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Investigating foliar application of bulk and nanoparticles titanium dioxide on fennel productivity to mitigate the negative effects of saline irrigation water

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Abstract

Background Fennel essential oils are fragrance compounds used in food and pharmaceutical sectors. One of the major impediments to expansion of fennel farming in Egypt's reclamation areas is saline water. Titanium dioxide (TiO₂) or TiO₂ nano particles (TiO₂NP) can be utilized to boost the yield of aromatic plants cultivated under saline irrigation water. Saline water, particularly which contains sodium chloride can harm fennel plant; consequently, it was predicted that fennel production would fail in Egypt's reclaimed area, where the primary source of irrigation is groundwater consisting sodium chloride. This study sought to help fennel respond to sodium chloride by applying Ti forms to their leaves in order to reduce the detrimental effects of sodium chloride on them for expanding their production in the newly reclamation areas as a natural source of essential oil. Ti forms were applied as foliar application at 0, 0.1, 0.2 TiO₂, 0.1 TiO₂NP, and 0.2 TiO₂NP, mM under irrigation with fresh water (0.4 dS m⁻¹), or saline water (51.3 mM or 4.7 dS m⁻¹).

Results Plants exposed to 0.1 mM TiO₂NP under fresh water resulted in the maximum values of morphological characters, estragole, oxygenated monoterpenes and photosynthetic pigments; while those subjected to 0.1 mM TiO₂NP under saline water gave the greatest values of essential oil, proline, antioxidant enzymes and phenols. The greatest amounts of soluble sugars were recorded with 0.2 mM TiO₂NP irrigated with saline water. Plants subjected to 0 mM TiO₂ under saline water produced the greatest values of flavonoids, hydrogen peroxide and malondialdehyde.

Conclusion To mitigate the negative effects of salty irrigation water on fennel plant production, TiO₂NP application is suggested as a potential strategy.

Keywords Fennel, Irrigation water, Ti forms, Essential oil, Chemical fractions

Introduction

Herbs have undergone substantial research due to the growing demand for their usage in different applications, including food, medicine and pharmaceuticals. Herbs provide a variety of byproducts that are economically valuable for raising the country's income [1]. The most vital active components of aromatic herbs are essential oils (EOs) [2]; they can be used in numerous industries including medicines, cosmetics, perfumes, fragrances, flavors, food additives, agrochemicals and biopesticides

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[3]. According to scientific research, utilizing EOs from aromatic herbs is preferable than using synthetic chemicals for a number of reasons because the latter have harmful side effects, particularly carcinogenic ones [3]. Fennel (*Foeniculum vulgare* Mill) is a medicinal and spice herb belonging to Apiaceae family [4]; its EO is used in food, aromatherapy and pharmaceutical industries as flavoring and fragrance ingredients [5].

Increasing population has caused serious problems in today's societies due to the lack of suitable regions for agriculture; governments frequently increase the cultivation of desert lands, which are frequently found in arid or semi-arid regions [6]. Water shortage is a global issue that is getting worse in arid and semi-arid regions; poor quality water, including salty groundwater is frequently utilized to make up for this deficit [7]. Using saline groundwater, which normally contains solutes in different amounts significantly, alters soil properties. With use of salt water for irrigation, soil becomes more salinized and salt deposits form in the root zone, which causes poor plant development and degradation [8]. Where, high amounts of salted irrigation water (SALIW) can lead to sodium and reactive oxygen species (ROS) buildup in plant tissues, decreases in basic elements and water uptakes causing oxidative and ionic stress, imbalance in nutrients, injuries to cell membranes and a drop in photosynthetic pigments (POTSP), which impedes development and growth [9]. On the other hand, SALIW is crucial for the production of proteins (PROTs), accumulation of total phenols (TPHEN) and total flavonoids (TFLAV), lipid metabolism, generation of carbohydrates (CARB) and a variety of natural product metabolites, particularly the composition of EOs in aromatic plants [10–13]. Impact of SALIW treatments on the equilibrium and uptake of essential macro- and micronutrients; this lowers protein and chlorophyll levels, which in turn cause reductions in growth and dry matter values [10–13]. Due to its ability to scavenge ROS, store CARB, and provide osmo-protection, CARB may be the primary factor contributing to the increase in CARB under SALIW [12,13]. In response to SALIW, plants produce more TPHEN, TFLAV, and EO, which is in line with their function as antioxidants that reduce oxidative damage [10–13]. The effects of SALIW vary depending on plant species [13, 14]. SALIW with sodium chloride (NaCl) in fennel leads to decreased plant growth, fresh and fruit yield, relative water content, EO, lipids, oleic and linoleic acids, POTSP, potassium (K), calcium (Ca) and magnesium (Mg); while, proline (PROL), CARB, TPHEN, malondialdehyde (MDA), hydrogen peroxide (H_2O_2), antioxidant enzymes activity (AOEA), total soluble solids (TSS), sodium (Na), major components of EO (limonene, fenchone, estragol, *trans*-anethol) and palmitic acid increased [15–21].

Scientific studies have been carried out to examine a number of strategies to increase the output of aromatic plants under stressful situations. Titanium (Ti) as titanium dioxide (TiO_2) or TiO_2 Nano particles (TiO_2 NP) can be used to increase the output of aromatic herbs grown under salt stress [22]. Ti is a favorable element found in the majority of rocks, sediments and sands [23], it is the tenth most prevalent element in the Earth's crust; furthermore, it is second transition metal [23]. Ti has a great affinity for oxygen, which it can be found naturally as TiO_2 [24]. TiO_2 NP may be produced naturally and is utilized in a variety of industrial applications [24]. Low concentrations of Ti applied through the roots or leaves have been reported to enhance crop performance by promoting the activity of specific enzymes, increasing chlorophyll content and photosynthesis delaying in chloroplast senescence, promoting nutrient uptake, enhancing stress tolerance and improve the plant production [25]. Ti forms (TiO_2 or TiO_2 NP) had a favorable impact on plant development, antioxidant capacity, AOEa activities, total soluble sugars (TSOLS), amino acids, EOs and their major constituents, pH value, titratable acidity (TA), electrical conductivity (EC) and TSS; as well as a decrease in H_2O_2 and MDA contents in plants grown in SALIW [26–28].

Egyptian government is working to increase the production of medicinal and aromatic plants that are grown in desert areas. This is in the context of the government's dedication to strengthening pharmaceutical economics and supporting the pharmaceutical sectors in Egypt. Unfortunately, those areas have only groundwater that contains salts, especially NaCl. Previous research has shown that fennel plant can tolerate small amounts of SALIW, but that excessive salt concentration, especially those containing NaCl can injure fennel plant; prior research has demonstrated that fennel plants can withstand saline levels of up to 1.6 dS m^{-1} ; above that point, the plant's output and active ingredient concentration start to decline [21]. Consequently, this research aimed to lessen the negative impacts of SALIW on fennel plants by foliar spraying fennel plants with Ti forms (TiO_2 or TiO_2 NP) to help them become tolerant to SALIW; this research has never been done previously, particularly in Egypt.

Materials and methods

TiO_2 and TiO_2 NPs characterization

Morphological characterization of TiO_2 and TiO_2 NPs was carried out using TEM (Fig. 1A and B). TEM showed that TiO_2 shape was semi-round and platy with average particle size 1933 nm ($1.9\text{ }\mu\text{m}$). DLS of TiO_2 showed the size of particulate matter was about 1800 nm. The zeta potential of TiO_2 in aqueous solution was reprocessed

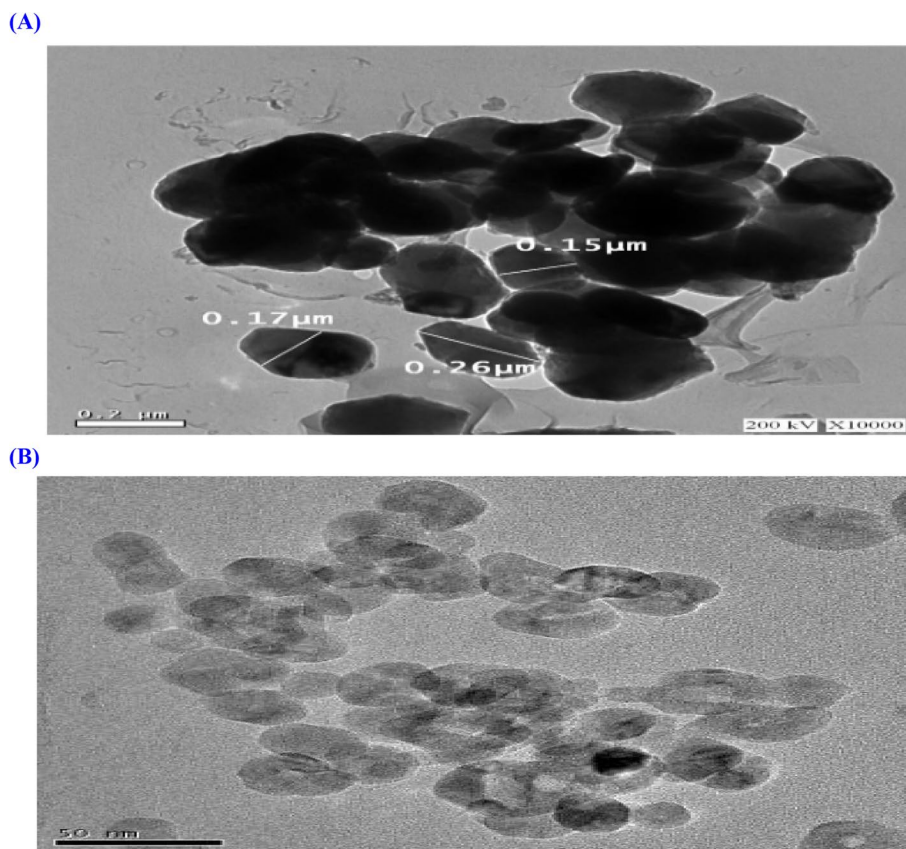


Fig. 1 **A** Transmission electron microscope (TEM) image of titanium dioxide (TiO₂). **B** TEM image of titanium dioxide nano particles (TiO₂NPs)

which was 16 mV. TEM presented that TiO₂ NPs were in definite particles with tetragonal shape at resolution scale 50 nm and approved by XRD. It is also clear from Fig. 1B that NPs are in dispersed as well as in aggregated form and the average size was 21 nm. DLS is used to determine the size of NPs which shows the size of particulate matter was about 19–20 nm. The zeta potential of TiO₂ NPs in aqueous solution was rescued which was 16.1 mV.

