NJ: **Co-expression tools for plant biology: opportunities for hypothesis generation and caveats.** *Plant Cell Environ* 2009, **32**(12):1633-1651.
84. Hirai MY, Yano M, Goodenow DB, Kanaya S, Kimura T, Awazuhara M, Arita M, Fujiwara T, Saito K: **Integration of transcriptomics and metabolomics for understanding of global responses to nutritional stresses in Arabidopsis thaliana.** *Proc Natl Acad Sci USA* 2004, **101**(27):10205-10210.
85. Hirai MY, Sugiyama K, Sawada Y, Tohge T, Obayashi T, Suzuki A, Araki R, Sakurai N, Suzuki H, Aoki K, Goda H, Nishizawa OI, Shibata D, Saito K: **Omics-based identification of Arabidopsis Myb transcription factors regulating aliphatic glucosinolate biosynthesis.** *Proc Natl Acad Sci USA* 2007, **104**(15):6478-6483.
86. Mao L, Van HJL, Dash S, Dickerson JA: **Arabidopsis gene co-expression network and its functional modules.** *BMC Bioinformatics* 2009, **10**:346.
87. Carrera J, Rodrigo G, Jaramillo A, Elena SF: **Reverse-engineering the Arabidopsis thaliana transcriptional network under changing environmental conditions.** *Genome Biol* 2009, **10**(9):R96.
88. Lorenz WW, Alba R, Yu YS, Bordeaux JM, Simoes M, Dean JF: **Microarray analysis and scale-free gene networks identify candidate regulators in drought-stressed roots of loblolly pine (P. taeda L.).** *BMC Genomics* 2011, **12**:264.
89. Weston DJ, Gunter LE, Rogers A, Wulfschleger SD: **Connecting genes, coexpression modules, and molecular signatures to environmental stress phenotypes in plants.** *BMC Systems Biol* 2008, **2**:16.
90. Weston DJ, Karve AA, Gunter LE, Jawdy SS, Yang X, Allen SM, Wulfschleger SD: **Comparative physiology and transcriptional networks underlying the heat shock response in Populus trichocarpa, Arabidopsis thaliana and Glycine max.** *Plant Cell Environ* 2011.
91. Sun X, Zou Y, Nikiforova V, Kurths J, Walther D: **The complexity of gene expression dynamics revealed by permutation entropy.** *BMC Bioinformatics* 2010, **11**:607.
92. Vanderauwera S, Zimmermann P, Rombauts S, Vandenabeele S, Langebartels C, Grisse W, Inze D, Van BF: **Genome-wide analysis of hydrogen peroxide-regulated gene expression in Arabidopsis reveals a high light-induced transcriptional cluster involved in anthocyanin biosynthesis.** *Plant Physiol* 2005, **139**(2):806-821.
93. Tognetti VB, Van AO, Morreel K, Vandenbroucke K, van dCB, De CI, Chiwocha S, Fenske R, Prinsen E, Boerjan W, Genty B, Stubbs KA, Inze D, Van BF: **Perturbation of indole-3-butyric acid homeostasis by the UDP-glucosyltransferase UGT74E2 modulates Arabidopsis architecture and water stress tolerance.** *Plant Cell* 2010, **22**(8):2660-2679.
94. Vanderauwera S, De BM, Van dSN, van dCB, Metzlaiff M, Van BF: **Silencing of poly(ADP-ribose) polymerase in plants alters abiotic stress signal transduction.** *Proc Natl Acad Sci USA* 2007, **104**(38):15150-15155.
95. De Block M, Verduyn C, De Brouwer D, Cornelissen M: **Poly(ADP-ribose) polymerase in plants affects energy homeostasis, cell death and stress tolerance.** *Plant J* 2005, **41**(1):95-106.
96. Kusano M, Tohge T, Fukushima A, Kobayashi M, Hayashi N, Otsuki H, Kondou Y, Goto H, Kawashima M, Matsuda F, Niida R, Matsui M, Saito K, Fernie AR: **Metabolomics reveals comprehensive reprogramming involving two independent metabolic responses of Arabidopsis to UV-B light.** *Plant J* 2011, **67**(2):354-369.