Experimental techniques

Two seasons (2021/2022 and 2022/2023) of experiments were conducted in a greenhouse at the National Research Centre in Giza, Egypt, with parameters set to 22/14 °C, 52/31% RH day/night and light intensity: 3700 lx. Fennel seeds were purchased from the Department of Medicinal and Aromatic Plants at Ministry of Agriculture in Giza, Egypt. In plastic pots (30 cm diameter and 50 cm height), seeds were planted in the second week of October during both seasons; there were 10 kg of air-dried soil in each pot. Six weeks after planting, seedlings were thinning to three plants per pot. Pots were separated into two main groups. The first one was subjected to various concentrations of TiO₂ forms at 0, 0.1, 0.2, 0.1NP, and 0.2NP

mM with fresh irrigation water (FRIW; 0.4 dS m⁻¹). The second group received the same TiO₂ treatments as the first, but it received 51.3 mM SALIW. Fennel plants were exposed to NaCl concentration after 45 days from planting (during vegetative stage); selection age was used according to Schreiber [29] and Day [30]. To make irrigation water highly soluble NaCl salt was employed. Salt concentration was measured by adding the proper quantity of NaCl to water and adjusting it using a portable EC meter; irrigation carried out according to field water capacity (FWC) which scored 60%. At every seven days, the soils found in pots of the second group were leached by tap water as a consequence of the salinity check; if there was no leaching when using SALIW, it can cause salt buildup in pots. TiO₂ (≥ 99%) and TiO₂NP (Nano powder 21 nm, particle size, TEAM, ≥ 99.5% trace metal basis, Sigma Aldrich. Sigma Aldrich Company supplied bulk anatase TiO₂ particles with 99% purity TiO₂NPs (<https://www.sigmaaldrich.com/EG/en/product/aldrich/637254>) with particle size 21 nm and purity more than 99%. The structural properties of TiO₂ and TiO₂ NPs were determined by Transmission Electron Microscope (TEM) model type of (JEOL-JEM-2100). TEM was

used for morphological characterization. For dynamic light scattering (DLS) and zeta potential, aqueous solution of desired concentration of TiO₂ and TiO₂NPs (1 g L⁻¹) was prepared by dissolving TiO₂ and TiO₂NPs in double de-ionized water (DDW). The solution was sonicated in a bath sonicator (~40 W) for 30 min. The particle size of the prepared TiO₂ and TiO₂NPs and zeta potential in an aqueous suspension were determined by measuring DLS with a Zetasizer (Malvern, UK). Due to selection of anatase type of bulk and nano titanium samples, X-Ray Diffraction (XRD) was not employed, since certified products were used with known particle shape. XRD measurement revealed that anatase level of the bulk TiO₂ particles are crystalline in nature and TiO₂ NPs were tetragonal particles [26–28]. TiO₂ and TiO₂NP were treated twice as foliar spray with equal amounts (2000 mL) to run-off to foliage. The first application was applied after 60 days of sowing and the second was after 15 days from the first application (during vegetative stage). TiO₂ and TiO₂NP were prepared in 1L volumetric flask by dissolve 0.05 or 0.1 g pure TiO₂ or TiO₂NP in distilled water to bring the total solution volume to 1L. The recommendations made by Egypt's Ministry of Agriculture were followed in all agricultural operations. According to Jackson [31] and Cottenie [32], Table 1 displays the physical and chemical characteristics of the soil employed in this investigation. Prior studies performed on fennel seedlings were used to assess the amounts of SALIW and TiO₂.

Harvesting

Fresh herbs (g plant⁻¹) were determined from each treatment during vegetative and flowering stages. Plant height (cm plant⁻¹) and herb dry weights (g plant⁻¹) were recorded during vegetative, flowering and fruiting stages. Fruit dry weights (g plant⁻¹) were measured during the ripening stage (240 days from the sowing).

Isolation of essential oil (EO)

From dried fruits in each treatment, three replicates of each treatment were made. Fifty grams of dried fruits from each replicate were hydro-distilled for three hours using Clevenger-style equipment [33]. Initial study suggests that hydro-distillation process should remain until no more EO could be extracted. For hydro-distillation, divided samples and 1L of water were introduced to a 2L round bottomed flask. EO boiling temperature was set to 100 °C for the purpose of extraction. EO was collected, cleaned of any traces of water using anhydrous sodium sulphate, and preserved at 4 °C in a sealed tube until use. Yield (g 100 plant⁻¹) and relative percentage (w/w) of EO content were determined.

Table 1 Specifications of the experimental soil

Items	values
Sand (%)	29.7
Silt (%)	17.6
Clay (%)	52.7
pH (1: 2.5)	7.7
EC (dS m ⁻¹)	1.8
Organic matter (%)	1.7
CaCO ₃ (%)	2.4
Total N (%)	27.8
P (mg 100 g ⁻¹ Soil)	17.3
Soluble Cations (mg 100 g ⁻¹ Soil)	
K	22.2
Fe	26.7
Mn	11.9
Zn	5.8
Cu	16.3
Ca	54.1
Mg	9.6
Na	44.6
Soluble Anions (mg 100 g ⁻¹ Soil)	
HCO ₃	68.8
Cl	42.8
CO ₃	27.9
SO ₄	33.8
NO ₃	17.9

Investigation of EO

Using a Shimadzu gas chromatography/mass spectrometric device (GC–MS), EO was investigated. Utilizing calibration curves generated using gas chromatography studies of common components; quantification was performed using an external standard approach that was developed. Retention index (RI), standard materials, and mass spectral data from the NIST/NBS and Wiley 275.1 libraries were used to identify the individual components of EO [34].

Analysis of POTSP

Using the techniques given by Anonymous [35], chlorophyll *a*, *b* and total carotenoids were quantified in fresh leaves {during vegetative stage (80 days from the sowing) and flowering stages (120 days from the sowing)} from each treatment. Fresh leaf tissues were pulverized in a mortar and pestle with 80% acetone. The optical density of the solution was measured using a spectrophotometer (Shimadzu UV-1700, Tokyo, Japan) at 662, 645 and 470 nm for Ch *a*, Ch *b* and carotenoids, respectively. Values for POTSP were given in mg g⁻¹ fresh weight.

Extraction and assessment of AOE

The technique for extracting AOE through vegetative and flowering stages was disclosed by Mukherjee [36]. The technique of Kar [37] was used to measure catalase activity (CAT), EC 1.11.1.6. Superoxide dismutase activity (SOD) EC 1.15.1.1 was identified by measuring inhibition of auto-oxidation of pyrogallol with the method of Marklund [38]. Peroxidase activity (POX) EC 1.11.1.7 assayed with the method of Kar [37] with slight modifications.

TSOLS determination

TSOLS content of dried leaves collected from each treatment during vegetative and flowering stages was determined using the Dubois [39]. To create the extract, a foliar tissue (0.03 g) was homogenized with 80% ethanol and a standard D-glucose solution was used to quantify using absorbance measurements at 490 nm.

Evaluation of TPHEN

According to Singleton [40], TPHEN content of leaf samples obtained at the vegetative and blooming phases was determined using the Folin-Ciocalteu reagent. In a nutshell, distilled water was used to dilute 2 mL of crude extract (1 mg mL^{-1}) to 3 mL, 0.5 mL of Folin-Ciocalteu reagent must be well mixed with the sample for three minutes, and 2 mL of 20% (w/v) sodium carbonate were added after that. The mixture was permitted to stand for a further hour in complete darkness, as well as measuring absorbance at 650 nm. The TPHEN content was calculated using the calibration curve, and the findings were shown as mg of gallic acid equivalent per gram of dry weight.

Estimation of TFLAV

According to Pourmorad [41], the aluminum chloride colorimetric technique was used to quantify the TFLAV in leaves taken throughout the vegetative and flowering periods. In a nutshell, 50 L of crude extract (1 mg mL^{-1} ethanol) were diluted with methanol to get 1 mL, 4 mL of distilled water was combined with 0.3 mL of a 5% NaNO_2 solution; after incubating for five minutes, 0.3 mL of 10% AlCl_3 solution was added, and the mixture was let to stand for six minutes. 2 mL of a 1 mol L^{-1} NaOH solution was then added, Double-distilled water was used to dilute the mixture to its final amount of 10 mL. The mixture was let to stand for 15 min, then, it was detected at 510 nm. TFLAV content was calculated using a calibration curve; it was then given as mg of rutin equivalent per g of dry weight.

Determining the PROL

Using the technique described by Bates [42], PROL content of fresh leaves taken throughout the vegetative and blooming periods was measured. PROL extract, acid ninhydrin, and glacial acetic acid were all added, and they were then incubated for one hour in a boiling water bath and then an ice bath. Using a Spekol Spectrocolorimeter VEB Carl Zeiss, the absorbance was determined at 520 nm. Authentic PROL was used at a known concentration to create a standard curve.

Measuring H_2O_2 and lipid peroxidation

Malondialdehyde (MDA) content was tested in order to determine the amount of lipid peroxidation using Heath's method [43]. A fresh leaf sample (collected at vegetative and flowering stages) weighing 0.5 g was homogenised in 10 ml of 5% tri-chloroacetic acid (TCA). Centrifuging the homogenate at $15,000 \times g$ for 10 min. 2.0 ml of the supernatant was mixed with 4.0 ml of 0.5% thiobarbituric acid (TBA) in 20% TCA. The mixture was heated for 30 min at 95°C , then immediately cooled in an ice bath and centrifuged for 10 min at $10,000 \times g$. using a JASCO V-750 spectrophotometer; the absorbance was measured at 500 nm and at 600 nm for non-specific absorption. A $155 \text{ mmol L}^{-1} \text{ cm}^{-1}$ extinction coefficient was used to compute the MDA concentration furthermore presented as $\text{nmol (MDA) g}^{-1} \text{ FW}$.

The technique of Yu [44] was used to measure the H_2O_2 in the leaf samples (taken at vegetative and blooming phases). In a mortar that had already been cooled, 0.5 g of fresh leaf tissue and 5.0 ml of 0.1% (w/v) TCA were combined. After 15 min of centrifugation at $12,000 g$, 0.5 ml of the supernatant was added to 1.0 ml of potassium iodide (KI) and 0.5 ml of potassium phosphate buffer (pH 7). Using a JASCO V-750 spectrophotometer, the absorbance was determined at 390 nm. Using the extinction value of $0.28 \text{ m}^{-1} \text{ cm}^{-1}$, the quantity of H_2O_2 was determined, and measured in $\text{nmol g}^{-1} \text{ FW}$.

Data evaluation

In this experiment, two variables were used: irrigation water sources (IRWS) such as FRIW & SALIW, and TiO_2 (5 rates). There were three replicates with ten pots for each one. The experiment employed a block design that was entirely random. According to Snedecor [45], the average data from both seasons were statistically analyzed using two ways analysis of variance. Variations between means were assessed by least of significant differences (LSD) at 0.05. Pearson's correlation was used to identify the relationships between growth characters and chemical variables during various growth stages. Also Pearson's correlation was used to identify the relationships EO

Table 2 Effect of IRWS, Ti forms and their interactions on the MORC and EO content

IRWS	Ti forms (mM)	Plant height (cm)			Weight of herb (g plant ⁻¹)					Fruit yield	EO	
					Fresh		Dry				g 100 plant ⁻¹	%
		.cm			g plant ⁻¹							
		VS	FLS	FRS	VS	FLS	VS	FLS	FRS			
FRIW	0.0	53.0	60.0	84.3	52.8	88.9	21.1	31.7	16.8	6.7	6.0	0.9
	0.1	53.3	60.7	85.3	62.4	96.2	24.6	37.8	18.7	7.8	8.6	1.1
	0.2	59.0	62.0	86.0	68.8	99.7	27.7	41.1	21.5	8.6	11.2	1.3
	0.1NP	61.0	68.0	101.3	77.6	112.6	31.8	45.8	26.6	16.8	28.6	1.7
	0.2NP	57.0	63.3	94.0	64.4	103.7	29.6	34.8	19.6	10.7	15.5	1.4
Overall FRIW		56.7	62.8	90.2	65.2	100.2	27.0	38.2	20.6	10.1	14.0	1.3
SALIW	0.0	35.6	43.0	52.0	36.3	54.9	12.9	23.4	11.2	4.9	5.4	1.1
	0.1	37.3	54.7	76.0	41.7	66.8	14.1	27.6	13.6	5.3	6.9	1.3
	0.2	38.3	56.0	77.0	48.9	79.7	17.1	31.8	16.5	6.4	9.6	1.5
	0.1NP	40.3	67.0	92.7	58.6	82.6	22.4	36.2	18.9	7.1	13.5	1.9
	0.2NP	39.6	61.0	82.5	50.3	77.3	19.6	30.7	15.6	6.1	9.8	1.6
Overall SALIW		38.2	56.3	70.0	47.2	72.3	17.2	29.9	15.2	6.0	9.0	1.5
Overall Ti forms	0.0	44.3	51.5	68.2	44.6	71.9	17.0	27.6	14.0	5.8	5.7	1.0
	0.1	45.3	57.7	80.7	52.1	81.5	19.4	32.7	16.2	6.6	7.8	1.2
	0.2	48.7	59.0	81.5	58.9	89.7	22.4	36.5	19.0	7.5	10.4	1.4
	0.1NP	50.7	67.5	97.0	68.1	97.6	27.1	41.0	22.8	12.0	21.1	1.8
	0.2NP	48.3	62.2	88.3	57.4	90.5	24.6	32.8	17.6	8.4	12.7	1.5
LSD (0.05)												
IRWS		1.1	2.1	2.5	2.1	1.2	0.9	1.2	3.3	0.9	2.3	ns
Ti forms		3.4	3.2	3.9	2.5	2.3	1.1	1.6	5.1	1.1	3.1	0.1
IRWS x Ti forms		4.2	4.1	5.1	2.9	2.7	1.3	2.1	7.2	1.2	5.4	ns

content and EO compositions. The interactions between IRW and TiO₂ as well as the individual (mean or overall) impacts of each factor were investigated in the results. The STAT-ITCF software claims that this approach was used in such ways [46].

Results

Ti forms, IRWS, and their interactions have an impact on the morphological characters (MORC)

Effects of TiO₂ or TiO₂NP, IRWS and their interactions on MORC such as plant height, fresh and dry mass production (g plant⁻¹) and fruit yield (g plant⁻¹) during different growth stages are shown in Table 2 and Figs. 2A-C and 3A). Changes in the rates of Ti forms and IRWS had a significant impact on MORC. Addition of SALIW resulted in a significant decrease in all MORC. Under varying amounts of Ti forms with IRWS, various MORC significantly enhanced. The maximum values of plant height (101.3 cm at fruiting stage) fresh mass (112.2 g plant⁻¹ at flowering stage), dry mass (26.6 g plant⁻¹ at flowering stage) and ripening fruit yield (16.8 g plant⁻¹) were recorded when plants received 0.1 mM TiO₂NP with FRIW.

Ti form, IRWS and their interactions' effects on EO composition

Significant reductions in EO output (g 100 plants⁻¹) were seen in response to SALIW compared with FRIW application (Table 2; Fig. 3B); however, it is evident that adding both Ti forms resulted in significant increases of EO yield. The greatest yield of EO (28.6 g 100 plants⁻¹) was recorded with the treatment of 0.1 mM TiO₂NP x FRIW (Table 2). According to Table (2), various increments in EO content have been observed under different IRWS, Ti forms and their interactions. Plants that received 0.1 mM TiO₂NP x SALIW had the highest amount of EO content (1.9%). The increments of EO contents were significant for Ti forms, but they were non significant for IRWS and Ti forms x IRWS.

EO from fennel fruits contained 23 components under IRWS, Ti forms and their interactions (Tables 3 and 4; Fig. 3C). Estragole, limonene, carvacrol, and carvone were the main ingredients. Different recognized components were present in three chemical fractions; Monoterpene hydrocarbons (MH) and sesquiterpene hydrocarbons (SH) were the minor fractions, whereas oxygenated monoterpenes (OM) made

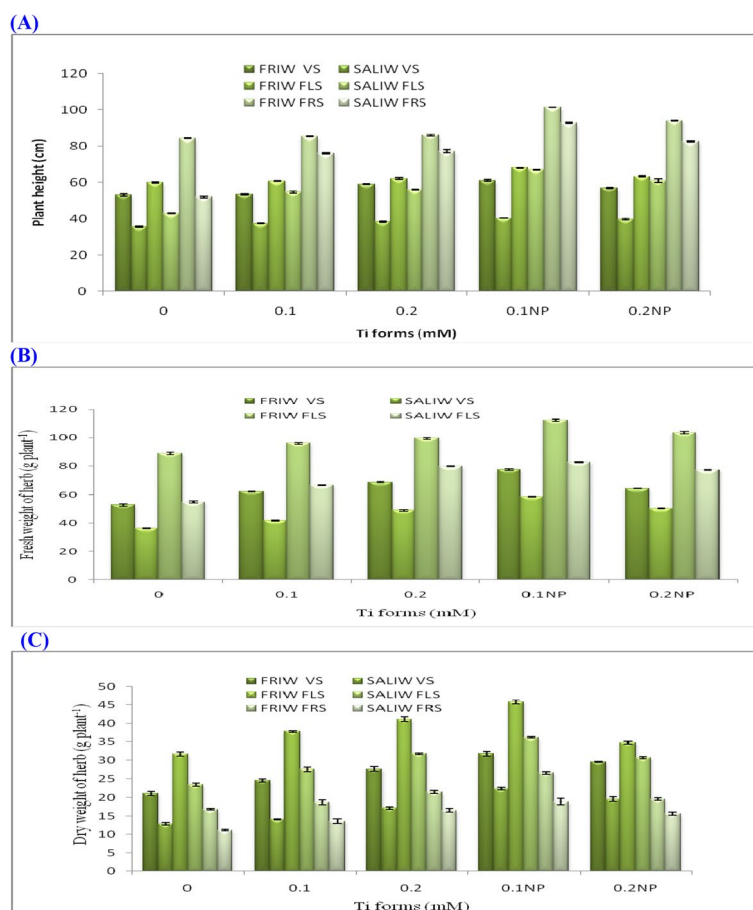


Fig. 2 **A** Effect of Ti forms and different irrigation water sources (IRWS) on plant height. **B** Impact of Ti forms and IRWS on fresh weight mass production. **C** Impact of Ti forms and IRWS on dry weight mass production. FLS, flowering stage; FRIW, fresh irrigation water; FRS, fruiting stage; SALIW, salted irrigation water; VS, vegetative stage. Every value is shown as mean ± SD (standard deviation)

up the majority. Significant variations were observed in the major components and all chemical groups as a result of IRWS, Ti forms, and their interactions. *Effect of IRWS x Ti forms* (Table 3): plants exposed to 0.2 mM TiO₂NP x SALIW produced the maximum amounts of estragole (51.9%), limonene (12.9%), carvacrol (12.9%), carvone (12.9%) and OM (81.5%). The greatest value of OM (19.7%) was obtained from plants subjected to 0.1 mM TiO₂NP x FRIW; while, plants treated with 0.2 mM TiO₂ x FRIW gave the maximum value of SH (4.7%). *Effect of Ti forms* (Table 4): treatment of 0.2 mM TiO₂NP gave the highest value of estragole (50.9%), limonene (12.8%), carvacrol (12.3%), carvone (12.0%) and OM (81.1%). The level of 0.1 mM TiO₂NP0 resulted in the maximum value of MH (18.7%), while, untreated plants gave the highest amount of SH (3.0%). *Effect of IRWS*: plants exposed to SALIW produced the highest amounts of estragole (50.7%), limonene (12.1%), carvacrol (11.6%), carvone (11.8%) and OM (80.3%); while those subjected to FRIW resulted in the highest values

of MH (17.8%) and SH (3.1%). The statistical changes in various components and their groups in response to IRWS, Ti forms and their interactions were presented in Tables 3 and 4.

Effects of Ti forms, IRWS and their interactions on the POTSP

During vegetative and blooming stages, SALIW led to a deficiency of POTSP (chlorophyll *a*, *b*, and total carotenoids); however, they were enhanced by applying both types of TiO₂ at different development phases (Table 5; Fig. 4A–C). The maximum values of chlorophyll *a* (19.3 mg g⁻¹) chlorophyll *b* (5.8 mg g⁻¹) and total carotenoids (7.8 mg g⁻¹) were obtained from the plant exposed to 0.1 mM TiO₂NP x FRIW during the flowering stage. The variations in chlorophyll *a* were significant for IRWS, Ti forms and IRWS x Ti forms. The changes in chlorophyll *b* and total carotenoids were highly significant for Ti forms; while they were not significant for IRWS or IRWS x Ti forms (Table 5).

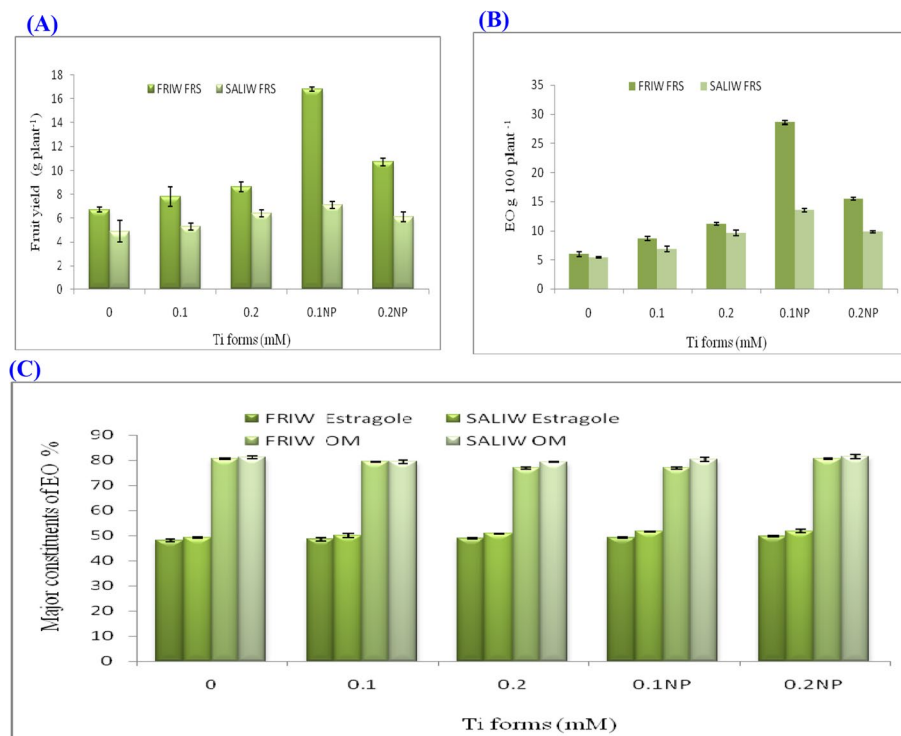


Fig. 3 **A** Effect of Ti forms and irrigation water sources (IRWS) on fruit yield. **B** Ti forms and IRWS's effect on essential oil (EO) yield. **C** Ti forms and IRWS changed the EO's estragole and oxygenated monoterpenes (OM). FRIW, fresh irrigation water; FRS, fruiting stage; SALIW, salted irrigation water. All values are given as mean \pm SD (standard deviation)

Effects of Ti forms, IRWS, and their interactions on the AOE A

AOEA (POX, CAT and SOD) in plant cells were significantly increased during vegetative and flowering stages when plants were exposed to various IRWS and Ti forms (Table 5; Fig. 5A-C). Plants exposed to SALIW x Ti forms displayed greater levels of AOE A than those exposed to FRIW. The maximum values of POX (12.1 unit/g FW. min), CAT (12.1 unit/g FW. min) and SOD (4.6 unit/g FW. min) were resulted from plant treated with SALIW x 0.1 mM TiO₂NP at flowering stage. The increments of POX, CAT and SOD were significant for IRWS, Ti forms and the interactions (Table 5).

Ti forms, IRWS, and their interactions' effects on the TSOLS

The contents of TSOLS significantly increased with various IRWS, TiO₂ from and TiO₂ from x IRWS at vegetative and flowering stages (Table 6; Fig. 6A). Plants treated with Ti forms x SALIW gave higher values in TSOLS than those exposed to Ti forms x FRIW. However, the highest value of TSOLS (33.2 mg g⁻¹) resulted from 0.2 mM TiO₂NP x SALIW during the flowering stage.

Effects of Ti forms, IRWS, and their interactions on TPHEN

Through both vegetative and flowering phases, application of various IRWS, Ti forms, and their interactions promoted

the assembly of TPHEN in fennel plants (Table 6; Fig. 6B). Plants treated with Ti forms x SALIW produced more TPHEN than plants subjected to FRIW. The highest amount of TPHEN (4.8 mg g⁻¹) was found in SALIW with 0.1 mM TiO₂NP treatment. At vegetative stage, the increases in TPHEN were non significant for IRWS or Ti forms treatments, but they were significant for the interaction treatments (Table 6); on the other hand, they were significant for all treatments during flowering stage.

Ti form, IRWS, and their interactions' effects on the TFLAV

Changes in the IRWS resulted in a significant increment of the TFLAV, either Ti forms or IRW x Ti forms produced several changes in the TFLAV contents during vegetative and flowering stages (Table 6; Fig. 6C). Plants treated with 0 mM TiO₂ x SALIW at flowering stage produced the greatest amount of TFLAV with the value of 3.1 mg g⁻¹.