97. Wilkins O, Brautigam K, Campbell MM: **Time of day shapes Arabidopsis drought transcriptomes.** *Plant J* 2010, **63**(5):715-727.
98. Raj S, Brautigam K, Hamanishi ET, Wilkins O, Thomas BR, Schroeder W, Mansfield SD, Plant AL, Campbell MM: **Clone history shapes Populus drought responses.** *Proc Natl Acad Sci USA* 2011.
99. Grimplet J, Deluc LG, Tillett RL, Wheatley MD, Schlauch KA, Cramer GR, Cushman JC: **Tissue-specific mRNA expression profiling in grape berry tissues.** *BMC Genomics* 2007, **8**:187.
100. Deluc LG, Quilici DR, Decendit A, Grimplet J, Wheatley MD, Schlauch KA, Merillon JM, Cushman JC, Cramer GR: **Water deficit alters differentially metabolic pathways affecting important flavor and quality traits in grape berries of Cabernet Sauvignon and Chardonnay.** *BMC Genomics* 2009, **10**:212.
101. Grimplet J, Wheatley MD, Jouira HB, Deluc LG, Cramer GR, Cushman JC: **Proteomic and selected metabolite analysis of grape berry tissues under well watered and water-deficit stress conditions.** *Proteomics* 2009, **9**:2503-2528.
102. Deluc LG, Decendit A, Papastamoulis Y, Merillon JM, Cushman JC, Cramer GR: **Water Deficit Increases Stilbene Metabolism in Cabernet Sauvignon Berries.** *J Agric Food Chem* 2011, **59**(1):289-297.
103. Zamboni A, Di Carli M, Guzzo F, Stocchero M, Zenoni S, Ferrarini A, Tononi P, Toffali K, Desiderio A, Lilley KS, Pe ME, Benvenuto E, Delledonne M, Pezzotti M: **Identification of putative stage-specific grapevine berry biomarkers and omics data integration into networks.** *Plant Physiol* 2010, **154**(3):1439-1459.
104. Usadel B, Blasing OE, Gibon Y, Poree F, Hohne M, Gunter M, Trethewey R, Kamlage B, Poorter H, Stitt M: **Multilevel genomic analysis of the response of transcripts, enzyme activities and metabolites in Arabidopsis rosettes to a progressive decrease of temperature in the non-freezing range.** *Plant Cell Environ* 2008, **31**(4):518-547.
105. Caldana C, Degenkolbe T, Cuadros-Inostroza A, Klie S, Sulpice R, Leisse A, Steinhäuser D, Fernie AR, Willmitzer L, Hannah MA: **High-density kinetic analysis of the metabolomic and transcriptomic response of Arabidopsis to eight environmental conditions.** *Plant J* 2011.
106. Yun KY, Park MR, Mohanty B, Herath V, Xu F, Mauleon R, Wijaya E, Bajic VB, Bruskiewich R, de LRBG: **Transcriptional regulatory network triggered by oxidative signals configures the early response mechanisms of japonica rice to chilling stress.** *BMC Plant Biol* 2010, **10**:16.
107. Urano K, Maruyama K, Ogata Y, Morishita Y, Takeda M, Sakurai N, Suzuki H, Saito K, Shibata D, Kobayashi M, Yamaguchi-Shinozaki K, Shinozaki K: **Characterization of the ABA-regulated global responses to dehydration in Arabidopsis by metabolomics.** *Plant J* 2009, **7**:1065-1078.

108. Cook D, Fowler S, Fiehn O, Thomashow MF: **A prominent role for the CBF cold response pathway in configuring the low-temperature metabolome of Arabidopsis.** *Proc Natl Acad Sci USA* 2004, **101**(42):15243-15248.