PROL rates and the effects of Ti forms, IRWS and their interactions

The accumulations of PROL were significantly promoted by applying various IRWS, Ti forms and IRWS x Ti forms during vegetative and flowering stages (Table 6; Fig. 7A). Plants exposed to SALIW x Ti forms gave higher values in PROL concentrations than those subjected to FRIW

Table 3 IRWS x Ti forms' impact on the constituents of EO

No	Constituents (%)	RI	IRWS	SALIW										LSD (0.05)		
				FRIW					SALIW							
				Ti forms (mM)												
0.0	0.1	0.2	0.1NP	0.2NP	0.0	0.1	0.2	0.1NP	0.2NP	0.0	0.1	0.2	0.1NP	0.2NP		
1	α-Pinene	930	1.5	0.5	0.8	0.9	0.6	0.7	0.7	0.5	0.6	0.6	0.7	0.5	0.6	0.3
2	Camphene	953	0.5	0.8	0.8	0.6	0.6	0.4	0.5	0.4	0.7	0.4	0.5	0.4	0.4	0.2
3	β-Pinene	980	0.7	0.9	0.7	0.9	0.5	0.5	1.1	0.8	0.2	0.3	0.5	0.2	0.3	0.5
4	Myrcene	991	0.4	1.4	1.6	1.4	0.4	0.6	0.5	0.7	0.7	0.7	0.5	0.7	0.7	0.4
5	α-Phellandrene	1005	0.2	1.4	0.9	0.5	0.4	0.3	1.1	0.6	0.7	0.8	0.6	0.7	0.6	0.6
6	p-cymene	1026	0.6	1.2	0.4	2.4	1.3	0.4	0.3	0.7	0.6	0.6	0.6	0.7	0.6	0.5
7	Limonene	1031	10.9	11.1	11.5	11.8	11.9	11.7	11.9	12.3	12.8	12.9	12.9	12.3	12.9	ns
8	.trans-β-Ocimene	1050	0.3	0.4	0.5	0.4	0.8	0.5	0.5	0.5	0.5	0.4	0.5	0.5	0.4	0.2
9	γ-Terpinene	1062	0.2	1.1	1.1	0.8	0.5	1.3	1.6	0.5	0.4	0.7	1.6	0.5	0.4	0.7
10	Fenchone	1094	2.5	1.6	0.7	0.8	0.5	1.5	0.8	0.6	0.5	0.6	1.5	0.6	0.5	0.8
11	Terpinen-4-ol	1177	0.7	1.7	0.9	0.3	0.4	0.6	0.5	0.9	0.5	0.7	0.6	0.9	0.5	0.5
12	α-Terpineol	1189	0.8	1.4	0.9	2.2	2.7	1.9	1.1	0.5	0.8	0.5	1.1	0.5	0.8	0.6
13	Estragole	1195	48.1	48.5	48.9	49.2	49.8	49.2	50.1	50.8	51.7	51.9	49.2	50.8	51.7	0.9
14	.cis-Carveol	1229	1.8	2.1	1.4	0.9	0.7	2.2	1.6	1.2	0.5	0.3	2.2	1.2	0.5	0.3
15	Pulegone	1237	2.9	2.8	1.6	0.7	0.5	1.1	1.5	0.9	0.6	0.4	1.1	0.9	0.6	0.7
16	Carvone	1242	9.7	9.9	10.4	10.5	11.1	10.5	11.4	11.9	12.5	12.9	10.5	11.9	12.5	0.8
17	Anethole	1255	1.5	0.8	0.8	0.5	0.9	1.7	0.7	0.3	0.4	0.8	1.7	0.3	0.4	0.9
18	Thymol	1290	2.8	0.5	0.5	0.8	2.5	2.2	0.8	0.7	0.6	0.5	2.2	0.7	0.6	0.7
19	Carvacrol	1298	9.9	10.1	10.7	10.9	11.6	10.3	10.9	11.5	12.2	12.9	10.3	11.5	12.2	0.9
20	Caryophyllene	1428	1.1	0.4	1.7	0.7	0.4	0.9	0.5	0.6	0.7	0.2	0.9	0.6	0.7	0.4
21	β-Humulene	1440	1.1	0.7	1.6	0.5	0.3	0.4	0.4	1.6	0.6	0.6	0.4	1.6	0.6	0.4
22	Germacrene D	1490	0.9	0.3	0.9	0.7	0.5	0.5	0.7	0.7	0.6	0.2	0.5	0.7	0.6	0.2
23	β-Bisabolene	1509	0.7	0.2	0.5	1.3	0.8	0.3	0.6	0.7	0.5	0.5	0.3	0.7	0.5	0.4
	MH (1-9)		15.3	18.8	18.3	19.7	17.0	16.4	18.2	17.0	17.2	16.8	16.4	18.2	17.0	1.1
	OM(10-19)		80.7	79.4	76.8	76.8	80.7	81.2	79.4	79.3	80.3	81.5	81.2	79.4	79.3	1.2
	SH (20-23)		3.8	1.6	4.7	3.2	2.0	2.1	2.2	3.6	2.4	1.5	2.1	3.6	2.4	1.3
	Overall		99.8	99.8	99.8	99.7	99.7	99.7	99.8	99.9	99.9	99.8	99.7	99.9	99.9	99.8

Table 4 The effect of Ti forms or IRWS on the components of EO

No	Constituents (%)	RI	IRWS		Ti forms (mM)					LSD	
			FRIW	SALIW	LSD	0.0	0.1	0.2	0.1NP		0.2NP
1	α-Pinene	930	0.9	0.7	0.1	1.1	0.6	0.7	0.8	0.8	0.2
2	Camphene	953	0.7	0.5	0.1	0.5	0.7	0.6	0.7	0.5	0.1
3	β-Pinene	980	0.7	0.6	0.2	0.6	1.0	0.8	0.6	0.4	0.4
4	Myrcene	991	1.0	0.6	0.3	0.5	1.0	1.2	1.1	0.6	0.3
5	α-Phellandrene	1005	0.7	0.7	ns	0.3	1.3	0.8	0.6	0.6	0.4
6	p-cymene	1026	1.2	0.5	0.3	0.5	0.8	0.6	1.5	1.0	0.6
7	Limonene	1031	11.4	12.1	0.3	11.3	11.5	11.9	12.3	12.8	0.3
8	.trans-β-Ocimene	1050	0.5	0.5	ns	0.4	0.5	0.5	0.5	0.6	ns
9	γ-Terpinene	1062	0.7	0.9	0.1	0.8	1.4	0.8	0.6	0.6	0.3
10	Fenchone	1094	1.2	0.8	0.2	2.0	1.2	0.7	0.7	0.6	0.2
11	Terpinen-4-ol	1177	0.8	0.6	0.1	0.7	1.1	0.9	0.4	0.6	0.5
12	α-Terpineol	1189	1.6	1.0	0.2	1.4	1.3	0.7	1.5	1.6	0.6
13	Estragole	1195	48.9	50.7	0.9	48.7	49.3	49.9	50.5	50.9	0.6
14	.cis-Carveol	1229	1.4	1.2	0.1	2.0	1.9	1.3	0.7	0.5	0.4
15	Pulegone	1237	1.7	0.9	0.3	2.0	2.2	1.3	0.7	0.5	0.6
16	Carvone	1242	10.3	11.8	0.8	10.1	10.7	11.2	11.5	12.0	0.8
17	Anethole	1255	0.9	0.8	ns	1.6	0.8	0.6	0.6	0.9	0.3
18	Thymol	1290	1.4	1.0	0.2	2.5	0.7	0.6	0.6	1.5	0.6
19	Carvacrol	1298	10.6	11.6	0.6	10.1	10.5	11.1	11.6	12.3	0.9
20	Caryophyllene	1428	0.9	0.6	0.2	1.0	0.5	1.2	0.7	0.3	0.2
21	β-Humulene	1440	0.8	0.7	ns	0.8	0.6	1.5	0.6	0.5	0.7
22	Germacrene D	1490	0.7	0.5	0.1	0.7	0.5	0.8	0.7	0.4	0.1
23	β-Bisabolene	1509	0.7	0.5	0.1	0.5	0.4	0.6	0.9	0.7	0.2
MH (1–9)			17.8	17.1	0.2	15.9	18.5	17.7	18.7	16.9	0.9
OM (10–19)			78.9	80.3	0.6	81.0	79.4	78.1	78.6	81.1	0.7
SH (20–23)			3.1	2.4	0.7	3.0	1.9	4.1	2.8	1.8	0.6
Overall			99.8	99.8		99.9	99.8	99.9	99.9	99.8	

Table 5 POTSP and AOEA are impacted by IRWS x Ti forms

IRWS	Ti forms (mM)		POTSP (mg g ⁻¹)		AOEA (unit/g FW. min)											
			Chlorophyll a		Chlorophyll b		Carotenoids		POX		CAT		SOD			
	VS	FLS	VS	FLS	VS	FLS	VS	FLS	VS	FLS	VS	FLS	VS	FLS		
FRW	0.0	12.0	2.3	2.6	3.3	4.3	3.4	5.6	1.7	8.3	1.4	2.8				
	0.1	10.1	2.8	3.5	4.0	5.0	3.7	5.8	2.4	9.1	1.5	2.9				
	0.2	10.2	3.0	3.8	4.1	5.0	4.0	6.4	3.3	9.8	1.7	3.0				
	0.1NP	14.0	19.3	4.5	5.8	5.7	7.8	4.1	6.6	4.7	10.5	1.9	3.1			
	0.2NP	12.1	12.3	3.3	3.7	5.1	5.9	4.6	6.7	4.1	10.2	2.0	3.2			
	10.8	13.8	3.2	3.9	4.4	5.6	4.0	6.2	3.3	9.6	1.7	3.0				
SALIW	0.0	7.6	1.8	2.4	2.7	3.7	4.7	7.4	5.5	10.8	2.1	3.3				
	0.1	7.5	2.1	3.4	3.2	3.8	4.8	8.0	5.9	11.2	2.1	3.5				
	0.2	11.0	12.7	3.3	4.2	4.6	4.9	8.5	6.7	11.6	2.3	3.7				
	0.1NP	14.9	16.3	4.1	4.5	5.8	6.1	5.2	7.9	13.6	2.6	4.0				
	0.2NP	10.9	15.6	3.6	4.2	4.6	5.8	5.3	7.2	12.1	2.4	4.6				
	10.2	12.3	3.0	3.7	4.2	4.9	5.0	9.3	6.6	11.8	2.3	3.8				
Overall Ti forms	0.0	7.0	2.1	2.5	3.0	4.0	4.1	6.5	3.6	9.5	1.7	3.1				
	0.1	8.8	2.5	3.5	3.6	4.4	4.3	6.9	4.2	10.2	1.8	3.2				
	0.2	10.6	12.8	3.2	4.0	4.4	5.0	4.6	7.4	10.7	2.0	3.4				
	0.1NP	14.5	17.8	4.3	5.2	5.8	7.0	4.7	8.5	12.0	2.3	3.6				
	0.2NP	11.5	14.0	3.5	4.0	4.9	5.9	5.0	9.4	11.1	2.2	3.9				
	10.2	12.3	3.0	3.7	4.2	4.9	5.0	9.3	6.6	11.8	2.3	3.8				
LSD (0.05)																
IRWS	0.2	0.1	ns	ns	ns	ns	0.1	0.1	0.1	0.4	0.1	0.2				
Ti forms	0.4	0.5	0.1	0.1	0.1	0.2	0.1	0.1	0.4	0.7	0.1	0.4				
IRWS x Ti forms	0.5	0.8	ns	ns	ns	ns	0.2	0.3	0.5	1.1	0.2	0.7				

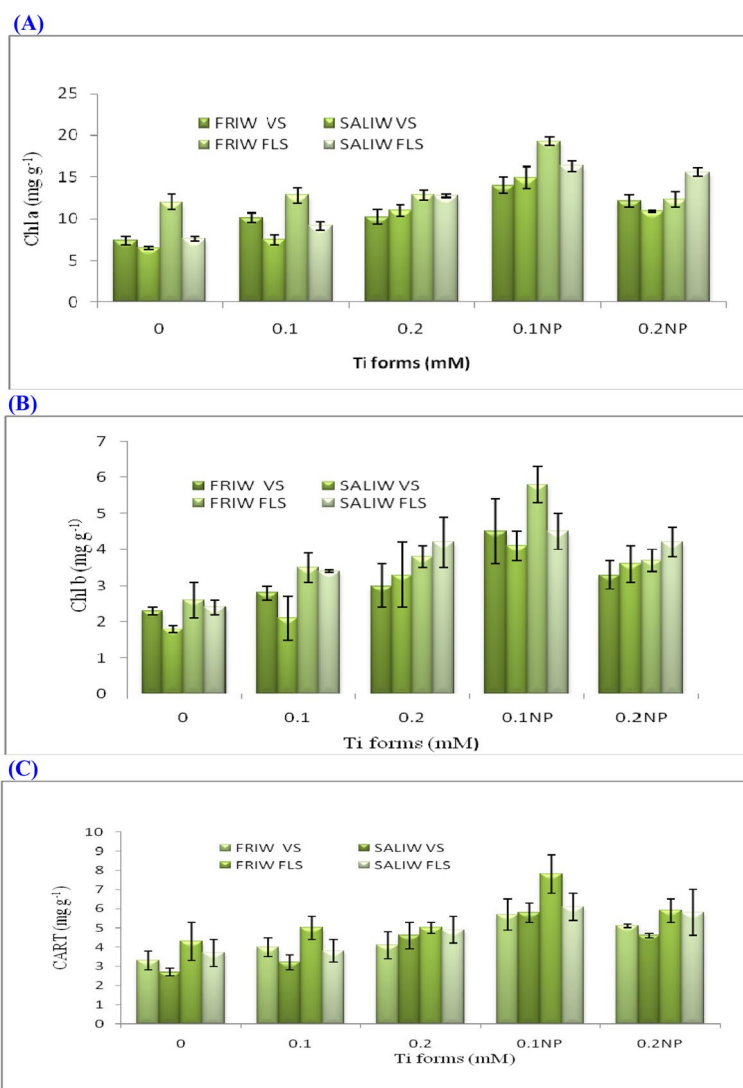


Fig. 4 **A** Effect of Ti forms and irrigation water sources (IRWS) on chlorophyll *a* (Chl *a*). **B** Ti forms and IRWS's impact on Chl *b*. **C** Impact of IRWS and Ti forms on carotenoids (CART). FLS, flowering stage; FRIW, fresh irrigation water; SALIW, salted irrigation water; VS, vegetative stage. Each value is displayed as mean ± SD (standard deviation)

x Ti forms during both stages. The maximum value of PROL ($19.1 \mu\text{moles g}^{-1}$) was recorded with plants treated with SALIW \times 0.1 mM TiO_2NP at flowering stage.

Effects of Ti forms, IRWS, and their interactions on the H_2O_2 levels

As SALIW, significant increments in H_2O_2 accumulations were observed at vegetative and flowering stages compared with FRIW application; however, by using Ti forms, they were reduced (Table 6; Fig. 7B). Plants exposed to SALIW without Ti forms throughout the flowering stage had the highest H_2O_2 level ($15.5 \mu\text{M}$).

Effects of Ti forms, IRWS, and their interactions on the MDA levels

Throughout both vegetative and flowering phases, application of various IRWS, Ti forms and their interactions promoted the assembly of MDA in fennel leaves (Table 6; Fig. 7C). Compared to plants exposed to IRWS in both phases, those exposed to IRWS x Ti forms produced lower concentrations of MDA. The peak MDA rate was discovered in SALIW without Ti forms (25 nmol g^{-1}) during the blooming stage. The variations in MDA rates for IRWS, Ti forms and their interactions were significant (Table 6).

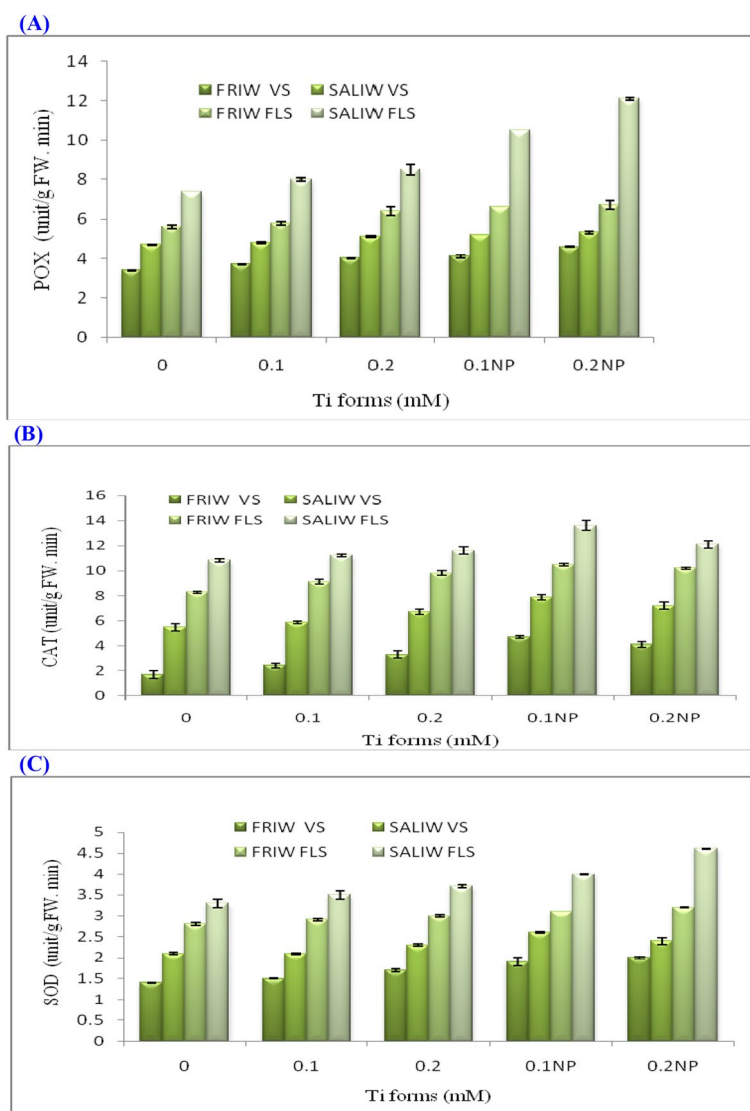


Fig. 5 **A** Effect of Ti forms and irrigation water sources (IRWS) on peroxidase (POX). **B** Impact of IRWS and Ti forms on catalase (CAT). **C** Response of superoxide dismutase (SOD) to IRWS and Ti forms. FLS, flowering stage; FRIW, fresh irrigation water; SALIW, salted irrigation water; VS, vegetative stage. Every value is shown as mean ± SD (standard deviation)

Pearson’s correlation

Pearson’s correlation was applied to analyze the relationship between MORC and chemical component in VS, FLS and FRS. Pearson’s correlation results exhibited positive and negative correlations between different variables. During VS and FLS (Fig. 8A and B), MORC variables and chemical component gave a strong correlation; highly strong correlation was found between PLH and FW or DW; FW and POTSP; AOEA and TPHEN. While low correlations were recorded between MDA, H₂O₂, PROL and AOEA. Urging FRS, PLH was strongly correlated with DW, fruit production and EO %; while low correlated with EO yield (Fig. 9A). Different correlations were

noticed between EO constituents (Fig. 9B). Estragole had highly strong positive correlation with carvone and carvacrol; but strong negative correlation with *cis*-carveol and pulegone. Low correlation was obtained between limonene, *trans*-β-ocimene with β-sabolenene.

Discussion

This research found that fennel plants exposed to SALIW experienced substantial reductions in plant height, fresh and dry weights and fruit yield; it might be as a result of SALIW, a source of toxic ions that raise the osmotic pressure of soil solution [47]; therefore, plant roots are unable to extract water from the surrounding soil, and plant

Table 6 IRWS x Ti forms' effects on the concentrations of TSOLS, TPHEN, TFLAV, PROL, H₂O₂, and MDA

IRWS	Ti forms (Mm)	TSOLS		TPHEN		TFLAV		PROL (μmoles g ⁻¹)		H ₂ O ₂ (μM)		MDA (nmol g ⁻¹)	
		(mg g ⁻¹)											
		VS	FLS	VS	FLS	VS	FLS	VS	FLS	VS	FLS	VS	FLS
FRIW	0.0	12.7	14.6	3.9	4.1	1.4	2.1	1.2	2.2	10.4	11.8	6.3	22.1
	0.1	14.4	17.5	4.0	4.3	1.8	2.6	3.8	4.5	5.3	8.2	5.9	15.7
	0.2	15.4	17.8	4.0	4.5	1.9	2.7	4.3	5.0	5.0	8.1	3.8	14.5
	0.1NP	18.6	20.4	4.1	4.6	1.5	2.2	1.7	2.3	4.5	6.5	1.4	9.2
	0.2NP	20.6	21.0	4.2	4.4	1.7	2.4	3.2	3.4	2.9	3.5	4.4	15.6
Overall FRIW		16.3	18.3	4.0	4.4	1.7	2.4	2.8	3.5	5.6	7.6	4.4	15.4
SALIW	0.0	13.9	16.3	4.1	4.3	2.5	2.8	5.2	6.5	14.6	15.5	9.6	25.0
	0.1	16.7	17.8	4.2	4.4	2.4	2.9	6.5	11.5	9.5	9.9	6.4	15.6
	0.2	17.3	25.1	4.3	4.7	2.3	3.1	6.6	13.8	8.9	9.6	4.4	15.5
	0.1NP	19.1	28.2	4.6	4.8	1.9	3.0	6.7	15.7	5.2	7.8	1.5	12.5
	0.2NP	31.3	33.2	4.4	4.6	2.1	2.0	6.3	19.1	5.5	7.3	2.6	13.4
Overall SALIW		19.7	24.1	4.3	4.6	2.2	2.8	6.3	13.3	8.7	10.2	4.9	16.4
Overall Ti forms	0.0	13.3	15.5	4.0	4.2	2.0	2.5	3.2	4.4	12.5	13.7	8.0	23.6
	0.1	15.6	17.7	4.1	4.4	2.1	2.8	5.2	8.0	7.4	9.1	6.2	15.7
	0.2	16.4	21.5	4.2	4.6	2.1	2.9	5.5	9.4	7.0	8.9	4.1	15.0
	0.1NP	18.9	24.3	4.4	4.7	1.7	2.6	4.2	9.0	4.9	7.2	1.5	10.9
	0.2NP	26.0	27.1	4.3	4.5	1.9	2.2	4.8	11.3	4.2	5.4	3.5	14.5
LSD (0.05)													
IRWS		0.2	0.3	ns	0.1	0.1	0.2	0.1	0.9	1.1	1.9	0.2	0.9
Ti forms		0.7	0.7	ns	0.1	0.2	0.4	0.3	1.3	2.6	3.1	0.7	1.2
IRWS x Ti forms		1.9	1.1	0.6	0.2	0.3	0.8	0.9	1.7	4.4	4.2	1.1	1.7

exposure to drought causes a drop in turgor, which causes a decrease in plant cell development, which is followed by growth inhibition [48]. Fennel, which is known to be susceptible to abiotic stress factors [21], may be made to tolerate SALIW stress better by applying Ti forms to its leaves. Prior research has demonstrated beneficial effects of TiO₂ form treatments (as foliar a spray) on the balance and absorption of critical macro and micronutrients; this causes chlorophyll and protein levels to rise, resulting in an increase of growth, dry matter content and yield under SALIW conditions [23]. However, the negative impact of SALIW on fennel's growth characteristics was overcome by the application of TiO₂NP; possible causes include: plants treated with TiO₂NP showed improved activities of AOE and enhanced accumulation of osmolytes. According to research, AOE are plants' first line of defense against ROS, and enhanced AOE can change how well plants tolerate stress [49]. Furthermore, osmotic stress caused by SALIW was balanced by an increase in osmolytes, which provided the plant cells with improved osmotic pressure, allowing them to absorb more water as seen by an increase in leaf relative water content [50].