109. Maruyama K, Takeda M, Kidokoro S, Yamada K, Sakuma Y, Urano K, Fujita M, Yoshiwara K, Matsukura S, Morishita Y, Sasaki R, Suzuki H, Saito K, Shibata D, Shinozaki K, Yamaguchi-Shinozaki K: **Metabolic pathways involved in cold acclimation identified by integrated analysis of metabolites and transcripts regulated by DREB1A and DREB2A.** *Plant Physiol* 2009, **150**(4):1972-1980.
110. Nakamichi N, Kusano M, Fukushima A, Kita M, Ito S, Yamashino T, Saito K, Sakakibara H, Mizuno T: **Transcript profiling of an Arabidopsis PSEUDO RESPONSE REGULATOR arrhythmic triple mutant reveals a role for the circadian clock in cold stress response.** *Plant Cell Physiol* 2009, **50**(3):447-462.
111. Kaplan F, Kopka J, Haskell DW, Zhao W, Schiller KC, Gatzke N, Sung DY, Guy CL: **Exploring the temperature-stress metabolome of Arabidopsis.** *Plant Physiol* 2004, **136**(4):4159-4168.
112. Wienkoop S, Morgenthal K, Wolschin F, Scholz M, Selbig J, Weckwerth W: **Integration of metabolomic and proteomic phenotypes: analysis of data covariance dissects starch and RFO metabolism from low and high temperature compensation response in Arabidopsis thaliana.** *Mol Cell Proteomics* 2008, **7**(9):1725-1736.
113. Salekdeh GH, Reynolds M, Bennett J, Boyer J: **Conceptual framework for drought phenotyping during molecular breeding.** *Trends Plant Sci* 2009, **14**(9):488-496.
114. Armengaud P, Sulpice R, Miller AJ, Stitt M, Amtmann A, Gibon Y: **Multilevel analysis of primary metabolism provides new insights into the role of potassium nutrition for glycolysis and nitrogen assimilation in Arabidopsis roots.** *Plant Physiol* 2009, **150**(2):772-785.
115. Hu H, Dai M, Yao J, Xiao B, Li X, Zhang Q, Xiong L: **Overexpressing a NAM, ATAF, and CUC (NAC) transcription factor enhances drought resistance and salt tolerance in rice.** *Proc Natl Acad Sci USA* 2006, **103**(35):12987-12992.
116. Xiao B, Huang Y, Tang N, Xiong L: **Over-expression of a LEA gene in rice improves drought resistance under the field conditions.** *TAG Theoretical and applied genetics Theoretische und angewandte Genetik* 2007, **115**(1):35-46.
117. Nelson DE, Repetti PP, Adams TR, Creelman RA, Wu J, Warner DC, Anstrom DC, Bensen RJ, Castiglioni PP, Donnarummo MG, Hinchey BS, Kumimoto RW, Maszle DR, Canales RD, Krolkowski KA, Dotson SB, Gutterson N, Ratcliffe OJ, Heard JE: **Plant nuclear factor Y (NF-Y) B subunits confer drought tolerance and lead to improved corn yields on water-limited acres.** *Proc Natl Acad Sci USA* 2007, **104**(42):16450-16455.
118. Oh SJ, Kim YS, Kwon CW, Park HK, Jeong JS, Kim JK: **Overexpression of the transcription factor AP37 in rice improves grain yield under drought conditions.** *Plant Physiol* 2009, **150**(3):1368-1379.
119. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, Amin N, Schwikowski B, Ideker T: **Cytoscape: a software environment for integrated models of biomolecular interaction networks.** *Genome Res* 2003, **13**(11):2498-2504.
120. Audenaert P, Van PT, Brondel F, Pickavet M, Demeester P, Van dPY, Michoel T: **CyClus3D: a Cytoscape plugin for clustering network motifs in integrated networks.** *Bioinformatics* 2011, **27**(11):1587-1588.
121. Xia T, Hemert JV, Dickerson JA: **OmicsAnalyzer: a Cytoscape plug-in suite for modeling omics data.** *Bioinformatics* 2010, **26**(23):2995-2996.