According to recent research, SALIW treatments reduce the production of POTSP; that since chloroplasts are

degrading; this results in a lower buildup of carotenoids, chlorophyll *a*, and *b* [51]. However, plants exposed to Ti forms x SALIW accumulated more POTSP than plants exposed to SALIW. The potential for valence shift makes the TiO₂ form unique among the transition forms; as a result, it takes part in activities involving the transfer of electrons related to photosynthesis [52]. Due to the development of a Ti-ascorbate compound in recent years, which can be applied as a foliar spray to plants, the usage of TiO₂ in various crops has been reported in certain research studies to have significantly boosted photosynthesis [52]. The rates of POTSP were accelerated by foliar application of TiO₂NP x SALIW [53]; this could be as a result of TiO₂NP's ability to increase light absorption, speed up the transport and conversion of light energy, shield chloroplasts from ageing, and extend the duration during which they are able to perform photosynthetic activity [54].

In response to the applications of SALIW and Ti forms, several modifications were seen in EOs and their components. Numerous alterations to EOs and their constituents occur when fennel plants are exposed to SALIW or SALIW x Ti forms; these modifications can be attributed to differences in the enzymatic activity of EO formations [10]. EO is thought to increase an aromatic plant's

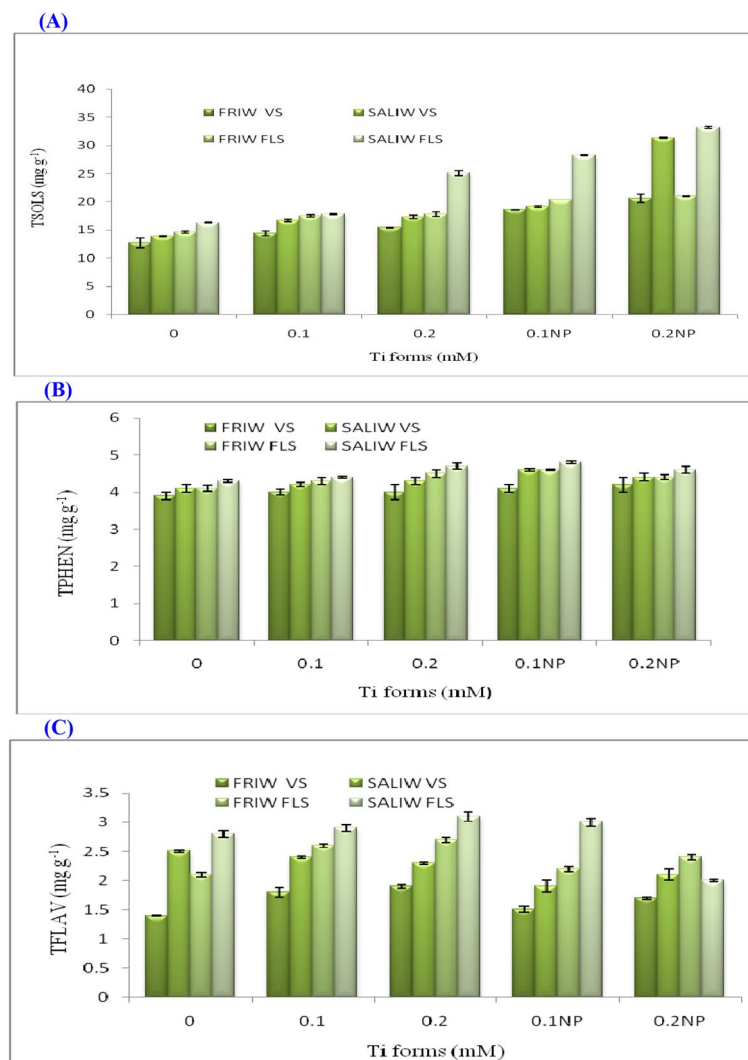


Fig. 6 **A** Impact of Ti forms and irrigation water sources (IRWS) on total soluble sugars (TSOLS). **B** Ti forms and IRWS effect on total phenols (TPHEN). **C** Total flavonoids (TFLAV) are affected by Ti forms and IRWS. FLS, flowering stage; FRIW, fresh irrigation water; SALIW, salted irrigation water; VS, vegetative stage. All values are shown as mean \pm SD (standard deviation)

tolerance to unfavorable circumstances like SALIW [10]. The elevation in EO (%) in response to SALIW with Ti forms may be explained by an increase in the number of glandular hairs as well as a corresponding increase in their densities [55]; while the differences in plant dry matter when exposed to SALIW or SALIW x Ti forms can be linked to the changes in EO yield [55, 56].

Treatments with SALIW resulted in an increase in the buildup of TSOLS; the storage of TSOLS for long-term energy supply, prolonged metabolism, and improved plant recovery may be the cause of the rise in TSOLS under SALIW [57]. In addition, SALIW x Ti forms treated plants showed greater accumulation TSOLS than SALIW-treated plants. Higher photosynthetic activity

was closely correlated with a greater buildup of TSOLS in leaves in plants treated with SALIW x Ti forms [58]. Lack of study has made it difficult to currently understand the relationship between SALIW and the accumulation of antioxidant molecules.

Plants increase their TPHEN and TFLAV production in response to SALIW, which, following their role as antioxidants in preventing oxidative damage, provide a twofold protective impact, as the health-improving substances found in food plants [59]. Because TPHEN and TFLAV have the ability to free radical scavenging (FRRS) that kills plant cells, increasing the antioxidant capabilities are correlated with antioxidants substances levels (polyphenols and TFLAV) [60, 61]. Plants treated with

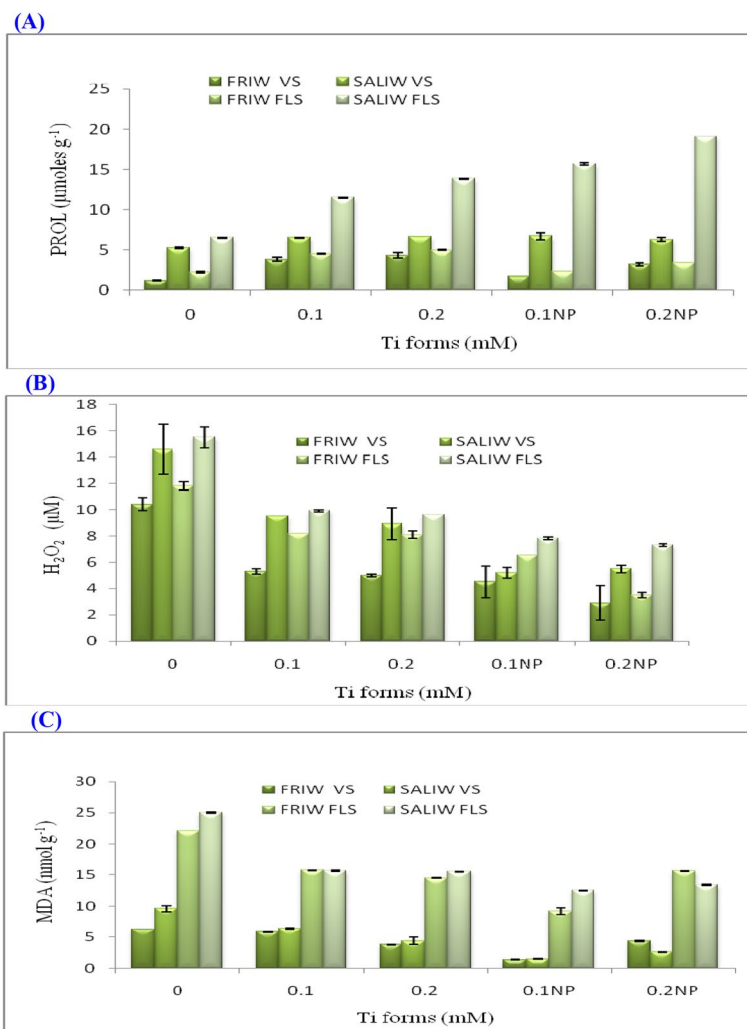


Fig. 7 **A** Impact of irrigation water sources (IRWS) and Ti forms on proline (PROL). **B** Ti forms and IRWS's effect on hydrogen peroxide (H₂O₂). **C** Ti forms & IRWS and their impact on malondialdehyde (MDA). FLS, flowering stage; FRIW, fresh irrigation water; SALIW, salted irrigation water; VS, vegetative stage. All values are displayed as mean ± SD (standard deviation)

SALIW x Ti forms gave higher amounts of TPHEN and TFLAV than those exposed to SALIW; these results confirmed by Ghorbanpour [62].

SALIW advocated for the accumulation of PROL in fennel leaves. These findings are consistent with the findings of Blum [63], as well as Slama [57] who revealed that PROL is regarded a source of energy, carbon, and nitrogen for regenerating plant tissues under SALIW. PROL content of fennel plants treated with Ti forms as a foliar spray x SALIW was higher than when treated with SALIW, these findings are confirmed by Lima [64].

The rise in H₂O₂ content in response to SALIW might be attributed to oxidative stress induced by SALIW, as well as peroxidation of membrane lipids, as represented by the higher concentration of H₂O₂; on the contrary,

foliar spray Ti forms to salt-stressed plants lowered the amount of oxidative stress and lipid peroxidation, as evidenced by a decrease in H₂O₂ concentration [65].

SALIW causes an increase in MDA content in plant tissues as a lipid peroxidation product, which shows the oxidative damage induced by SALIW, leading to membrane damage; an increase in MDA level causes membrane damage and subsequent leaking of membrane electrolytes [66]. Mustafa [67] validated these findings that Ti forms inhibit MDA generation.

There are some earlier research publications on various plants that corroborate our conclusions on the usage of Ti forms in increasing plant development and yield under SALIW. TiO₂ may act as an elicitor to improve secondary metabolism in sage plants for biosynthesis of natural

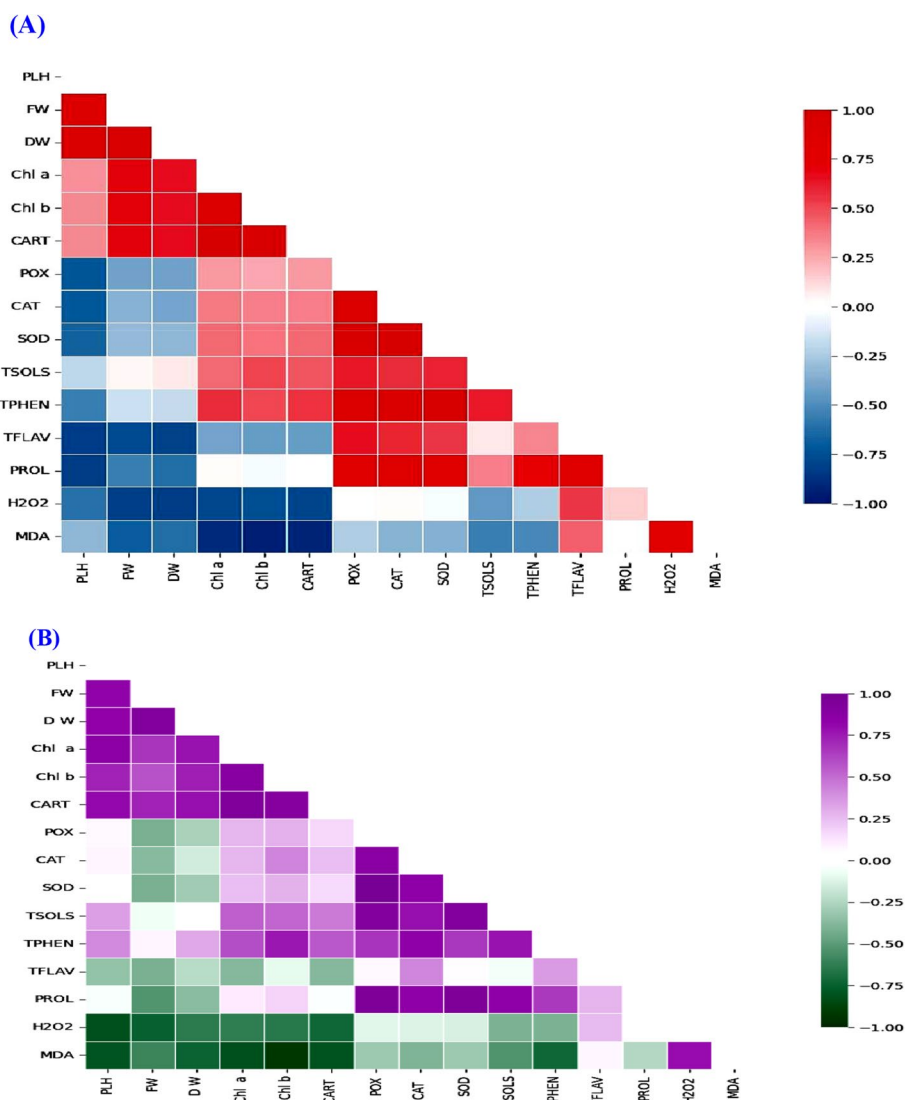


Fig. 8 **A** Pearson's correlation between morphological characters (MORC) and chemical contents variables in response to Ti forms and irrigation water sources (IRWS) during vegetative stage (VS). **B** Pearson's correlation between MORC and chemical contents variables in response to Ti forms and IRWS during flowering stage (FLS). CART, carotenoids; CAT, catalase; Chl, chlorophyll; DW, dry weight; FW, fresh weight; H₂O₂, hydrogen peroxide; MDA, malondialdehyde; PLH, plant height; POX, peroxidase; PROL, proline; SOD, superoxide dismutase; TFLAV, total flavonoids; TPHEN, total phenols; TSOLS, total soluble sugars

antioxidants (TPHEN and TFLAV), EO and its main constituents (camphene, *p*-cymene, 1, 8-cineol, *cis*-thujene and camphor) [62]. It was discovered that using TiO₂NP as a spray under abiotic stress factors had a significant influence on thyme plant growth characteristics and EO compositions [55]. TiO₂NP improved tomato plant growth, yield, and quality under SALIW by increasing the rates of carbonic anhydrase, nitrate reductase, AOE, PROL, and Glycine Betaine [65]. TiO₂ x SALIW treatments increased tomato yield parameters; while they reduced pH value, TA, EC, TSS and TSS/TA ratio in the fruits [27]. TiO₂ or TiO₂NP applied via leaves at low concentrations has

been documented to improve crop performance through stimulating the activity of certain enzymes, enhancing chlorophyll content and photosynthesis, promoting nutrient uptake, strengthening stress tolerance and improving crop yield and quality [25]. TiO₂NP application on Moldavian balm cultivated under SALIW enhanced agronomic characteristics, EO yield, geraniol, citral, geranyl acetate, geraniol, and AOE; however, it decreased H₂O₂ production [28]. TiO₂NP increased wheat germination properties, yield features, osmotic, water potential, carotenoids, TPHEN, TFLAV, TSOLS, proteins, PROL, amino acid contents and AOE; whereas it lowered MDA at various

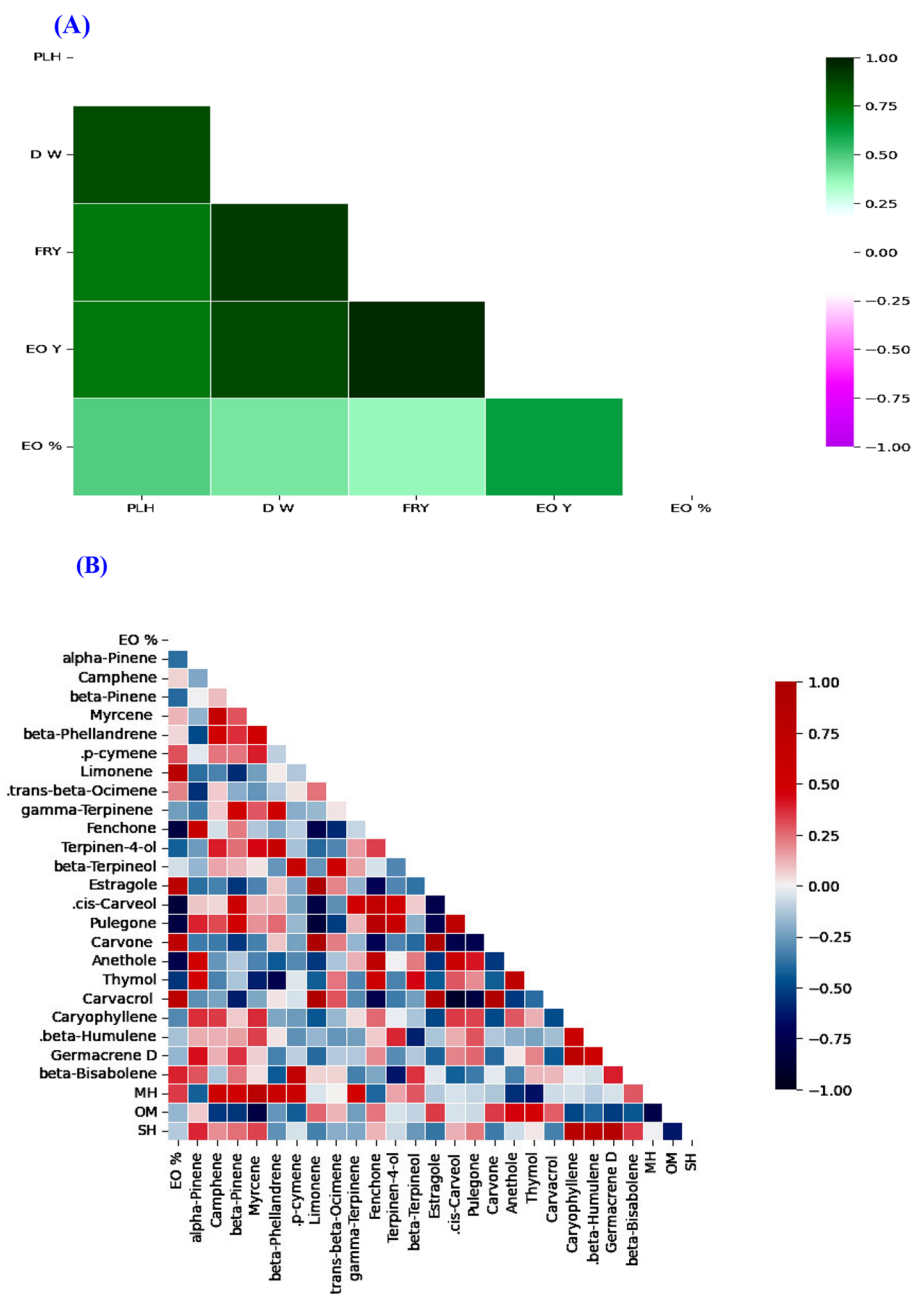


Fig. 9 **A** Pearson's correlation between morphological characters (MORC) and essential oil (EO) variables in response to Ti forms and irrigation water sources (IRWS) during fruiting stage (FRS). **B** Pearson's correlation between EO (%) and EO constituent's variables in response to Ti forms and IRWS. DW, dry weight; EO, essential oil; EOY, essential oil yield; FRY, fruit yield; MH, monoterpene hydrocarbons; OM, oxygenated monoterpenes; PLH, plant height; SH, sesquiterpene hydrocarbons

SALIW doses; as a result, foliar application of TiO₂NP can aid in plant resistance to SALIW [67]. TiO₂ have a positive effect on the yield, macro & micronutrients and antioxidant activity of tomato plants under SALIW [68]. Several farmers would benefit from this experiment since they will be able to produce fennel plant in Egypt's new territories,

which would reduce the negative impacts of SALIW and boost the chances of exporting fennel to other countries. Furthermore, it has been stated that the creation of fennel EO would allegedly require Ti application, as they greatly affect its synthesis, resulting in the extension of its biological domain as a natural source of EO.

Conclusion

TiO₂ forms increased production of fennel plants exposed to SALIW. Ti responded to SALIW by increasing POTSP, PROL, carbohydrates, EO, and antioxidants while lowering H₂O₂ and MDA. Application of TiO₂NP is proposed as a viable strategy to lessen the detrimental effects of salinized irrigation water on fennel plant.

Abbreviations

AlCl ₃	Aluminum chloride
AOEA	Antioxidant enzymes activity
Ca	Calcium
CAT	Catalase
Chl	Chlorophyll
CART	Carotenoids
DW	Dry weight
DLS	Dynamic light scattering
dS m	Deci siemens per metre
EC	Electrical conductivity
EO	Essential oil
Fig	Figure
FLS	Flowering stage
FRIW	Fresh irrigation water
FRRS	Free radical scavenging
FRS	Fruiting stage
FW	Fresh weight
FWC	Field water capacity
GC/MS	Gas chromatography/mass spectrometric
H ₂ O ₂	Hydrogen peroxide
IRWS	Irrigation water sources
K	Potassium
L	Liter
LSD	Least of significant differences
min	Minute
MDA	Malondialdehyde
Mg	Magnesium
MH	Monoterpene hydrocarbons
mM	milli Molar
MORC	Morphological characters
Na	Sodium
NaCl	Sodium chloride
NaNO ₂	Sodium nitrite
NP	Nano particles
ns	non significant
OM	Oxygenated monoterpenes
PLH	Plant height
POTSP	Photosynthetic pigments
POX	Peroxidase activity
PROL	Proline
PROT	Proteins
RI	Retention index
ROS	Reactive oxygen species
SALIW	Saltened irrigation water
SD	Standard deviation
SH	Sesquiterpene hydrocarbons
SOD	Superoxide dismutase
TA	Titrateable acidity
TEM	Transmission electron microscope
TCARB	Total carbohydrates
TFLAV	Total flavonoids
Ti	Titanium
TiO ₂	Titanium dioxide
TiO ₂ NP	Titanium dioxide nano particles
TPHEN	Total phenols
TSOLS	Total soluble sugars
TSS	Total soluble solids
VS	Vegetative stage
XRD	X-ray diffraction

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Authors' contributions

The trials in the greenhouse were conducted by AMAA and FSZ, who also assessed the chemical contents and carried out the statistical analysis. KAK determined the components of the EO and wrote the paper. The final version was read and approved by the authors.

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Availability of data and materials

The corresponding author can provide the datasets used and/or analyzed during the current work upon proper request.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

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Competing interests

The authors declare no competing interests.

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References

- Kanel KR. Sustainable management of medicinal and aromatic plants in Nepal: a strategy. New Delhi: A study commissioned by IDRC/SARO, Medicinal and Aromatic Plants Program in Asia (HMAPP); 2000.
- Dhifi W, Bellili S, Jazi S, Bahloul N, Mnif W. Essential oils' chemical characterization and investigation of some biological activities: a critical review. *Med*. 2016;3:1–16.
- Bakkali F, Averbeck S, Averbeck D, Idaomar M. Biological effects of essential oils: a review. *Food Chem Toxicol*. 2008;46:446–75.
- Omidbaigi R. Production and processing of medicinal plants. Tehran: Astane Ghods Razavi; 2007.
- Guillen MD, Manzanos MJ. A contribution to study Spanish wild grown fennel (*Foeniculum vulgare* Mill.) as a source of flavor compounds. *Chemie, Mikrobiologie Technologie der Lebensmittel*. 1994;16:141–5.
- Huang TM, Pang ZH. The role of deuterium excess in determining the water salinisation mechanism: a case study of the arid Tarim River Basin, NW China. *Appl Geochem*. 2012;27:2382–8.
- Chartzoulakis KS. Salinity and olive: growth, salt tolerance, photosynthesis and yield. *Agric Water Manage*. 2005;78:108–21.
- Verma AK, Gupta SK, Isaacs RK. Use of saline water for irrigation in monsoon climate and deep water table regions: simulation modeling with SWAP. *Agric Water Manage*. 2012;115:186–93.
- Golezani GK, Abdoli S. Physiological and biochemical responses of medicinal plants to salt stress. In: Aftab T, editor. *Environmental challenges and medicinal plants. Environmental challenges and solutions*. Cham: Springer; 2022. ISBN 978-3-030-92050-0 (eBook).
- Burbott AJ, Loomis D. Evidence for metabolic turnover monoterpene in peppermint. *Plant Physiol*. 1969;44:173–9.
- Solinas V, Deiana S. Effect of water and nutritional conditions on the *Rosmarinu officinalis* L phenolic fraction and essential oil yields. *Riv Ital Eposs*. 1996;19:189–98.
- Parida AK, Das AB. Salt tolerance and salinity effects on plants. *Ecol Environ Saf*. 2005;60:324–49.