122. Schulze WX: **Proteomics approaches to understand protein phosphorylation in pathway modulation.** *Curr Opin Plant Biol* 2010, **13**(3):280-287.
123. Sugiyama N, Nakagami H, Mochida K, Daudi A, Tomita M, Shirasu K, Ishihama Y: **Large-scale phosphorylation mapping reveals the extent of tyrosine phosphorylation in Arabidopsis.** *Mol Syst Biol* 2008, **4**:193.
124. Nakagami H, Sugiyama N, Mochida K, Daudi A, Yoshida Y, Toyoda T, Tomita M, Ishihama Y, Shirasu K: **Large-scale comparative phosphoproteomics identifies conserved phosphorylation sites in plants.** *Plant Physiol* 2010, **153**(3):1161-1174.
125. de la Fuente van Bentem S, Anrather D, Roitinger E, Djamei A, Hufnagl T, Barta A, Csaszar E, Dohnal I, Lecourieux D, Hirt H: **Phosphoproteomics reveals extensive in vivo phosphorylation of Arabidopsis proteins involved in RNA metabolism.** *Nucleic Acids Res* 2006, **34**(11):3267-3278.
126. Chen Y, Hoehenwarter W, Weckwerth W: **Comparative analysis of phytohormone-responsive phosphoproteins in Arabidopsis thaliana using TiO2-phosphopeptide enrichment and mass accuracy precursor alignment.** *Plant J* 2010, **63**(1):1-17.
127. Kline KG, Barrett-Wilt GA, Sussman MR: **In planta changes in protein phosphorylation induced by the plant hormone abscisic acid.** *Proc Natl Acad Sci USA* 2010, **107**(36):15986-15991.
128. Zhang X, Yazaki J, Sundaresan A, Cokus S, Chan SW, Chen H, Henderson IR, Shinn P, Pellegrini M, Jacobsen SE, Ecker JR: **Genome-wide high-resolution mapping and functional analysis of DNA methylation in Arabidopsis.** *Cell* 2006, **126**(6):1189-1201.
129. Gregory BD, Yazaki J, Ecker JR: **Utilizing tiling microarrays for whole-genome analysis in plants.** *Plant J* 2008, **53**(4):636-644.
130. Matsui A, Ishida J, Morosawa T, Mochizuki Y, Kaminuma E, Endo TA, Okamoto M, Nambara E, Nakajima M, Kawashima M, Satou M, Kim JM, Kobayashi N, Toyoda T, Shinozaki K, Seki M: **Arabidopsis transcriptome analysis under drought, cold, high-salinity and ABA treatment conditions using a tiling array.** *Plant Cell Physiol* 2008, **49**(8):1135-1149.
131. Zeller G, Henz SR, Widmer CK, Sachsenberg T, Ratsch G, Weigel D, Laubinger S: **Stress-induced changes in the Arabidopsis thaliana transcriptome analyzed using whole-genome tiling arrays.** *Plant J* 2009, **58**(6):1068-1082.
132. Nie L, Wu G, Culley DE, Scholten JC, Zhang W: **Integrative analysis of transcriptomic and proteomic data: challenges, solutions and applications.** *Crit Rev Biotechnol* 2007, **27**(2):63-75.
133. Bohmer M, Schroeder JI: **Quantitative transcriptomic analysis of abscisic acid-induced and reactive oxygen species-dependent expression changes and proteomic profiling in Arabidopsis suspension cells.** *Plant J* 2011, **67**(1):105-118.
134. Yang S, Vanderbeld B, Wan J, Huang Y: **Narrowing down the targets: towards successful genetic engineering of drought-tolerant crops.** *Mol Plant* 2010, **3**(3):469-490.
135. Reiland S, Finazzi G, Endler A, Willig A, Baerenfaller K, Grossmann J, Gerrits B, Rutishauser D, Gruissem W, Rochaix JD, Baginsky S: **Comparative phosphoproteome profiling reveals a function of the STN8 kinase in fine-tuning of cyclic electron flow (CEF).** *Proc Natl Acad Sci USA* 2011.

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