13. Khalid K, da Silva JT. Yield, essential oil and pigment content of *Calendula officinalis* L. flower heads cultivated under salt stress conditions. *Scien Hort*. 2010;126:297–305.
14. Khalid AK, Ahmed MA. Growth and certain biochemical components of black cumin cultivated under salinity stress factor. *J Mater Environ Sci*. 2017;8:7–13.
15. Ashraf M, Akhtar N. Influence of salt stress on growth, ion accumulation and seed oil content in sweet fennel. *Biol Plant*. 2004;48:461–4.
16. Semiz GD, Ünlükara A, Yurtseven E, Suarez DL, Telci İ. Salinity impact on yield, water use, mineral and essential oil content of fennel (*Foeniculum vulgare* Mill.). *J Agric Sci*. 2012;18:177–86.
17. Cucci G, Lacolla G, Boari F, Cantore V. Yield response of fennel (*Foeniculum vulgare* Mill.) to irrigation with saline water. *Acta Agric Scand - B Soil Plant Sci*. 2014;64:129–34.
18. Rebey IB. Variation in fatty acid and essential oil composition of sweet fennel (*Foeniculum vulgare* Mill) seeds as affected by salinity. *J New Sci*. 2016;6:1233–40.
19. Beykhhormizi A, Sarghein SH, Ardakani MRS, Moshtaghion SM, Kouhi SMM. Alleviation of salinity stress by vermicompost extract: a comparative study on five fennel landraces. *Commun Soil Sci Plant Anal*. 2018;49:2123–30.
20. Shafeiee M, Ehsanzadeh P. Physiological and biochemical mechanisms of salinity tolerance in several fennel genotypes: existence of clearly-expressed genotypic variations. *Indus Crop Prod*. 2019;132:311–8.
21. Sisakht JN, Ehsanzadeh P, Ehteman H. Fennel outperforms ajwain and anise in the saline environment: physiological response mechanisms in germinating seeds and mature plants. *Ital J Agron*. 2022;17:2096.
22. Lyu S, Wei X, Chen J, Wang C, Wang X, Pan D. Titanium as a beneficial element for crop production. *Front Plant Sci*. 2017;25(8):597.
23. Bacillieri FS, Vasconcelos AC, Quintao LRM, Mageste JG, Torres JLR. Titanium (Ti) in plant nutrition-A review. *Aust J Crop Sci*. 2017;11:382–6.
24. Wen J, Li X, Liu W, Fang Y, Xie J, Xu Y. Photocatalysis fundamentals and surface modification of TiO₂ nanomaterials. *Chin J Catal*. 2015;36:2049–70.
25. Chaudhary I, Singh V. Titanium dioxide nanoparticles and its impact on growth, biomass and yield of agricultural crops under environmental stress: a review. *Res J Nanosci Nanotechnol*. 2021;10:1–8.
26. Abdel Latef AAH, Srivastava AK, El-sadek MSA, Kordrostami M, Tran LSP. Titanium dioxide nanoparticles improve growth and enhance tolerance of broad bean plants under saline soil conditions. *Land Degrad Deve*. 2018;29(1065):1073.
27. Carbajal-Vazquez VH, Gomez-Merino FC, Herrera-Corredor JA, Contreras-Oliva A, Alcantar-Gonzalez G, Trejo-Tellez LI. Effect of titanium foliar applications on tomato fruits from plants grown under salt stress conditions. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*. 2020;48:924–37.
28. Gohari G, Mohammadi A, Akbari A, Panahirad S, Dadpour MR, Fotopoulos V, Kimura S. Titanium dioxide nanoparticles (TiO₂ NPs) promote growth and ameliorate salinity stress effects on essential oil profile and biochemical attributes of *Dracocephalum moldavica*. *Sci Rep*. 2020;10:912.
29. Schreiber HA, Stanbery CO. Barley production as influenced by timing of soil moisture and timing on Na application. *Agron J*. 1965;57:442–5.
30. Day AD, Thompson PK. Effect of soil moisture regimes on growth of barley. *Agron J*. 1975;67:430–3.
31. Jackson ML. Soil chemical analysis. 1st ed. New Delhi: Prentice Hall Ltd publishing; 1973.
32. Cottenie A, Verloo M, Kiekens L, Velgh G, Camerlynck R. Chemical analysis of plant and soils. Laboratory of analytical and agro chemistry. Belgium: State University of Gent publishing; 1982.
33. Clevenger JF. Apparatus for determination of essential oil. *J Amer Pharm Assoc*. 1928;17:346–9.
34. Adams RP. Identification of essential oil components by gas chromatography/mass spectrometry. Carol Stream: Allured Publ Corp; 1995.
35. Anonymous. Official methods of analysis. 20th ed. Washington, DC: Association of Official Analytical Chemists; 2016.
36. Mukherjee SP, Choudhuri MA. Implications of water stress-induced changes in the levels of endogenous ascorbic acid and hydrogen peroxide in *Vigna* seedlings. *Plant Physiol*. 1983;58:166–70.
37. Kar M, Mishra D. Catalase, peroxidase and polyphenoloxidase activities during rice leaf senescence. *Plant Physiol*. 1976;57:315–9.
38. Marklund S, Marklund G. Involvement of the superoxide anion radical in the autoxidation of pyrogallol and a convenient assay for superoxide dismutase. *Eur J Biochem*. 1974;47:469–74.
39. Dubois M, Gilles KA, Hamilton JK, Roberts PA, Smith F. Phenol sulphuric acid method for carbohydrate determination. *Ann Chem*. 1956;28:350–9.
40. Singleton VL, Orthofer R, Lamuela-Raventos RM. Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. *Methods Enzymol*. 1999;299:152–78.
41. Pourmorad F, Hosseinimehr S, Shahabimajid N. Antioxidant activity, phenol and flavonoid contents of some selected Iranian medicinal plants. *Afr J Biotechnol*. 2006;5:1142–5.
42. Bates LS, Waldren RP, Teare ID. Rapid determination of free proline of water stress studies. *Plant Soil*. 1973;39:205–7.
43. Heath RL, Packer L. Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. *Arch Biochem Biophys*. 1968;125:189–98.
44. Yu CW, Murphy TM, Lin CH. Hydrogen peroxide-induced chilling tolerance in mung beans mediated through ABA-independent glutathione accumulation. *Funct Plant Biol*. 2003;30:955–63.
45. Snedecor GW, Cochran WG. Statistical methods. 11th ed. Ames Iowa: Iowa State Univ Press; 1990.
46. Foucart T. Analyse factorielle, programmation sur micro-ordinateur. Masson ITCF Paris. 1982. ISBN-13: 978-2225764509.
47. Jabeen N, Ahmed R. Demonstration of growth improvement in sunflower (*Helianthus annuus* L.) by the use of organic fertilizers under saline conditions. *Pak J Bot*. 2009;41:1373–84.
48. Misra A, Srivastava NK. Influence of water stress on Japanese mint. *J Herb Spi Med Plants*. 2000;7:51–8.
49. Li T, Hu Y, Du X, Tang H, Shen C, Wu J. Salicylic acid alleviates the adverse effects of salt stress in *Torreya grandis* cv. Merrillii seedlings by activating photosynthesis and enhancing antioxidant systems. *PLoS One*. 2014;10:e109492.
50. Wutipraditkul N, Wongwean P, Buaboocha T. Alleviation of salt-induced oxidative stress in rice seedlings by proline and/or glycinebetaine. *Biol Plant*. 2015;59:547–53.
51. Aspinall D, Paleg LG. Proline accumulation. In: Paleg LG, Aspinall D, editors. Physiology and biochemistry of drought resistance in plants, physiology aspects. New York: Academic Press; 1981.
52. Alcaraz CF, Sánchez MF, Giménez JL. Ascobato de titanio fertilizante foliar. *Agri*. 1991;708:636–8.
53. Akbari GA, Morteza E, Moaveni P, Alahdadi I, Bihamta MR, Hasanloo T. Pigments apparatus and anthocyanins reactions of borage to irrigation, methylalcohol and titanium dioxide. *Int J Biosci*. 2014;4:192–208.
54. Yang F, Hong F, You W, Liu C, Gao F, Wu C, Yang P. Influence of nano-anatase TiO₂ on the nitrogen metabolism of growing spinach. *Biol Trace Elem Res*. 2006;110:179–90.
55. Nasab BF, Sirousmehr AR, Azad H. Effect of titanium dioxide nanoparticles on essential oil quantity and quality in *Thymus vulgaris* under water deficit. *J Med Plant By-prod*. 2018;2:125–33.
56. El-Sherif AF, Shehata SM, Youssif RM. Response of tomato seedlings to zinc application under different salinity levels. *Egypt J Hortic Sci*. 1990;17:131–42.
57. Slama I, Ghnaya T, Hessini K, Messedi D, Savouire A, Abdelly C. Comparative study of the effects of mannitol and PEG osmotic stress on growth and solute accumulation in *Sesuvium portulacastrum*. *Environ Exp Bot*. 2007;61:10–7.
58. Vatankhah A, Aliniaiefard S, Nezhad MM, Abdi S, Mokhtarpour Z, Reezi S, Tsaniklidis G, Fanourakis D. Plants exposed to titanium dioxide nanoparticles acquired contrasting photosynthetic and morphological strategies depending on the growing light intensity: a case study in radish. *Sci Rep*. 2023;13:5873.
59. Martinez V, Mestre TC, Rubio F, Girones-Vilaplana A, Moreno DA, Mittler R. Accumulation of flavonols over hydroxycinnamic acids favors oxidative damage protection under abiotic stress. *Front Plant Sci*. 2016;7:838.
60. Huang YC, Chang YH, Shao YY. Effects of genotype and treatment on the antioxidant activity of sweet potato in Taiwan. *Food Chem*. 2006;98:529–38.
61. Ben Taarit M, Msaada K, Hosni K, Marzouk B. Fatty acids, phenolic changes and antioxidant activity of clary sage (*Salvia sclarea* L.) rosette leaves grown under saline conditions. *Ind Crop Pro*. 2012;38:58–63.
62. Ghorbanpour M. Major essential oil constituents, total phenolics and flavonoids content and antioxidant activity of *Salvia officinalis* plant in response to nano-titanium dioxide. *Ind J Plant Physiol*. 2015;20:249–56.

63. Blum A, Ebercon A. Genotype responses in sorghum to drought stress. III. Free proline accumulation and drought resistance. *Crop Sci.* 1976;16:379–86.
64. Lima MGS. Efeito do estresse salino sobre a concentração de pigmentos e prolina em folhas de arroz. *Bragantia.* 2004;63:335–40.
65. Khan MN. Nano-titanium dioxide (Nano-TiO₂) mitigates NaCl stress by enhancing antioxidative enzymes and accumulation of compatible solutes in tomato (*Lycopersicon esculentum* Mill.). *J Plant Sci.* 2016;11:1–11.
66. Liu T, Hou GG, Cardin M, Marquart L, Dubat A. Quality attributes of whole wheat four tortillas with sprouted whole wheat four substitution. *LWT.* 2017;77:1–7.
67. Mustafa N, Raja NI, Ilyas N, Abasi F, Ahmad MS, Ehsan M, Mehak A, Badshah I, Pročków J. Exogenous application of green titanium dioxide nanoparticles (TiO₂ NPs) to improve the germination, physiochemical and yield parameters of wheat plants under salinity stress. *Molecules.* 2022;30:4884.
68. Carbajal-Vázquez VH, Gómez-Merino FC, Alcántar-González EG, Sánchez-García P, Trejo-Téllez LI. Titanium increases the antioxidant activity and macronutrient concentration in tomato seedlings exposed to salinity in hydroponics. *Plants.* 2022;11:1036.

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